General Synthesis Route to Benanomicin-Pradimicin Antibiotics

Minoru Tamiya,^[a] Ken Ohmori,^[a] Mitsuru Kitamura,^[a, b] Hirohisa Kato,^[a] Tadamasa Arai,^[a] Mami Oorui,^[a] and Keisuke Suzuki^{*[a]}

Dedicated to the memory of Professor Tsuguo Mizuochi

Abstract: A general approach to the regio- and stereoselective total synthesis of the benanomicin-pradimicin antibiotics (BpAs) is described. Construction of the aglycon has been achieved by 1) the diastereoselective ring-opening of a biaryl lactone by using (R)-valinol as a chiral nucleophile and 2) the stereocontrolled semi-pinacol cyclization of the aldehyde acetal by using SmI₂ in the presence of BF₃•OEt₂ and a proton source to afford the ABCD tetracyclic monoprotected diol. This strat-

Introduction

The benanomicin-pradimicin antibiotics (BpAs) were isolated from the culture of the *Actinomyces* species in 1988 by two independent groups.^[1] To date, more than 20 congeners have been identified (Figure 1) that share a unique structure, which consists of a benzo[*a*]naphthacene core with different amino acid and disaccharide moieties. Their potent antifungal and anti-HIV activities are attributed to Ca²⁺mediated specific binding to the mannose-rich oligosaccharides presented on fungi or virus surfaces, and therefore, they are called "lectin-mimics".^[2]

[a] Dr. M. Tamiya, Dr. K. Ohmori, Dr. M. Kitamura, Dr. H. Kato, T. Arai, M. Oorui, Prof. Dr. K. Suzuki Department of Chemistry, Tokyo Institute of Technology SORST-JST, 2-12-1 O-okayama, Meguro-ku Tokyo 152-8551 (Japan) Fax: (+81)3-5734-2788 E-mail: ksuzuki@chem.titech.ac.jp
[b] Dr. M. Kitamura Current Address: Department of Applied Chemistry

Kyushu Institute of Technology 1-1 Sensui, Tobata-ku, Kitakyushu, Fukuoka 804-8550 (Japan)

Supporting information for this article is available on the WWW under http://www.chemeurj.org/ or from the author.

egy enabled us to control the two stereogenic sites in the B ring (C-5 and C-6) and the regioselective introduction of the carbohydrate moiety. The ABCD tetracycle could serve as an ideal platform for the divergent access to various BpAs. The amino acid (Dalanine) was introduced onto the

Keywords: antibiotics • benanomicin • natural products • pradimicin • total synthesis ABCD tetracycle. Glycosylation was promoted by the combination of Cp_2HfCl_2 and AgOTf (1:2 ratio). Construction of the E ring followed by deprotection completed the first total synthesis of benanomicin A (2a), benanomicin B (2b), and pradimicin A (1a). The route is flexible enough to allow the synthesis of other congeners differing in their amino acid and carbohydrate moieties.

Because of these intriguing biological properties as well as their structural novelty, the synthetic community,^[3] which includes our group,^[4–7] has focused considerable attention on the BpAs. This paper will detail our efforts towards the synthesis of BpAs with particular emphasis on the new strat-

СОН

O _≫ NH										
MeO = HO =										
	R ¹	R ²		\mathbb{R}^1	R ²					
pradimicin A (1a)	N(Me)H	Me	pradimicin FA-1 (1e)	N(Me)H	CH ₂ OH					
pradimicin B (1b) ^[a]	N(Me)H	Me	pradimicin FA-2 (1f)	NH_2	$\mathrm{CH}_{2}\mathrm{OH}$					
pradimicin D (1e)	N(Me)H	Н	benanomicin A (2a)	OH	Me					
pradimicin E (1d)	NH_2	Н	benanomicin B (2b)	$\rm NH_2$	Me					
[a] Sugar moiety of 1b is shown below.			(= pradimicin C)							
HOLOT			BMY-28864 (2c) ^[b]	NMe ₂	CH_2OH					
			[b] Unnatural product.							
HC			•							

Figure 1. Structures of the benanomicin-pradimicin antibiotics.

© 2007 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim



egies and tactics developed during the course of this synthetic study.

In planning the total synthesis of the BpAs, three problems were considered: 1) The construction of the densely functionalized pentacyclic chromophore, 2) the control of the diol stereochemistry in the B ring, and 3) the regioselective introduction of the sugar moiety.

Results and Discussion

Strategic considerations

Pseudo-symmetry problem: The major challenge in this whole synthetic venture was the regio- and stereochemical control of the vicinal diol at C-5 and C-6 with pseudo-symmetry. To highlight the essence of the problem, Scheme 1 shows a hypothetical route based on the asymmetric epoxidation of benzo[a]naphthacene core A (Step 1) followed by hydrolytic epoxide cleavage (Step 2). Even if Step 1 occurred with high enantiofacial selectivity, Step 2 must occur regioselectively to obtain the enantiopure diol. This would not be straightforward because the two ends of the epoxide have similar reactivities, both are benzylic and there is little difference in the sterics in the vicinity of the epoxide moiety. In addition, even if this pseudo- C_s -symmetry problem were somehow managed, one must confront the pseudo- C_2 symmetry of the resulting diol, which would raise the problem of regioselective installation of the carbohydrate moiety (Step 3) that would once again not be a trivial issue.

Stereochemical clues from model studies: With these issues in mind, we became interested in another possible route to the stereogenic diol motif, that is, the pinacol cyclization of a biaryl dialdehyde (Scheme 2). The problems of diastereoand enantiocontrol need to be addressed for this particular approach to be valid.



Scheme 2. Pinacol cyclization strategy.

To our delight, initial attempts to solve the diastereocontrol problem immediately gave positive results.^[5] Reaction of biaryl dialdehyde $\mathbf{3}^{[8]}$ with SmI₂^[9] cleanly afforded *trans*diol $\mathbf{4}^{[10]}$ as the sole product [Eq. (1)]. Consistently high yields and *trans* selectivity were attained in the intramolecular pinacol coupling reaction of various biaryl dialdehydes by using various reducing agents (cf. Sm, V, Ti).^[5] In particular, SmI₂ gave excellent *trans* selectivity.



The clue to enanticocontrol was obtained when pinacol cyclization was applied to chiral, nonracemic biaryl dialdehyde **5**. For example, reaction of (M)-**5** with SmI₂ gave a quantitative yield of the *trans*-diol (S,S)-**6** [Eq. (2)]. Thus, the axial stereochemistry in **5** was completely transmitted into the stereogenicities of *trans*-diol **6**.^[11]





Scheme 1. Hypothetical route to BpAs based on a three-step protocol.

9792 -

www.chemeurj.org

© 2007 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

Additional information: During the structure determination of diol **6**, we discovered an interesting fact concerning its axial stereochemistry. Diacetate **7**, which was derived from diol **6**, has a low barrier (ca. 15 kcalmol⁻¹) for conformational flipping, as estimated by variable-temperature NMR spectroscopy [Eq. (3)].^[5] The ratio of **7**_{diequatorial} and **7**_{diaxial} at -40 °C was about 1:1.



This observation was in line with the pioneering work of Mislow and Harvey who reported that the flipping barrier of tethered biaryl **B** ($\Delta G^{\pm} = 24.1 \text{ kcal mol}^{-1}$) is much lower than that of the nontethered counterpart **C** ($\Delta H^{\pm} = 50.8 \text{ kcal mol}^{-1}$, calculated; Figure 2).^[12a] In general, a two-atom bridge renders the biaryl configurationally labile, allowing the interconversion of **B** and *ent*-**B**, in sharp contrast to the behavior of nontethered biaryl **C**.



Figure 2. Properties of biaryl compounds.

Such properties of tethered biaryls (configurationally labile) and tetra-*ortho*-substituted biaryls (configurationally inert) inspired us to envision a three-step strategy for a stereocontrolled access to the key tetrol structure **H** embedded in the target molecules (Figure 3): 1) Biaryl lactone **E**, even if it is sterically encumbered, would be available by taking advantage of intramolecular C–C bond formation in the ester precursor **D** (Step 1),^[13] 2) chiral biaryl dialdehyde **F** would be obtained by the Bringmann-type asymmetric cleavage of the configurationally labile lactone **E**^[14] by employing a chiral, nonracemic reagent (Step 2), and 3) dihydrophenanthrenetetrol **G** could be constructed by stereospecific pinacol cyclization of enantioenriched biaryl dialdehyde **F** (Step 3).

This strategy takes advantage of the stereochemical lability of lactone \mathbf{E} to drive the enantioselective ring-opening completely to a single enantiomer of \mathbf{F} , which subsequently undergoes stereospecific pinacol coupling to *trans*-diol \mathbf{G} .



FULL PAPER

Figure 3. Three-step strategy for the construction of the tetrol substructure.

Although the axial stereochemistry, set inertly during the formation of **F** from **E**, becomes labile again in **G**, the stereochemical interplay between the stereochemistry of the diol and the biaryl axis results in the preservation of stereochemical information.^[10a, 12]

Retrosynthetic analysis: Scheme 3 shows our retrosynthetic analysis of the BpAs, which starts with the fragmentation of the target into three moieties, a sugar, an amino acid, and aglycon **I**. By assuming that the B ring would be constructed by the pinacol cyclization reaction, the enantiopure biaryl dialdehyde **II** was set as the precursor, which could in principle be available by the asymmetric ring-opening reaction of lactone **III** pioneered by Bringmann and co-workers.^[14] To cope with the sterically encumbered biaryl bond formation between the anthracene and the A ring segment, we envisioned an intramolecular biaryl C–C bond formation (cf. Figure 3).^[13] Ester **IV** could be further dissected into anthracene **VI** and the orsellinic acid derivative **V**. Anthracene **VI** would be constructed by the Diels–Alder reaction of naph-thoquinone **VIII** with diene **VII**.^[15]

First-generation total synthesis

Initial approach to the aglycon: Based on these analyses, Aring segment **14** was synthesized from the known orsellinic acid derivative **8** (Scheme 4).^[16] Phenol **8** was converted into triflate **9** by selective triflation of the phenol *para* to a methoxycarbonyl group followed by protection of the remaining phenol as a methoxymethyl (MOM) ether. Carbonyla-





Scheme 4. Preparation of the A-ring segment (unless otherwise noted, the reactions were performed at ambient temperatures). Reagents and conditions: a) PhNTf₂, K₂CO₃, acetone, 24 h, 70%; b) MOMCl, *i*Pr₂NEt, CH₂Cl₂, 5.5 h, quant; c) Pd(OAc)₂, DPPF, CO, PhOH, DMF, $60 \rightarrow 80$ °C, 3 h, quant; d) LiBH₄, THF, 24 h, 98%; e) MOMCl, *i*Pr₂NEt, CH₂Cl₂, 3 h, 88%; f) CF₃CO₂H, CH₂Cl₂, 0°C, 3 h, 99%; g) BnMe₃N+ICl₂⁻, NaHCO₃, MeOH, CH₂Cl₂, 0°C, 10 h, 83%. Tf=trifluoromethanesulfonyl, DPPF= diphenylphosphinoferrocene.

tion of triflate **9** in the presence of phenol afforded phenyl ester **10**.^[17] Selective reduction of the phenyl ester in **10** was cleanly achieved with LiBH₄ without affecting the methyl ester moiety to give alcohol **11**, which was protected as bis-MOM ether **12**. Acid treatment of **12** gave selective removal of the MOM protecting group of the phenol to give **13** and its iodination by using BnMe₃N⁺ICl₂⁻ gave iodophenol **14**.^[18]

Synthesis of the CDE-ring segment started with commercially available aldehyde **15** (Scheme 5). Wittig-Horner re-



Scheme 5. Preparation of the CD-ring segment (unless otherwise noted, the reactions were performed at ambient temperatures). Reagents and conditions: a) **19**, LiCl, DBU, CH₃CN, 20 h, 77% (E/Z=20:1); b) CF₃CO₂H, H₂O, 3.5 h, 90%; c) NaOAc, Ac₂O, 140 °C, 30 min, 99%; d) Ce(NH₄)₂(NO₂)₆, CH₃CN, H₂O, 0°C, 10 min, quant. DBU=1,8-diazabicyclo[5.4.0]undec-7-ene.

action of **15** with phosphonate **19**^[19] gave the corresponding unsaturated ester, the *tert*-butyl ester of which was selectively hydrolyzed under acidic conditions to give unsaturated acid **17**.^[20] Treatment of **17** with acetic anhydride in the presence of sodium acetate afforded naphthalene **18a**,^[20,21] which corresponds to the CD-ring system.

To annulate the E ring, naphthalene **18a** was oxidized to the corresponding naphthoquinone **20a** in preparation for a Diels–Alder reaction with siloxydiene **21** (Table 1).^[15] Because the regioselectivity of the Diels–Alder reaction of 1,4naphthoquinones is highly sensitive to the substitution pattern, we tested four related quinones **20a–d** with different protecting groups (R) for comparison purposes.

The reactions were conducted by adding diene 21 to a solution of quinone 20a-d in THF at 0°C. After stirring for

Table 1. Diels-Alder reactions of quinone 20 a-d. MeO MeO THE CO₂Et ÓTMS ö TMSÓ 21 20a-d OH 0 OR workup MeC conditio (i) hydrolysis CO₂Et MeC CO₂Et ii) tautomerization R'Ó č ö liii) oxidation 22' R' = H 23 22': R' = Me Yield^[b] [%] Entry Substrate R Hydrolysis 22:22':23 conditions^[a] 1 20 a SiO 2.0.134 Ac 2 MOM 40 20 b SiO₂ 4:3:1 3 20 c 34:1:0 59 SiO₂ Me 4 20 d Η SiO₂ 0:0:1 49 5 20 b nBu₄NF 1:0:0 84 MOM

[a] In THF at 0 °C, 7 h. [b] Combined yield of 22, 22', and 23.

7 h at room temperature, acidic silica gel was added to hydrolyze the silyl acetal. The resulting ketone underwent tautomerization and spontaneous air-oxidation to afford a varied mixture of desired anthraquinone 22, undesired methyl ether 22', and undesired regioisomer 23 (entries 1–3). When 20d was used as the substrate, only undesired regioisomer 23 was obtained (entry 4).^[22]

For ease later in the synthesis (for deprotection of the R group in 22 in the palladium-mediated cyclization reaction, cf. Scheme 6) and to optimize the regioselectivity of the Diels-Alder reaction, we decided to employ MOM ether 20b as the substrate for the synthesis. The remaining problems in this context were the low yield and the presence of another mode of acetal cleavage that produces methyl ether 22'. TLC analysis suggested that the cycloaddition step proceeded without a problem and it seemed that the low yield stemmed from the workup conditions: 1) The hydrolysis of the silyl acetal, 2) the tautomerization step to the hydroquinone, and 3) the oxidation step.

Extensive optimization studies revealed a protocol that would achieve smooth conversion to desired anthraquinone **22** with complete regioselectivity (entry 5). Thus, upon treatment of the crude cycloadduct with nBu_4NF under air in THF, the color changed from yellow to purple, and after several minutes, the color returned to light yellow, which gave the anthraquinone **22** in a yield of 84%.

Anthraquinone 22, thus obtained, was converted into anthracene 26 because the quinone moiety would be incompatible with the reductive conditions of the pinacol cyclization reaction (Scheme 6). After protection of the phenol in 22, the resulting benzyl ether 24 was subjected to reductive dimethylation to afford anthracene 25, which was hydrolyzed to give carboxylic acid 26.

With A-ring segment 14 and CDE-ring segment 26 in hand, their combination was attempted. After esterification of 14 and 26 by using 2,4,6-trichlorobenzoyl chloride, $^{[23,24]}$



FULL PAPER

Scheme 6. Preparation of biaryl lactone **28** (unless otherwise noted, the reactions were performed at ambient temperatures). Reagents and conditions: a) Cs_2CO_3 , BnBr, DMF, 7.5 h, 89%; b) $Na_2S_2O_4$, cat. nBu_4NBr , THF, 30 min; c) Me_2SO_4 , 50% aq. KOH, 1 h, 98% (2 steps); d) 5 M aq. KOH, EtOH, reflux, 1.5 h, 91%; e) 2,4,6-trichlorobenzoyl chloride, Et₃N, toluene, 3 h, then **14**, DMAP, 0.5 h, 94%; f) CF_3CO_2H , CH_2Cl_2 , 2 h, 87%; g) Pd(OCOCF₃)₂, P(tBu)₃, PPh₃, $tBuCO_2Na$, DMA, 110°C, 25 min, 64%. Bn = benzyl, TMS = trimethylsilyl, DMAP = 4-dimethylaminopyridine, DMA = *N*,*N*-dimethylacetamide.

the resulting ester was treated with CF_3CO_2H to detach the MOM group to give ester **27** in high yield. Because ester **27** turned out to be rather labile, easily undergoing color change upon exposure to air and/or light, it was immediately subjected to the palladium-mediated cyclization reaction.^[13] After careful optimization, we were able to obtain lactone **28** in a yield of 64% by using Pd(OCOCF₃)₂. Unfortunately, handling of this advanced intermediate **28** was even more difficult, particularly in terms of its air-sensitivity, and therefore, we decided to abandon this synthetic route.

Successful pathway to aglycon: Recognizing the instability of anthracene derivatives, we decided to postpone the construction of the E ring until a later stage of the synthesis (Scheme 7). To secure the regioselectivity of the Diels–Alder reaction with siloxydiene **21**, we planned to introduce a chlorine atom (X=Cl) into the CD-ring precursor **VIII**' en route to the ABCD-ring segment **IX**.

Chloronaphthalene **32**, the CD-ring intermediate,^[4a] was synthesized from the known compound $29^{[25]}$ (Scheme 8). Regioselective hydroxymethylation of **29** under Casiraghi conditions,^[26] followed by methylation of the phenol, and MnO₂ oxidation of the benzylic alcohol afforded benzalde-hyde **30**. The same sequence of reactions, which was already described for the conversion of **15** into **18** (see Scheme 5), was applied to aldehyde **30** to give naphthalene **31** in an overall yield of 76%. Hydrolysis of ester **31** gave carboxylic acid **32** in a yield of 87%.



Scheme 7. Naphthalene route.



Scheme 8. Preparation of the CD-ring segment (unless otherwise noted, the reactions were performed at ambient temperatures). Reagents and conditions: a) (HCHO)_m, Me₂AlCl, CH₂Cl₂, 5 h, 93 %; b) Me₂SO₄, K₂CO₃, acetone, reflux, 13 h, 91 %; c) MnO₂, CH₂Cl₂, 40 °C, 17 h, 90 %; d) NaH, **19**, THF, 0 °C \rightarrow RT, 1 h; e) CF₃CO₂H, H₂O, 40 min; f) NaOAc, Ac₂O, reflux, 1 h, 76% (3 steps); g) 3 M aq. NaOH, THF, EtOH, 70 °C, 1 h, 87%.

Joining the A-ring segment **14** and the CD-ring segment **32** was achieved by using a water-soluble carbodiimide (EDCI: 1-(3-dimethyaminopropyl)-3-ethylcarbodiimide hydrochloride) to give ester **33**, which was ready for biaryl bond formation (Scheme 9). After considerable optimization, the palladium-catalyzed cyclization of ester **33** proceeded smoothly in the presence of sodium pivalate to give tetracycle **34**.^[13] However, the product **34** turned out to be highly



Scheme 9. Preparation of triol **35**. Reagents and conditions: a) EDCI, DMAP, CH_2Cl_2 , RT, 3 h, 78 %; b) Pd(OAc)₂, PPh₃, *t*BuCO₂Na, DMA, 110 °C, 1.5 h; c) NaBH₄, THF, MeOH, -40 °C, 3 h, 86 % (2 steps).

9796

K. Suzuki et al.

prone to hydrolysis during purification by silica gel column chromatography. Lactone **34**, which was once obtained in a small amount, was stable as a crystalline material. In spite of various attempts to improve the workup procedure, we were unable to find a suitable protocol for the high-yielding formation of **34**. At this juncture, we opted to reduce the crude product of **34** with NaBH₄ to obtain alcohol **35** in a yield of 86%. Thus, at this stage, the opportunity for the planned asymmetric lactone-opening was lost and we decided to resort to optical resolution as an alternative means of obtaining the requisite biaryl enantiomer.

Scheme 10 shows the optical resolution protocol. After detachment of the MOM group in **35**, the benzyl alcohol



Scheme 10. Optical resolution protocol. Reagents and conditions: a) 6 M aq. HCl, DME, 3 h, 50 °C, 93 %; b) TBSCl, imidazole, DMF, RT, 25 min, 84 %; c) (1*S*,4*R*)-(–)-camphanoyl chloride, DMAP, pyridine, RT, 20 h; separation by silica gel column chromatography (see text), 38 %. DME = 1,2-dimethoxyethane, TBS = *tert*-butyldimethylsilyl.

moieties in the resulting tetrol were protected by TBS groups to give bis-phenol **36**, which was treated with (–)-(1*S*,4*R*)-camphanoyl chloride. Esterification occurred selectively at the phenol *ortho* to the methoxycarbonyl in the A ring to give a diastereomeric mixture of monoesters **37a** and **37b**. Column chromatography (SiO₂, hexane/EtOAc=7:3) allowed separation of these diastereomers, which gave the more polar isomer **37a** (R_f =0.34, hexane/EtOAc=7:3; 38%) and the less polar isomer **37b** (R_f =0.42, hexane/EtOAc=7:3; 40%), respectively. Isomer **37a** possessed the requisite *M* configuration.^[27]

The requisite diastereomer **37a** was then converted in two steps into enantiopure tetrol (*M*)-**38** (Scheme 11). After protection of the phenols in (*M*)-**38** as methyl ethers, the remaining benzyl alcohols were oxidized to afford enantiomerically pure biaryl dialdehyde (*M*)-**39**.^[28] Upon treatment of (*M*)-**39** with SmI₂ (0 °C, THF, 10 min), the pinacol cyclization reaction proceeded smoothly to afford *trans*-diol **40** as a single product in quantitative yield with high stereoselectivity (>99% enantiomeric excess (*ee*)), as shown by HPLC analysis.^[29] The circular dichroism (CD) measurement revealed an absolute configuration of 5*S*,6*S* for *trans*-diol



Scheme 11. Synthesis of benanomicinone methyl ester (unless otherwise noted, the reactions were performed at ambient temperatures). Reagents and conditions: a) HF, CH₃CN, $0^{\circ}C \rightarrow RT$, 45 min, 97%; b) K₂CO₃, MeOH, 19 h, quant; c) MeI, K₂CO₃, acetone, 40°C, 11 h, 91%; d) MnO₂, CH₂Cl₂, 26 h, 79%; e) SmI₂, THF, 0°C, 10 min, quant; f) Ac₂O, pyridine, DMAP, 20 min, 98%; g) Ce(NH₄)₂(NO₃)₆, CH₃CN, H₂O, 0°C, 10 min, quant; h) diene **21**, THF, 0°C \rightarrow RT, 2 h; i) SiO₂, 12 h; j) K₂CO₃, THF, CH₂Cl₂, 2.5 h, 90% (3 steps); k) BCl₃, CH₂Cl₂, -10°C, 30 min, 99%; l) 2M aq. NaOH, 70°C, 2.5 h; m) D-alanine methyl ester-HCl, BOP, Et₃N, DMF, 1.5 h, 80% (2 steps).

40.^[10] Pleasingly, the axial stereochemistry in **39** was completely transmitted into the stereogenicities of *trans*-diol **40**.

For elaborating tetracycle **40** into the pentacyclic full carbon skeleton, the two hydroxy groups were protected as diacetate **41**. Reaction of **41** with $Ce(NH_4)_2(NO_3)_6$ (CAN) effected the selective oxidation of the D ring to afford a quantitative yield of chloroquinone **42**, which was then subjected to the Diels–Alder reaction with siloxydiene **21** to annulate the E ring.^[15]

We were pleased to find that the following optimized protocol enabled the formation of pentacycle **43** in high yield. This method for E-ring annulation turned out to be highly reliable and was repeatedly employed in the later syntheses (cf. Scheme 17, Scheme 19, and Scheme 20).

A solution of siloxydiene **21** in THF was added to a solution of **42** in THF at 0°C and stirring was continued until the consumption of the starting material was complete (TLC assay, 2 h). The resulting silyl acetal was selectively hydrolyzed by treatment with acidic silica gel. Elimination of HCl was achieved with K_2CO_3 to give naphthacenequinone **43** in a yield of 90% from **42**. Note that the chloro group played two roles: 1) To direct the regioselective cycloaddition and 2) to facilitate the aromatization.

Having pentacycle 43 in hand, selective removal of the methyl ethers proximal to the carbonyl groups was readily achieved with BCl₃ to give ester 44, which was saponified to give carboxylic acid 45. Compounds 44-46 were scarcely soluble in solvents other than water, DMF, and/or DMSO,

presumably owing to the presence of the quinone moiety, the handling of **45** was especially difficult.

Condensation of acid **45** with D-alanine methyl ester was effected by using benzotriazol-1-yloxytris(dimethylamino)-phosphonium hexafluorophosphate (BOP) in DMF, which gave benanomicinone methyl ester (**46**) in a yield of 80% from **45**, which was identical to an authentic specimen.^[30]

Glycosylation study-towards the formation of benanomicin B: With aglycon 46 in hand, the stage was set for the final installation of the carbohydrate moiety. Considering the potential problems expected from the presence of the quinone functionality (e.g., deactivation of the Lewis acid used for the glycosylation,^[31] poor solubility), we chose to reduce the oxidation level of the aglycon moiety (Scheme 12). According to the procedure of Oki and co-workers, conversion of anthraquinone 46 into the corresponding anthraquinone diacetate 48 was carried out.^[2g] After protection of the 1,2diol in 46 as an acetonide, the resulting product was subjected to reductive acetylation (acetic anhydride, zinc powder, pyridine) to afford anthracene 47, in which all of the phenols as well as two hydroquinone phenoxides, reductively generated in situ, were fully acetylated. Again, careful handling was required to cope with the sensitive nature of the anthracene derivative to light and oxygen after the quinone functionality of 46 had been reduced. Careful removal of the acetonide and purification afforded pentacycle 48 in a yield of 68% (three steps), which was ready for glycosylation.



benanomicin B·HCI (2b·HCI)

Scheme 12. Glycosylation study and synthesis of benanomicin B (unless otherwise noted, the reactions were performed at ambient temperatures). Reagents and conditions: a) $CH_2=C(OCH_3)CH_3$, TsOH·H₂O, DMF, 18 h; b) Ac₂O, Zn, pyridine, 10 h, 73 % (2 steps); c) TsOH·H₂O, CH₃CN, H₂O, 2 h, 93 %; d) fluoride **51**, Cp₂HfCl₂/AgClO₄ (1:2), MS4A, CH₂Cl₂, -12 °C, 25 min, 44 % (**49/50**=3:2); e) H₂, Pd/C, 1m aq. HCl, MeOH, 6 h; f) 1m aq. NaOH, MeOH, 2 h, then 2m aq. HCl, 68 % (2 steps). Ts=*p*-toluene-sulfonyl.

Benanomicin B (2b) was selected as the initial target. In accordance with the results of model studies on the glycosidation of glycosyl fluoride 51, Cp₂HfCl₂ and AgClO₄ (1:2 ratio) were used in combination as promoters.^[32,33] A suspension of Cp₂HfCl₂ and AgClO₄ in the presence of molecular sieves (MS4A) in CH₂Cl₂ was stirred at room temperature for 10 min and chilled to -78°C. A CH₂Cl₂ solution of glycosyl fluoride 51 and diol 48 was added to this mixture. Reaction at -12°C for 25 min afforded two regioisomeric βglycosides 49 and 50 in yields of 26 and 18%, respectively (Scheme 12). Extensive NMR studies, which included HMBC and HMQC analyses, showed that 49 was the necessary regioisomer for the total synthesis.^[34] Although unfortunate, the poor regioselectivity at this stage was not surprising if one took into consideration the approximate C_2 symmetry of the diol.

In any event, the final stage of the total synthesis was examined. After reduction of the azide in **49** by hydrogenolysis in the presence of aqueous HCl, treatment with 1 M aqueous NaOH accomplished the detachment of all of the acyl groups and hydrolysis of the methyl ester. The reaction mixture was acidified with 2 M aqueous HCl (pH 3.5) to afford the final product, benanomicin B hydrochloride (**2b**-HCl), as a red solid. As the ¹H NMR spectrum proved highly dependent on the pH, the concentration, and the temperature, identification of **2b** was not an easy issue. However, its identification was finally achieved by the measurement of a ¹H NMR spectrum of a sample that contained a mixture of synthetic and authentic **2b** at a low pH (ca. 3.5).^[35]

Second-generation total synthesis

Improved strategies for the regio- and stereocontrolled total synthesis of BpAs: In the total synthesis described above, two issues remained unsolved (Figure 4): 1) The axially



Figure 4. The two remaining tasks for the fully-controlled total synthesis.

chiral biaryl dialdehyde (*M*)-**39** was only obtained by tedious optical resolution, which inevitably produced the useless enantiomer (*P*)-**39** and 2) the sugar moiety was introduced with poor regioselectivity, originating from the approximate C_2 -symmetric structure. Seeking a fully controlled total synthesis, we next addressed these two problems.^[4b]

Control of axial stereochemistry: Our original plan to tackle the first problem, that is, the control of axial stereochemistry, involved using the Bringmann-type asymmetric ringopening of the biaryl lactone.^[14] This strategy was, however, hampered by the difficulty in preparing the related lactone **34** as a result of its high lability to hydrolysis during silica gel chromatography (cf. Scheme 9). Thus, the prerequisite was the preparation of the lactone itself, which had seemed hopeless in the early phase of our investigation.

Fortunately, however, the following protocol enabled us to achieve this goal (Scheme 13). After Yamaguchi esterification of carboxylic acid **52** with the A-ring segment **53** (TIPS was used as the protecting group of the benzyl alcohol (TIPS: triisopropylsilyl)), the MOM protecting group of the resulting ester was removed by CF_3CO_2H to afford the ester **54**, which was subjected to the palladium-catalyzed cyclization reaction. Again, lactone product **55** was highly labile to hydrolysis during purification by silica gel chromatography. However, addition of water to the reaction mixture at 0°C effected the precipitation of lactone **55** as fine needles that were collected by filtration. The overall yield from **52** was 59% in three steps.^[34]

Prior to asymmetric ring-opening, we observed the dynamic behavior of lactone 55 by using variable-temperature



Scheme 13. Preparation of biaryl lactone 55. Reagents and conditions: a) 2.4.6-Trichlorobenzovl chloride, Et₃N, DMAP, toluene, RT, 20 min, 99%; b) CF₃CO₂H, CH₂Cl₂, RT, 3.5 h, 99%; c) Pd(OAc)₂, PPh₃, tBu-CO2Na, DMA, 110°C, 20 min, 60%.

¹H NMR analysis of lactone **55** (Figure 5). At -15°C, a singlet was observed at 4.95 ppm, which could be assigned to the benzyl protons. Upon cooling, the peak broadened and finally started to split at -75°C (coalescence temperature) and two peaks appeared upon further cooling (-95°C).

The estimated barrier for the conformational interchange was $\Delta G^{\pm} = 9.8 \text{ kcal mol}^{-1}$, which indicated an extremely facile interconversion between enantiomers (P)-55 and (M)-55.



Figure 5. Variable-temperature ¹H NMR spectra of 55.

Chem. Eur. J. 2007, 13, 9791-9823

© 2007 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

With the requisite lactone 55 in hand, asymmetric ringopening was examined (Table 2). For the enantioselective



[a] No reaction. [b] Amide 57a was obtained in a yield of 54% with a



ring-opening of 55, we first applied borane reduction catalyzed by a chiral oxazaborolidine which, unfortunately, gave

enantioselectivity poor (entry 1). The enantioselectivities were also disappointing in reactions carried out with LiBH₄ or NaBH₄ in the presence of various stoichiometric chiral modifiers, such as N,Ndibenzoylcysteine (entries 2 and 3)^[36] and diethyl L-tartrate (entries 4 and 5).^[37] However, an unexpected but important pointer was obtained when Lvaline was employed (entry 6).^[38] Although the result from the reaction itself was again poor (8% yield, 2% ee), the formation of a sizable amount of a polar by-product was noted which was identified as amide 57 a, that is, the direct addition product of L-valine. Importantly, amide 57 a was highly enriched in one of the diastereomers (90:10 diastereomeric ratio).

This finding encouraged us to pursue the diastereoselective ring-opening of 55 by

www.chemeurj.org

9799

using various stoichiometric chiral nucleophiles (Table 3). Of course, the first choice was L-valine in the absence of $NaBH_4$. However, no reaction occurred, presumably as a result of the insolubility of the amino acid in less polar or-

Table 3. Diastereoselective biaryl lactone ring-opening.



[a] The ratio was determined by ¹H NMR analysis. [b] RT=room temperature. [c] No reaction. [d] Ester **58** was obtained in a yield of 14 % (3:2 d.r.). [e] (-)-Norephedrine was employed.



ganic solvents such as CH_2Cl_2 (entry 1). When L-valine ethyl ester was used as a "soluble nucleophile", a slow reaction proceeded, which gave the ring-opened product in a yield of 51%, albeit with poor stereoselectivity (entry 2).

A breakthrough came with the use of an amino alcohol; treatment of lactone **55** with (*S*)-valinol at room temperature for 1.8 h gave amide **57c** in a yield of 72% in a diastereomeric ratio (d.r.) of 90:10 (entry 3). Although the result was adequate, the material balance was low. TLC inspection (TLC A, Table 3) showed three new polar spots (R_f =0.38, 0.25, and <0.1, hexane/EtOAc=1:1), the first two spots correspond to the minor/major diastereomers of amide **57c** and the most polar product corresponds to a small amount (14%) of the *O*-addition product **58** formed with a low diastereomeric ratio (3:2).

In contrast, when the reaction was prolonged (11 h), ester **58** disappeared (see TLC B, Table 3) and only amide **57c** was obtained in an improved yield (88%) with a slightly lower diastereomeric ratio (84:16, entry 4). We ascribed this phenomenon to the in situ conversion of ester **58** into amide **57c**.

Indeed, such an N,O acyl rearrangement happens without mutation of the axial stereochemistry. Upon retreatment with valinol, isolated ester **58** (d.r. 3:2) was converted into amide **57c** in a yield of 86% (d.r. 3:2).^[39] Thus, albeit a

minor path, the competing *O*addition process, which gave ester **58**, serves as an indirect supplier of amide **57c**, although with a lower diastereomeric ratio.

To eliminate this stereodeteriorating O-addition pathway, the O-methylated valinol derivative was used. However, a very slow and nonselective ring-opening was observed, after one month amide **57d** was obtained in a yield of 82% with a 1:1 diastereomeric ratio (entry 5). Thus, the hydroxy group in the amino alcohol is essential for the smooth progress of the reaction and for high diastereoselection.

Fortunately, optimization experiments showed that the reaction temperature was decisive. The reaction at 0°C for 43 h gave the best result to afford a yield of 90% of amide **57 c** in a diastereomeric ratio of 91:9 (entry 6). The *O*-addition pathway seemed to be less pronounced under these conditions.

Hoping for further enhance-

ment of the diastereoselectivity, other amino alcohols, such as *tert*-leucinol, phenylalaninol, alaninol, and norephedrine, were also examined, but their use only resulted in lower selectivities (entries 7–10).

In summary, (*S*)-valinol turned out to be the best chiral nucleophile for this diastereoselective biaryl lactone ringopening process. The structure of the major isomer of **57 c** was unambiguously established by single-crystal X-ray analysis. After separation of the diastereomers of **57 c** by column chromatography (silica gel, hexane/EtOAc=1:2), the major isomer of **57 c** gave single crystals suitable for X-ray analysis (Figure 6).^[40] The biaryl axis was revealed to have the *P* configuration, which was the wrong stereochemistry for our synthetic purposes (cf. Scheme 11).

Thus, (R)-valinol was proven to be the right enantiomer for establishing the requisite M configuration in the biaryl intermediate in the total synthesis (see Scheme 15).

Discrimination of the 1,2-diol moiety: To address the pivotal issue of diol discrimination at the B ring, we planned a semi-pinacol cyclization (Scheme 14). The hope was that



Figure 6. X-ray structure of the major isomer of **57c**. Hydrogen atoms have been omitted for clarity.



Scheme 14. Semi-pinacol cyclization strategy.

mono-masked diol **XII** would be accessible from the acetal aldehyde **XI** by activation with an acid (e.g., Lewis acid) and a net two-electron reduction. If the strategy proved to be viable then the product would be ideally suited to regio-selective installation of the sugar moiety.^[6] Of course, we expected that a careful choice of the reaction conditions would be necessary because the timing between the generation of the oxocarbenium intermediate (**XI** \rightarrow **XII**) and net two-electron reduction (**XIII** \rightarrow **XII**) would be critical.

With this scenario in mind, we began extensive modeling studies^[6] by using acetal aldehyde **59** as the substrate (Table 4). The first attempt proceeded smoothly by treatment of (\pm) -**59** with SmI₂^[9] in the presence of BF₃·OEt₂.^[41] Thus, SmI₂ in THF (0.1 M) was added to a solution of (\pm) -**59** and BF₃·OEt₂ in degassed THF at -78 °C. Stirring at 0 °C for 30 min afforded cyclized product **60** in a yield of 70% in a fully *trans*-selective manner. However, this key reaction turned out to be capricious because all of the subsequent attempts suffered from poor reproducibility; the yields fluctuated widely in the range of 0 to 70% with concomitant formation of varying amounts of by-products (e.g., cyclic acetal **61**, aldehyde **62**, alcohol **63**, and/or dialdehyde **64**), even with slight changes in the reaction parameters (entry 1).

After numerous unfruitful experiments, we finally found that the presence of a suitable proton source was the key to securing reproducibility. In the presence of H_2O , the reaction occurred cleanly to give semi-pinacol **60** in a yield of 85% (entry 2) and methanol proved to be even more effective (entry 3).

As the need for a proton source for the facile formation of 60 is clearly evident from entries 2 and 3, other acids were also examined in the absence/presence of a proton





			[%]				[%]
1	BF ₃ •OEt ₂	_	_[a]	6	TiCl ₄	_	_[b]
2	$BF_3 \cdot OEt_2$	H_2O	85	7	$TiCl_4$	H_2O	32 ^[d]
3	$BF_3 \cdot OEt_2$	MeOH	95	8	TfOH	-	-
4	TMSOTf	-	_[b]	9	TfOH	H_2O	29 ^[e]
5	TMSOTf	H_2O	71 ^[c]				

[a] The yield of **60** fluctuated, varying between 0 and 70%, and sizable amounts of by-products were produced. [b] Many unidentified by-products were produced. [c] Acetal **61** was obtained in a yield of 21%. [d] Dialdehyde **64** was obtained in a yield of 60%. [e] Acetal **61** was obtained



source. In all cases (entries 4–9), compound **60** was only formed when H₂O was added to the reaction (entries 5, 7, and 9), otherwise a complex mixture was produced (entries 4, 6, and 8). When TMSOTf was employed with H₂O, compound **60** was formed in a yield of 71% along with acetal **61** (entry 5). In the case of a combination of TiCl₄ and H₂O, the yield of **60** decreased to 32% and was accompanied by the formation of dialdehyde **64** (entry 7). TfOH was also used as a Brønsted acid in the presence of H₂O. The yield of **60** was 29% with **61** was also formed in a yield of 41% (entry 9).

In summary, $BF_3 \cdot OEt_2$ turned out to be the best acid for the semi-pinacol cyclization reaction.

To gain a further insight into this transformation, the reaction was monitored by time-resolved IR spectroscopy (Figure 7). Thus, in a solution of **59** in THF at -78 °C, absorptions were observed that are characteristic of the C=O stretching of the methoxycarbonyl (1732 cm⁻¹) and formyl groups (1698 cm⁻¹). Amazingly, these signals persisted at -78 °C even after the addition of BF₃·OEt₂, MeOH, and SmI₂ (Time A).

However, when the reaction temperature was raised to 0 °C (Time B), the absorption at 1698 cm⁻¹ disappeared with the emergence of a strong absorption at 1572 cm⁻¹ which could be assigned to the stretching vibration of a ketyl radical ('C-O⁻) species.^[42]

This observation suggested that the one-electron reduction of the formyl group to form the ketyl radical species is the initial step, though the precise mechanism still remains uncertain.



Figure 7. Time-resolved IR monitoring (React-IR) of the semi-pinacol reaction of **59**.

Preparation of enantiopure tetracycle (S,S)-60: After establishing these two stereoselective processes, that is, the lactone ring-opening of biaryl lactone with valinol and the semi-pinacol cyclization of acetal aldehyde 59, we applied these to the preparative-scale synthesis of chiral, nonracemic 60 (Scheme 15). We were pleased to find that asymmetric ring-opening was viable for the gram-scale reaction, and lactone 55 (2.0 g), upon treatment with (*R*)-valinol (0 °C, 43 h), afforded a combined yield of 90% of amide 57 c with a diastereoselectivity of 91:9. The desired diastereomer 57 c with



Scheme 15. Preparation of tetracycle (*S*,*S*)-**60** (unless otherwise noted, the reactions were performed at ambient temperatures). Reagents and conditions: a) (*R*)-Valinol, CH₂Cl₂, 0°C, 43 h, 90% (*M*/*P*=91:9); b) BnBr, Cs₂CO₃, DMF, 0°C, 1 h, 97%; c) PPh₃, I₂, imidazole, CH₂Cl₂, 20 min, quant; d) MeOTf, 2,6-di-*tert*-butylpyridine, CH₂Cl₂, 1 h; e) L-Selectride, 0°C, 20 min; f) SiO₂, 17 h, 96% (3 steps); g) BnOTMS, cat. TMSOTf, toluene, $-78 \rightarrow -15$ °C, 4 h, quant; h) *n*Bu₄NF, THF, 1 h, 0°C, quant; i) MnO₂, CH₂Cl₂, 15 h, 99%; j) BF₃·OEt₂, SmI₂, MeOH, THF, $-78 \rightarrow 0$ °C, 30 min, 95%. L-Selectride = lithium tri-*sec*-butylborohydride.

R,M configurations was isolated as yellow crystals (1.9 g) by silica gel column chromatography and recrystallization.

After the selective benzylation of the two phenolic hydroxy groups in the amide (R,M)-**57 c**, the hydroxy amide moiety in the resulting bis-benzyl ether **65** was cyclized (PPh₃, I₂, imidazole), which gave oxazoline **66** in quantitative yield. After *N*-methylation of **66**, the activated C=N bond underwent facile reduction with L-Selectride, and the resulting *N*,*O*-acetal was hydrolyzed by exposure to silica gel to give aldehyde **67** in a yield of 96%.^[43] Conversion of **67** into the corresponding dibenzyl acetal,^[44] followed by detachment of the silyl protection, and MnO₂ oxidation afforded the enantiopure acetal aldehyde (*M*)-**59**, which was ready for the key cyclization reaction.^[45]

SmI₂ (2.5 equiv) was added to a solution of (*M*)-**59** in the presence of BF₃·OEt₂ (3.0 equiv) and MeOH (1.0 equiv) in THF at -78 °C. The temperature was immediately raised to 0 °C and the stirring was continued for 30 min to give (*S*,*S*)-**60** in a yield of 95%. Importantly, this semi-pinacol cyclization reaction also allowed the specific transmission of the axial stereochemistry, as shown by HPLC analyses on a chiral stationary phase (Figure 8).^[46]



Figure 8. HPLC analyses of the semi-pinacol cyclization reaction. HPLC conditions: for (*M*)-**59** and *rac*-**59**, Chiralpak AD-H (0.46×25 cm) column, 25 °C, UV detection at 254 nm, flow rate: 1.0 mLmin⁻¹, eluent: hexane/*i*PrOH 9:1; for (*S*,*S*)-**60** and *rac*-**60**: Chiralpak AD-H (0.46×25 cm) column, 25 °C, UV detection at 300 nm, flow rate: 1.0 mLmin⁻¹, eluent: hexane/EtOH 9:1.

Divergent approach to BpAs: Enantiopure tetracycle (*S*,*S*)-60, thus obtained, could serve as an ideal platform for the divergent access to various natural/unnatural congeners of BpAs. Figure 9 represents the three branching points for the divergency: 1) introduction of the amino acid moiety, 2) glycosylation with a disaccharide derivative, and 3) construction of the E ring by Diels–Alder reaction with a diene. To demonstrate such divergency, we report herein the total syn-



Figure 9. Divergent access to various BpAs.

thesis of three natural BpAs, that is, benanomicin B (2b), pradimicin A (1a), and benanomicin A (2a). These three BpAs share D-alanine as the amino acid, but differ in their disaccharide moieties. Among the possibilities in assembling these three modules, we chose to form the amide bond first, then perform the glycosylation reaction, and finally the synthesis was completed by E-ring annulation. This choice has the advantage of reserving the generation of the quinone functionality until the latest stage because we knew that quinones might pose problems associated with chemical transformations and poor solubility (cf. Scheme 11).

Scheme 16 shows the installation of the amino acid, the methyl ester of D-alanine. Tetracycle (S,S)-**60** was hydrolyzed under basic conditions to afford the corresponding carboxylic acid. This saponification reaction proceeded only under harsh basic conditions that nonetheless did not cause aromatization. The resulting carboxylic acid was condensed with the methyl ester of D-alanine by using BOP to give the



Scheme 16. Installation of the amino acid moiety into (S,S)-60. Reagents and conditions: a) 5 M aq. KOH, EtOH, sealed tube, 100 °C, 3 h; b) BOP, Et₃N, D-alanine methyl ester-HCl, DMF, RT, 1 h, 84 % (2 steps).

amide **68** in excellent yield whose structure was unambiguously established by single-crystal X-ray analysis (Figure 10).^[47]



Figure 10. X-ray structure of amide **68**. Hydrogen atoms have been omitted for clarity.

Amide **68** served as a key intermediate for the divergent synthesis of the three natural products **2b**, **1a**, and **2a**, prepared by installing three different disaccharides followed by E-ring annulation.

Benanomicin B (2b): The first target was benanomicin B (2b), which has a primary amine in the sugar moiety. Glycosyl fluoride 51 was used as the glycosyl donor (cf. Scheme 12) because it has an azide that can be converted into the corresponding primary amine at the appropriate time (Scheme 17).

The glycosylation process was initially attempted by using a combination of Cp2HfCl2 and AgClO4 (1:2 ratio). Although this procedure had previously given the best result,^[33] it only induced the decomposition of **68** at the reaction temperature of -36°C.^[48] After careful reinvestigation of the protocol (Ag salt, molarity, and temperature), optimal conditions were found: Treatment of fluoride 51 and alcohol 68 with a 1:2 ratio of Cp₂HfCl₂ and AgOTf (instead of AgClO₄) in the presence of MS4A at -36 °C for 11 h gave β -glycoside **69** in a yield of 72% along with its α anomer in a yield of 9%. After separation, β anomer 69 was oxidized with CAN and the resulting unstable chloroquinone was immediately subjected to a Diels-Alder reaction with siloxydiene 21.^[15] The previously described protocol (see Scheme 11, $42 \rightarrow 43$) worked well to afford pentacycle 70 in a fully regiocontrolled manner (74% overall yield in three steps).

The next stage was the removal of the protecting groups in **70**, which was rather difficult as a result of the high polarity and the poor solubility of the partially deprotected intermediates in organic solvents. Careful optimization showed that the removal of the four acyl protecting groups and saponification of the methyl ester was possible by treatment

Chem. Eur. J. 2007, 13, 9791-9823

© 2007 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

www.chemeurj.org

- 9803



Scheme 17. Synthesis of benanomicin B (unless otherwise noted, the reactions were performed at ambient temperatures). Reagents and conditions: a) Fluoride **51**, Cp₂HfCl₂/AgOTf (1:2), MS4A, CH₂Cl₂, -36° C, 11 h, 9% for the α anomer, 72% for the β anomer; b) CAN, H₂O, CH₃CN, 20 min; c) diene **21**, THF, 20 min, then SiO₂, 8 h; d) K₂CO₃, CH₂Cl₂, THF, 40°C, 3 h, 74% (3 steps); e) 2M aq. NaOH, MeOH, 8.5 h; f) H₂, Pd/C, 1M aq. HCl, MeOH, DMF, 2 h, then 2M aq. HCl, 53% (2 steps), for purification see text.

with 2M aqueous NaOH (room temperature, 8.5 h), which gave compound **71**. Although the acyl protecting groups were removed fairly rapidly (2.5 h), hydrolysis of the methyl ester required a longer time (8.5 h).

The final stage of the synthesis required the removal of the three benzyl protecting groups and conversion of the azide into the primary amine, which could be achieved under catalytic hydrogenation conditions. This stage was again troublesome owing to the poor solubility of the partially deprotected intermediates and also to catalyst deactivation by the resulting amino group. After considerable screening of the reaction conditions, an appropriate choice of solvent and additives allowed us to achieve the total synthesis: Treatment of 71 with 5% Pd/C in MeOH and DMF (3:1)^[49] in the presence of dilute aqueous HCl^[50] effected the desired conversion. During the filtration of the catalyst in air through Hyfro Super-Cel, the color of the filtrate changed from orange to purple, which suggested the transformation of the in situ formed hydroquinone back to the quinone.

The filtrate was purified by successive column chromatography on Cosmosil (C18, CH₃CN/H₂O=3:7, phosphate buffer pH 3) and then on Sephadex LH-20 (DMF) to afford benanomicin B as a DMF solvate that was dissolved in MeOH and the pH adjusted to 3.5 by the addition of 2 M aqueous HCl. After removal of the solvent, the residue was dissolved in a small amount of DMSO to which CHCl₃ was added. The resulting red precipitate was collected by filtration to afford benanomicin B hydrochloride (**2b**-HCl), which was identical to an authentic specimen (m.p. 213–215 °C, $[\alpha]_D^{22} = +360$ (c = 0.0501 in H₂O); lit.:^[1a] m.p. > 220 °C, $[\alpha]_D^{30} = +353$ (c = 0.05 in H₂O)).

Pradimicin A (1a): The second target was pradimicin A (1a), the most abundant congener of the pradimicins. This compound possesses an N-monomethylamino group instead of the primary amino group in 2b. Because the direct conversion of a primary amine into the corresponding monoal-kylamine is often hampered by bis-alkylation,^[51] indirect routes are often employed, such as temporary protection, al-kylation, and deprotection.

However, we have developed a method for the selective conversion of an azide into the corresponding *N*-monomethylamino group without forming the bis-methylation product [Eq. (4)].^[7] Trimethylphosphine reacts with an azide to form the corresponding iminophosphorane,^[52] which undergoes aza-Wittig reaction with (HCHO)_n to form the corresponding imine. In situ reduction of this imine with NaBH₄ affords the requisite *N*-monomethylamine.

$$R^{-}N_{3} \xrightarrow{\mathsf{PMe}_{3}} R^{-}N^{=}\mathsf{PMe}_{3} \xrightarrow{(\mathsf{HCHO})_{n}} R^{-}N^{=}\mathsf{CH}_{2} \xrightarrow{\mathsf{H}^{-}} R^{-}N_{\mathsf{H}}^{\mathsf{CH}_{3}}$$
(4)

By adapting this synthetic protocol, the azide-containing intermediates used for the synthesis of **2b** could be utilized for the synthesis of **1a**. In terms of the timing of the transformation ($-N_3 \rightarrow -NHMe$), there were three optional stages (Scheme 18),^[53] stage 1: advanced intermediate **70**; stage 2: tetracyclic glycoside **69**; stage 3: glycosyl donor **51**.

We examined these three possibilities and stage 2 turned out to be the best choice, as described below. stage 1 proved to be inappropriate because the attempted conversion of **70** only produced an intractable mixture of multiple products, presumably owing to side reactions at the quinone functionality.

On the other hand, stage 3 revealed a different problem. Azide sugar **51** was successfully converted into monomethylamine **74** and subsequently to trifluoroacetylated glycosyl fluoride **75**. However, glycosidation of **75** with acceptor **68** gave exclusively the undesired α -glycoside **76** owing to the remote participation of the trifluoroacetamido group.^[54]

Fortunately, stage 2 was a viable one: Glycoside **69** in CH_2Cl_2 was successively treated with Me_3P , $(HCHO)_n$, and $NaBH_4$ to give monomethylamine **73** in a yield of 75% (see Scheme 18), which was successfully converted into the target **1a** (see below).

Scheme 19 illustrates the final stages en route to target **1a**. The amino group in **73** was masked with a trifluoroacetyl group to give β -**76** in a yield of 90%. After oxidation of β -**76** with CAN, construction of the E ring was achieved by the standard protocol (see Scheme 11) to give pentacycle **77** in a yield of 75% (three steps). After hydrogenolysis of **77** to detach the three benzyl groups, final removal of the five acyl groups (including a trifluoroacetyl group) and hydroly-

9804



Scheme 18. Three optional stages for the conversion of azide into the *N*-monomethylamine derivative (unless otherwise noted, the reactions were performed at ambient temperatures). Reagents and conditions: a) Me₃P, 1.5 h, (HCHO)_n, CH₂Cl₂, 15.5 h, NaB(CN)H₃, MeOH, 0°C, 10 min, 60% (contaminated with unidentified by-products); b) Me₃P, 1.5 h, (HCHO)_n, CH₂Cl₂, 11 h, NaBH₄, MeOH, 0°C, 10 min, 75%; c) Me₃P, 1.5 h, (HCHO)_n, CH₂Cl₂, 13 h, NaBH₄, MeOH, 0°C, 10 min, 78%; d) (CF₃CO)₂O, pyridine, CH₂Cl₂, 0°C, 30 min, 91%; e) amide **68**, Cp₂HfCl₂/AgOTf (1:2), MS4A, CH₂Cl₂, -25°C, 11 h, 93%.



Scheme 19. Synthesis of pradimicin A (unless otherwise noted, the reactions were performed at ambient temperatures). Reagents and conditions: a) (CF₃CO)₂O, pyridine, CH₂Cl₂, 20 min, 0°C, 90%; b) CAN, H₂O, CH₃CN, 20 min; c) diene **21**, THF, 20 min then SiO₂, 14 h; d) K₂CO₃, CH₂Cl₂, THF, 2 h, 40°C, 75% (3 steps); e) H₂, Pd/C, MeOH, DMF, 30 min; f) 0.5 M aq. NaOH, 24 h, then 2 M aq. HCl, 61% (2 steps), for purification, see text.

sis of the methyl ester were achieved by treatment with 0.5 M aqueous NaOH without MeOH (cf. Scheme 17).^[55] After acidification with 2 M aqueous HCl (pH 3.5), the filtrate was purified by successive column chromatography on Cosmosil (C18, CH₃CN/H₂O=3:7, phosphate buffer pH 3) and Sephadex LH-20 (DMF) to afford pradimicin A as a

DMF solvate that was dissolved in MeOH and the pH was adjusted to 3.5 by the addition of 2 M aqueous HCl. The reprecipitation protocol described above (see above) gave a red powder of pradimicin A hydrochloride (**1a**·HCl) in a yield of 61%^[56] which was identical to an authentic sample (m.p. 194–197°C, $[\alpha]_D^{26} = +280$ (c = 0.0100 in 1.00 M aq. HCl); lit.:^[1d] m.p. 193–195°C, $[\alpha]_D^{30} = +287$ (c = 0.01 in 1.0 M aq. HCl)).

Benanomicin A (2a): The last target was benanomicin A (2a), which has a neutral sugar (Scheme 20). Fluoride 80



Scheme 20. Synthesis of benanomicin A (unless otherwise noted, the reactions were performed at ambient temperatures). Reagents and conditions: a) **80**, Cp₂HfCl₂/AgOTf (1:2), MS4A, CH₂Cl₂, -30° C, 17 h, 7% for the α anomer and 74% for the β anomer; b) CAN, H₂O, CH₃CN, 20 min; c) diene **21**, THF, 30 min then SiO₂, 14 h; d) K₂CO₃, CH₂Cl₂, THF, 40°C, 2 h, 66% (3 steps); e) H₂, Pd/C, MeOH, DMF, 50 min; f) 0.5 M aq. NaOH, 15 h, then 1 M aq. HCl, 68% (2 steps).

was used as the glycosyl donor.^[57] The glycosidation reaction was initially attempted under exactly the same conditions as those used for the synthesis of 69. Unexpectedly, however, the reactivity of this neutral glycosyl fluoride 80 was lower than that of the azide-containing glycosyl donor 51, and therefore, no reaction occurred at -36°C. Fortunately, a subtle difference in the reaction temperature proved decisive and the reaction proceeded smoothly at -30°C. Fluoride 80 and alcohol 68 were treated with Cp₂HfCl₂ and AgOTf (1:2 ratio) in the presence of MS4A in CH₂Cl₂ at -78 °C and stirred at -30 °C for 17 h to give β -glycoside 78 (74%) along with its α anomer (7%). After isolation and oxidation of β -78 by CAN, the resulting unstable chloroquinone was immediately subjected to the previously described E-ring annulation protocol (see Scheme 11) to give pentacycle 79 in a fully regiocontrolled manner (66% yield over three steps). After hydrogenolysis of 79 in a mixed solvent

www.chemeurj.org

- 9805

(MeOH/DMF=3:1) for 50 min,^[49] removal of the five acyl groups and hydrolysis of the methyl ester were achieved by treatment with 0.5 m aqueous NaOH for 15 h. After acidification with 2 m aqueous HCl (pH 3.5), the residue was purified by successive column chromatography on Diaion HP-20 (H₂O/MeOH=1:0 to 0:1, gradient elution) and Sephadex LH-20 (DMF). After removal of the DMF, the residue was dissolved in a small amount of DMSO to which CHCl₃ was added. The resulting precipitates were collected by filtration to afford benanomicin A (**2a**) as a red powder (68 % yield). The synthetic sample showed spectroscopic properties identical to those of the natural product (m.p. 242–244 °C (decomp); lit.:^[1b] m.p. > 220 °C).

Conclusion

A general synthetic route to benanomicin-pradimicin antibiotics has been developed that is based on three significant findings: 1) Diastereoselective biaryl lactone ring-opening, 2) correlation of the biaryl educt configuration to pinacol product stereochemistry, and 3) semi-pinacol cyclization for the discrimination of the diol moiety in the B ring. By using this synthetic strategy, it should be possible to synthesize various natural/unnatural BpA congeners.

Experimental Section

General methods: All experiments involving air- and moisture-sensitive compounds were conducted under an atmosphere of dry argon. Dichloromethane was successively distilled from P2O5 and CaH2 and stored over 4 Å molecular sieves. For TLC analyses, Merck precoated plates (silica gel 60 F254, Art 5715, 0.25 mm) were used. For flash column chromatography, silica gel 60N (Spherical, neutral, 23-210 µm) from Kanto Chemicals was used. Preparative TLC (PTLC) was performed on Merck silica gel 60 PF₂₅₄ (Art 7747). Melting point (m.p.) determinations were performed by using a Yanako MP-S3 or MP-500 instrument and are uncorrected; ¹H and ¹³C NMR spectra were recorded by using a JEOL JNM EX-270 (270 MHz), a JNM AL-400 (400 MHz), a JNM lambda-400 (400 MHz), or a Bruker DRX-500 (500 MHz) spectrometer in the solvent indicated; chemical shifts (δ) are given in ppm relative to tetramethylsilane and coupling constants (J) are reported in Hz. Multiplicities are described by the following abbreviations s = singlet, d = doublet, t = triplet, q = quartet, sept=septet, m=multiplet, br=broad. Infrared (IR) spectra were recorded by using a Jasco IR-Report-100 or a Perkin-Elmer Spectrum 100 spectrometer. Attenuated total reflectance Fourier-transform infrared (ATR-FTIR) spectra were recorded by using a Perkin–Elmer 1600 FTIR spectrometer. In situ IR spectra were recorded by using a REMSPEC mid-IR fiber-Optic probe with an MCT detector. Ten scans were acquired per spectrum at a resolution of 4 cm⁻¹ by using the Time Base software package (Perkin-Elmer). Optical rotations ($[\alpha]_D$) were measured by using a Jasco DIP-1000 polarimeter. UV spectra were recorded by using a Hitachi 300 spectrometer. CD spectra were recorded by using a Jasco J500C spectropolarimeter. High-performance liquid chromatography (HPLC) analyses were performed by using a Jasco 880PU instrument with UV detection at 254 or 300 nm. Low-resolution mass spectra (LRMS) were obtained with a Shimadzu Kratos Kompact MALDI IIS spectrometer. High-resolution mass spectra (HRMS) were obtained with a JEOL JMS-700 spectrometer. Elementary analyses were performed on a Perkin-Elmer series II 2004 instrument. X-ray crystallographic data were recorded with a Rigaku RAXIS-RAPID diffractometer at the Tokyo Institute of Technology X-ray Laboratory.

First-generation total synthesis

Methyl 2-hydroxy-6-methyl-4-(trifluoromethylsulfonyloxy)benzoate: K₂CO₃ (4.55 g, 24.7 mmol) and PhNTf₂ (5.88 g, 16.5 mmol) were added to a solution of 8 (3.00 g, 16.5 mmol) in acetone (30 mL) at 0°C. After stirring for 24 h at room temperature, the reaction mixture was filtered through a Celite pad (washed with methanol). The filtrate was concentrated in vacuo, the mixture was diluted with H2O, and extracted with EtOAc $(\times 3)$. The combined organic extracts were washed with brine. dried (Mg₂SO₄), and concentrated in vacuo. Flash column chromatography (silica gel, hexane/EtOAc=95:5 to 80:20, gradient elution) gave the undesired regioisomer methyl 4-hydroxy-2-methyl-6-(trifluoromethylsulfonyloxy)benzoate as a white crystalline solid (1.09 g, 21 %) and somewhat impure methyl 2-hydroxy-6-methyl-4-(trifluoromethylsulfonyloxy)benzoate contaminated with PhNHTf, which was further purified by washing with saturated aqueous K2CO3, followed by extraction with EtOAc to afford a pure form of methyl 2-hydroxy-6-methyl-4-(trifluoromethylsulfonyloxy)benzoate as a white amorphous solid (3.63 g, 70%). M.p. 52–53 °C (Et₂O); ¹H NMR (400 MHz, CDCl₃): $\delta = 2.58$ (s, 3H), 3.99 (s. 3H), 6.65 (d. J=2.6 Hz, 1H), 6.78 (d. J=2.6 Hz, 1H), 11.6 ppm (s. 1 H; OH); ¹³C NMR (100 Hz, CDCl₃): $\delta = 24.2$, 52.6, 108.4, 115.4, 118.6 (q, ${}^{1}J_{CF}$ =317 Hz), 128.6, 114.5, 152.4, 164.4, 171.2 ppm; IR (film): $\tilde{\nu}$ = 3090, 3050, 2990, 2970, 1660, 1610, 1590, 1510, 1460, 1440, 1410, 1380, 1350, 1310, 1270, 1240, 1220, 1200, 1140, 1130, 1060, 1040, 970, 950, 880, 850, 810, 770 cm⁻¹; elemental analysis calcd (%) for C₁₀H₉F₃O₆S: C 38.22, H 2.89, S 10.20; found: C 38.03, H 2.90, S 10.11. Methyl 4-hydroxy-2methyl-6-(trifluoromethylsulfonyloxy)benzoate: M.p. 88-89°C (hexane and Et₂O); ¹H NMR (400 MHz, CDCl₃): $\delta = 2.41$ (s, 3H), 3.93 (s, 3H), 5.77 (s, 1H; OH), 6.65 (d, *J*=2.4 Hz, 1H), 6.71 ppm (d, *J*=2.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 20.7, 52.7, 107.1, 117.6, 118.2, 118.4$ (q, ${}^{1}J_{CF}$ =318 Hz), 141.9, 148.1, 158.2, 166.2 ppm; IR (KBr): $\tilde{\nu}$ =3380, 2950, 1700, 1610, 1570, 1490, 1450, 1410, 1300, 1240, 1210, 1120, 1100, 1040, 970, 940, 850, 810, 780, 760, 700, 680 cm⁻¹; elemental analysis calcd (%) for C10H9F3O6S (314.24): C 38.22, H 2.89, S 10.20; found: C 38.25, H 3.02, S 10.05.

 $\label{eq:2-methoxy} 2-methoxy-6-methyl-4-(trifluoromethylsulfonyloxy) ben-$ Methyl zoate (9): iPr₂NEt (33 mL, 0.191 mol) and a solution of MOMCl (11.6 mL, 0.152 mol) in CH₂Cl₂ (80 mL) were added over 1 h to a solution of methyl 2-hydroxy-6-methyl-4-(trifluoromethylsulfonyloxy)benzoate (40.0 g, 0.127 mol) in CH2Cl2 (100 mL) at 0°C. After the reaction mixture had been stirred for 5.5 h at RT, the reaction was stopped by adding pH 7 phosphate buffer and then the mixture was extracted with CH_2Cl_2 (×3). The combined organic extracts were washed with 2M aq. HCl, saturated aqueous NaHCO₃, and brine, dried (Na₂SO₄), and concentrated in vacuo. After adding Et2O and hexane to the residue, the precipitate was collected by filtration to afford 9 (1st crop: 25.1 g, 55%; 2nd crop: 10.7 g, 23%) as a white solid. The filtrate was concentrated in vacuo and the residue was purified by flash column chromatography (silica gel, hexane/EtOAc=86:14) to afford 9 (9.83 g, 22 %, total yield: quant) as a white solid. M.p. 41-42 °C (hexane and Et₂O); ¹H NMR (400 MHz, CDCl₃): $\delta = 2.33$ (s, 3H), 3.47 (s, 3H), 3.93 (s, 3H), 5.18 (s, 2 H), 6.79 (d, J = 2.0 Hz, 1 H), 6.95 ppm (d, J = 2.0 Hz, 1 H); ¹³C NMR (100 Hz, CDCl₃): $\delta = 19.4$, 52.4, 56.3, 94.9, 106.1, 115.9, 118.9 (q, ${}^{1}J_{C,F} =$ 319 Hz), 124.8, 138.7, 150.0, 155.1, 167.2 ppm; IR (neat): $\tilde{\nu} = 3460$, 3100, 2960, 2830, 2080, 1970, 1730, 1590, 1480, 1430, 1280, 1210, 1140, 1050, 1000, 970, 930, 860, 840, 760 cm⁻¹; elemental analysis calcd (%) for C₁₂H₁₃O₇F₃S: C 40.23, H 3.66, S 8.95; found: C 40.16, H 3.73, S 8.90.

1-Methyl 4-phenyl 2-methoxymethoxy-6-methylterephthalate (**10**): A mixture of **9** (2.37 g, 6.62 mmol), Pd(OAc)₂ (75.6 mg, 0.337 mmol), 1,1'-bis(diphenylphosphino)ferrocene (181 mg, 0.326 mmol), Et₃N (2.11 g, 20.9 mmol), and PhOH (3.04 g, 32.3 mmol) in DMF (13 mL) was bubbled with CO gas for 10 min. The reaction mixture was stirred for 1.5 h at 60 °C and then for 1.5 h at 80 °C. After cooling to RT, the reaction mixture was diluted with water and extracted with Et₂O (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=84:16) to afford **10** as a white solid (2.18 g, quant). M.p. 71.5–72.0 °C (hexane and EtOAc); ¹H NMR (400 MHz, CDCl₃): δ =2.37 (s, 3H), 3.49 (s, 3H), 3.95 (s, 3H), 5.25 (s, 2H), 7.27 (t,

 $J=7.8~{\rm Hz}, 1~{\rm H}), 7.27~({\rm d}, J=7.8~{\rm Hz}, 2~{\rm H}), 7.43~({\rm t}, J=7.8~{\rm Hz}, 2~{\rm H}), 7.72~({\rm s}, 1~{\rm H}), 7.77~{\rm ppm}~({\rm s}, 1~{\rm H}); {\rm ^{13}C}~{\rm NMR}~(100~{\rm MHz}, {\rm CDCl}_3); \delta=19.2, 52.4, 56.4, 94.7, 113.5, 121.7, 125.3, 126.0, 129.5, 129.6, 131.2, 136.9, 150.8, 153.9, 164.5, 167.8~{\rm ppm};~{\rm IR}~({\rm KBr}); ~\tilde{\nu}=2950, 2830, 1730, 1590, 1490, 1430, 1400, 1320, 1300, 1270, 1200, 1160, 1120, 1100, 1080, 1050, 1000, 940, 930, 840, 790, 760, 730~{\rm cm}^{-1};$ elemental analysis calcd (%) for ${\rm C}_{18}{\rm H}_{18}{\rm O}_6$: C 65.45, H 5.49; found: C 65.41, H 5.57.

4-hydroxymethyl-2-methoxymethoxy-6-methylbenzoate Methvl (11): LiBH₄ (1.62 g, 74.5 mmol) was added to a solution of 10 (24.6 g, 74.5 mmol) in THF (260 mL) at 0°C. The reaction temperature was allowed to rise to RT and the mixture was stirred for 24 h. The reaction was stopped by adding AcOH and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=1:1) to afford 11 as a colorless oil (17.5 g, 98%). ¹H NMR (400 MHz, CDCl₃): $\delta = 2.28$ (s, 3H), 2.31 (t, J=6.0 Hz, 1H; OH), 3.45 (s, 3H), 3.91 (s, 3H), 4.61 (d, J = 6.0 Hz, 2 H), 5.16 (s, 2 H), 6.83 (s, 1 H), 6.96 ppm (s, 1 H); ¹³C NMR $(100 \text{ MHz}, \text{ CDCl}_3)$: $\delta = 19.3, 52.2, 56.2, 64.7, 94.6, 110.4, 121.7, 123.8,$ 136.8, 143.7, 154.1, 168.7 ppm; IR (film): $\tilde{v} = 3430, 2950, 2830, 1740, 1720,$ 1610, 1580, 1440, 1400, 1360, 1280, 1210, 1190, 1150, 1090, 1050, 1000, 930, 850, 820, 780, 750 cm $^{-1}\!;$ elemental analysis calcd (%) for $C_{12}H_{16}O_5\!:$ C 59.99, H 6.71; found: C 59.87, H 6.94.

Methyl 2-methoxymethoxy-4-(methoxymethoxy)methyl-6-methylbenzoate (12): iPr₂NEt (17.0 mL, 97.2 mmol) and MOMCl (7.2 mL, 95 mmol) were added to a solution of 11 (7.82 g, 32.5 mmol) in CH₂Cl₂ (65 mL) at 0°C. The reaction mixture was allowed to warm to RT and stirred for 3 h. The reaction was stopped by adding water and the mixture was extracted with EtOAc (\times 3). The combined organic extracts were washed with 2M aq. HCl, saturated aqueous NaHCO3, and brine, dried (Na2SO4), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=8:2) to afford 12 as a colorless oil (8.11 g, 88 %). ¹H NMR (400 MHz, CDCl₃): $\delta = 2.29 \text{ (s}, 3 \text{ H}), 3.41 \text{ (s}, 3 \text{ H}),$ 3.47 (s, 3H), 3.91 (s, 3H), 4.54 (s, 2H), 4.69 (s, 2H), 5.18 (s, 2H), 6.86 (brs, 1H), 6.98 ppm (brs, 1H); 13 C NMR (100 MHz, CDCl₃): $\delta = 19.3$, 52.1, 55.4, 56.2, 68.7, 94.7, 95.7, 111.4, 122.7, 124.1, 136.7, 140.5, 154.1, 168.6 ppm; IR (neat): $\tilde{v} = 3620, 2950, 2830, 2070, 1970, 1900, 1730, 1620,$ 1580, 1560, 1440, 1400, 1370, 1320, 1270, 1210, 1190, 1150, 1090, 1050, 1010, 950, 920, 850, 820 cm⁻¹; elemental analysis calcd (%) for $C_{14}H_{20}O_6$: C 59.14, H 7.09; found: C 58.92, H 7.26.

Methyl 2-hydroxy-4-(methoxymethoxy)methyl-6-methylbenzoate (13): Trifluoroacetic acid (1.2 mL, 16 mmol) was added to a solution of 12 (1.45 g, 5.09 mmol) in CH₂Cl₂ (25 mL) at 0°C. After stirring for 3 h, the reaction was stopped by adding water and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with saturated aqueous NaHCO₃, water, and brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=9:1) to afford phenol 13 as a colorless oil (1.22 g, 99%). ¹H NMR (400 MHz, CDCl₃): δ =2.54 (s, 3H), 3.41 (s, 3H), 3.95 (s, 3H), 4.52 (s, 2H), 4.71 (s, 2H), 6.70 (s, 1H), 6.84 (s, 1H), 11.33 ppm (s, 1H; OH); ¹³C NMR (100 MHz, CDCl₃): δ =24.1, 52.1, 55.5, 68.2, 95.9, 111.3, 114.1, 121.7, 141.5, 144.8, 163.1, 172.1 ppm; IR (neat): $\tilde{\nu}$ =2950, 2890, 1730, 1660, 1620, 1570, 1440, 1420, 1380, 1360, 1320, 1260, 1210, 1150, 1100, 1050, 1010, 990, 940, 920, 850, 810, 750 cm⁻¹; elemental analysis calcd (%) for C₁₂H₁₆O₃: C 59.99, H 6.71; found: C 59.97, H 6.79.

Methyl 2-hydroxy-3-iodo-4-(methoxymethoxy)methyl-6-methylbenzoate (14): NaHCO₃ (5.23 g, 62.7 mmol) and BnMe₃N⁺ICl₂⁻ (10.8 mL, 30.9 mmol) were added to a solution of 13 (7.48 g, 31.1 mmol) in CH₂Cl₂/ MeOH (5:2, 310 mL) at 0 °C. The reaction mixture was stirred for 10 h at 0 °C. The reaction was stopped by adding 10% aqueous Na₂S₂O₃ and the mixture was extracted with CH₂Cl₂ (×3). The combined organic extracts were washed with 10% aqueous Na₂S₂O₃ and brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by recrystallization from EtOH to afford 14 as a white solid (1st crop: 8.00 g, 70%; 2nd crop: 0.75 g, 6.5%; 3rd crop: 0.75 g, 6.5%; total yield: 83%). M.p. 67.5–68.0°C (EtOH); ¹H NMR (400 MHz, CDCl₃): δ =2.54 (s, 3H), 3.45 (s, 3H), 3.99 (s, 3H), 4.59 (s, 2H), 4.79 (s, 2H), 6.94 (s, 1H), 12.36 ppm (s, 1H; OH); ¹³C NMR (100 MHz, CDCl₃): δ =2.39, 52.6, 55.7, 73.4, 85.6, 96.4, 111.3, 122.6, 141.5, 147.1, 160.9, 171.8 ppm; IR (KBr): $\tilde{\nu}$ =2960, 2890, 1660, 1600, 1540, 1450, 1390, 1370, 1350, 1290, 1250, 1200, 1150, 1110, 1060, 1000, 960, 940, 910, 860, 810, 770, 710 cm⁻¹; elemental analysis calcd (%) for C₁₂H₁₅O₅I: C 39.36, H 4.13; found: C 39.47, H 4.34.

4-tert-Butyl 1-ethyl 2-(2,5-dimethoxybenzylidene)succinate (**16**): Phosphonate **19** (4.90 g, 14.5 mmol) in CH₃CN (15 mL), DBU (2.20 g, 1.44 mmol), and 2,5-dimethoxybenzaldehyde (2.00 g, 12.0 mmol) were added to a suspension of LiCl (0.827 g, 19.6 mmol) in CH₃CN (90 mL) at RT and the reaction mixture was stirred for 44 h. The reaction was stopped by adding water and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/Et₂O/CHCl₃=14:3:3) to afford the *E* (3.12 g, 73%) and *Z* isomers (156 mg, 3.7%) of **16** as yellow oils.

16 (*E* isomer): ¹H NMR (400 MHz, CDCl₃): δ =1.32 (t, *J*=7.1 Hz, 3 H), 1.42 (s, 9H), 3.39 (s, 2H), 3.73 (s, 3H), 3.77 (s, 3H), 4.25 (q, *J*=7.1 Hz, 2H), 6.81 (d, *J*=9.0 Hz, 1H), 6.85 (dd, *J*=2.7, 9.0 Hz, 1H), 6.89 (d, *J*= 2.7 Hz, 1H), 7.91 ppm (s, 1H); ¹³C NMR (100 MHz, CDCl₃): δ =14.2, 27.9, 35.3, 55.6, 55.9, 60.8, 80.7, 111.6, 114.8, 115.5, 124.7, 126.9, 137.4, 151.7, 153.1, 167.3, 170.5 ppm; IR (neat): $\tilde{\nu}$ =2730, 2725, 1720, 1710, 1640, 1580, 1500, 1460 cm⁻¹; elemental analysis calcd (%) for C₁₉H₂₆O₆: C 65.13, H 7.48; found: C 65.43, H 7.50.

16 (*Z* isomer): ¹H NMR (400 MHz, CDCl₃): δ =1.08 (t, *J*=7.1 Hz, 3 H), 1.44 (s, 9H), 3.38 (s, 2H), 3.73 (s, 3H), 3.75 (s, 3H), 4.09 (q, *J*=7.1 Hz, 2H), 6.74–6.81 (m, 3H), 6.86 ppm (s, 1H); ¹³C NMR (100 MHz, CDCl₃): δ =13.7, 28.0, 42.0, 55.7, 56.0, 60.4, 81.0, 111.4, 114.4, 115.6, 125.8, 128.0, 134.5, 151.2, 152.9, 167.7, 170.0 ppm; IR (neat): $\tilde{\nu}$ =2730, 2725, 1720, 1490, 1220 1160 cm⁻¹; elemental analysis calcd (%) for C₁₉H₂₆O₆: C 65.13, H 7.48; found: C 65.43, H 7.74.

3-(2,5-dimethoxybenzylidene)succinic acid monoethyl ester (**17**): Trifluoroacetic acid (5.2 mL) was added to a solution of (*E*)-**16** (2.99 g, 8.55 mmol) in water (0.58 mL). After stirring for 3.5 h, volatile materials were removed by azeotropic distillation with toluene (×3). The resulting solid was filtered to afford **17** as a white powder (2.13 g, 85%). The filtrate was purified by flash column chromatography (silica gel, hexane/ EtOAc=1:1 then MeOH/CHCl₃=1:9) to afford **17** as a white solid (0.14 g, 5%; total yield: 90%). ¹H NMR (270 MHz, CDCl₃): δ =1.35 (t, *J*=7.1 Hz, 3H), 3.51 (s, 2H), 3.77 (s, 3H), 3.80 (s, 3H), 4.31 (q, *J*= 7.1 Hz, 2H), 6.82–6.82 (m, 3H), 7.99 (s, 1H), 10.6 ppm (s, 1H).

Ethyl 4-acetoxy-5,8-dimethoxy-2-naphthoate (18a): NaOAc (0.800 g, 9.75 mmol) was added to a solution of 17 (2.10 g, 7.13 mmol) in acetic anhydride (11.5 mL) at RT and the reaction mixture was stirred for 30 min at 140°C. After cooling to RT, the resulting precipitate was collected by filtration to afford 18a as a white powder (1.99 g, 88%). The filtrate was concentrated in vacuo. The residue was azeotroped with toluene (×2) to remove volatile materials and recrystallized (hexane/EtOAc) to afford 18a (0.162 g, 7%). The mother liquor was extracted with EtOAc (×3) and the combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=7:3) to afford 18a as a white solid (0.0937 g, 11%; total yield: 99%). ¹H NMR (270 MHz, CDCl₃): δ =1.43 (t, J=7.1 Hz, 3H), 2.38 (s, 3H), 3.89 (s, 3H), 3.97 (s, 3H), 4.42 (q, J=7.1 Hz, 2H), 6.77 (d, J=8.6 Hz, 1H), 6.87 (d, J=8.6 Hz, 1H), 7.69 (d, J=1.7 Hz, 1H), 8.90 ppm (d, J=1.7 Hz, 1H).

Ethyl 4-hydroxy-5,8-dimethoxy-2-naphthoate (18d): K₂CO₃ (56.8 g, 0.411 mol) was added to a solution of 18a (26.3 g, 0.0826 mol) in EtOH (410 mL) at RT. After stirring for 1 h at 70 °C, the reaction was extracted with EtOAc (×3) and the combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by recrystallization (hexane/EtOAc) to afford 18d as a white powder (20.1 g, 89%). The mother liquor was concentrated in vacuo and the residue was purified by flash column chromatography (silica gel, hexane/EtOAc=65:35) to afford 18d as a white powder (2.03 g, 9%; total yield: 98%). M.p. 120.0–120.2 °C (hexane and EtOAc); ¹H NMR (400 MHz, CDCl₃): δ =1.43 (t, *J*=7.2 Hz, 3 H), 3.97 (s, 3 H), 4.03 (s, 3 H), 4.41 (q, *J*=7.2 Hz, 2H), 6.69 (d, *J*=8.4 Hz, 1H), 6.79 (d, *J*=8.4 Hz, 1H), 7.49 (d, *J*=1.2 Hz, 1H), 8.46 (d, *J*=1.2 Hz, 1H), 9.49 ppm (s, 1H; OH); ¹³C NMR (100 MHz, CDCl₃): δ =14.4, 55.7, 56.3, 61.0, 103.5, 105.7, 110.5, 115.7,

Chem. Eur. J. 2007, 13, 9791-9823

© 2007 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

117.5, 127.4, 128.8, 149.5, 150.9, 154.4, 166.5 ppm; IR (KBr): $\tilde{\nu}{=}3370,$ 1710, 1620, 1520, 1450, 1390, 1290, 1250, 1230, 1110, 1050, 1030, 810, 770 cm^{-1}; elemental analysis calcd (%) for $C_{15}H_{16}O_5$: C 65.21, H 5.84; found: C 65.33, H 5.90.

Ethyl 5,8-dimethoxy-4-methoxymethoxy-2-naphthoate (18b): Compound 18d (20.9 g, 0.0766 mmol) and MOMCl (9.33 g, 0.116 mol) were added to a suspension of NaH (60% dispersion in mineral oil, washed with hexane (4.02 g, 0.100 mol)) in DMF (240 mL) at 0°C. After stirring for 20 min, the reaction was stopped by adding water and the mixture was extracted with Et_2O (×3). The combined organic extracts were washed with brine, dried (MgSO₄), and concentrated in vacuo. After adding Et₂O and hexane to the residue, the precipitate was collected by filtration to afford 18b as a white solid (23.8 g, 99%). M.p. 78.7-79.2 °C (hexane and EtOAc); ¹H NMR (400 MHz, CDCl₃): $\delta = 1.44$ (t, J = 7.2 Hz, 3H), 3.62 (s, 3H), 3.92 (s, 3H), 3.98 (s, 3H), 4.43 (q, J=7.2 Hz, 2H), 5.41 (s, 2H), 6.78 (d, J = 8.4 Hz, 1 H), 6.90 (d, J = 8.4 Hz, 1 H), 7.66 (d, J = 1.6 Hz, 1 H), 8.70 ppm (d, J = 1.6 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 14.5$, 55.8, 56.6, 57.3, 61.1, 96.7, 104.6, 109.5, 112.8, 119.4, 121.4, 127.5, 127.9, 150.2, 150.4, 153.6, 166.4 ppm; IR (KBr): $\tilde{\nu} = 2980$, 1710, 1600, 1520, 1470, 1380, 1280, 1150, 1090, 1040, 920, 800, 760, 730, 720, 660 cm⁻¹; elemental analysis calcd (%) for C₁₇H₂₀O₆: C 63.74, H 6.29; found: C 63.53, H 6.45.

Ethyl 8-methoxymethoxy-1,4-naphthoquinone-6-carboxylate (20b): A solution of CAN (20.1 g, 36.6 mmol) in water (63 mL) was added to a solution of 18b (5.80 g, 18.3 mmol) in CH₃CN (183 mL) at 0 °C. After stirring for 10 min, the reaction was stopped by adding water, the precipitate was collected by filtration, and washed with Et2O. After adding water and stirring for 10 min, precipitate was collected by filtration to afford 20b (4.36 g, 83 %) as a yellow solid. The filtrate was extracted with EtOAc (× 3). The combined organic extracts were washed with brine, dried (MgSO₄), and concentrated in vacuo. The precipitate was collected by filtration and washed with Et₂O to afford **20b** as a yellow solid (0.683 g, 13%; total yield: 96%). M.p. 107-108°C (hexane and EtOAc); ¹H NMR (400 MHz, CDCl₃): $\delta = 1.44$ (t, J = 7.2 Hz, 3H), 3.56 (s, 3H), 4.45 (q, J =7.2 Hz, 2H), 5.42 (s, 2H), 6.92 (d, J=10.4 Hz, 1H), 6.90 (d, J=10.4 Hz, 1H), 8.16 (d, J=1.6 Hz, 1H), 8.41 ppm (d, J=1.6 Hz, 1H); ¹³C NMR $(100 \text{ MHz}, \text{ CDCl}_3): \delta = 14.3, 56.8, 62.0, 95.1, 122.4, 123.0, 134.0, 135.9,$ 137.0, 140.7, 157.0, 164.5, 183.5, 184.1 ppm; IR (KBr): $\tilde{\nu} = 3680$ 3630, 1720, 1660, 1610, 1600, 1560, 1550, 1430, 1370, 1340, 1320, 1270, 1210, 1170, 1110, 1090, 1000, 920, 880, 850, 770 cm⁻¹; elemental analysis calcd (%) for $\rm C_{15}H_{14}O_6$: C 62.06, H 4.86; found: C 61.84, H 4.95.

Ethyl 8-hydroxy-6-methoxy-4-methoxymethoxy-9,10-anthraquinone-2-carboxylate (22 b): Diene 21 (3.47 g, 10.5 mmol) was added to a solution of 20b (2.50 g, 8.73 mmol) in THF (87 mL) at 0°C. After stirring the reaction mixture for 7 h at -78 °C, nBu₄NF (1.0 M in THF, 13 mL, 13 mmol) was added and the mixture bubbled with O_2 for 5 min at RT. The mixture was poured into water (150 mL) and stirred for 15 min. The resulting precipitate was collected by filtration to afford 22b as a yellow solid (2.11 g, 84%). M.p. 195.0-195.4°C (hexane and EtOAc); ¹H NMR (400 MHz, $CDCl_3$): $\delta = 1.45$ (t, J = 7.2 Hz, 3H), 3.59 (s, 3H), 3.94 (s, 3H), 4.46 (q, J =7.2 Hz, 2H), 5.45 (s, 2H), 6.69 (d, J=2.4 Hz, 1H), 7.32 (d, J=2.4 Hz, 1H), 8.16 (d, J=1.5 Hz, 1H), 8.62 (d, J=1.5 Hz, 1H), 12.71 ppm (s, 1H; OH); ¹³C NMR (100 MHz, CDCl₃): $\delta = 14.3$, 56.0, 56.9, 62.1, 95.2, 106.0, 107.7, 110.2, 121.3, 122.5, 124.8, 136.0, 136.1, 136.4, 158.0, 164.7, 165.1, 166.7, 181.2, 185.5 ppm; IR (KBr): $\tilde{\nu} = 2380, 2360, 1730, 1680, 1650, 1630,$ 1590, 1560, 1430, 1400, 1300, 1260, 1150, 1010, 960, 930, 820 cm⁻¹; elemental analysis calcd (%) for $C_{20}H_{18}O_8$: C 62.17, H 4.70; found: C 62.11, H 4.78

Ethyl 8-benzyloxy-6-methoxy-4-methoxymethoxy-9,10-anthraquinone-2carboxylate (24): Cs₂CO₃ (4.29 g, 13.2 mmol) and BnBr (1.44 mL, 11.6 mmol) were added to a solution of anthraquinone 22b (2.02 g, 5.27 mmol) in DMF (150 mL) at 0 °C. After stirring for 7.5 h at RT, the reaction was stopped by adding water and the mixture was extracted with CH₂Cl₂ (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The precipitate was collected by filtration and washed with Et₂O to afford 24 as a yellow powder (2.23 g, 89%). M.p. 179–180°C (hexane and EtOAc); ¹H NMR (500 MHz, CDCl₃): δ = 1.43 (t, J = 7.0 Hz, 3H), 3.59 (s, 3H), 3.92 (s, 3H), 4.43 (q, J = 7.0 Hz, 2H), 5.27 (s, 2H), 5.43 (s, 2H), 6.78 (s, 1H), 7.33 (t, *J*=7.6 Hz, 1H), 7.38 (s, 1H), 7.43 (t, *J*=7.6 Hz, 2H), 7.61 (d, *J*=7.6 Hz, 2H), 8.07 (s, 1H), 8.61 ppm (s, 1H); ¹³C NMR (125 MHz, CDCl₃): δ = 14.3, 55.8, 56.7, 61.8, 71.0, 95.3, 103.7, 105.7, 115.8, 120.9, 121.8, 124.3, 126.8, 127.9, 128.7, 136.00, 136.01, 137.6, 139.0, 157.1, 161.0, 164.8, 165.0, 179.9, 182.3 ppm; IR (KBr): $\tilde{\nu}$ =2460, 1710, 1670, 1650, 1600, 1560, 1460, 1390, 1310, 1240, 1150, 1110, 1030, 970, 930, 760 cm⁻¹; elemental analysis calcd (%) for C₂₇H₂₄O₈: C 68.06, H 5.08; found: C 67.96, H 5.31.

Ethvl 8-benzyloxy-6.9.10-trimethoxy-4-(methoxymethoxy)anthracene-2carboxylate (25): A solution of Na₂S₂O₄ (5.14 g, 29.4 mmoL) in degassed water (112 mL) was added to a suspension of anthraquinone 24 (1.40 g, 2.94 mmol) and nBu₄NBr (237 mg, 0.735 mmol) in degassed THF (50 mL). After stirring for 30 min at RT, 50 % aqueous KOH (9 mL) was added to the mixture, which was stirred for 30 min. After adding Me₂SO₄ (0.735 mL, 7.35 mmoL) and stirring for 1 h, the reaction was stopped by adding water and N,N-dimethyl-1,3-propanediamine (2 mL) at 0°C and the organic layer was extracted with CH_2Cl_2 (×3). The combined organic extracts were washed with brine, dried (Na2SO4), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=75:25) to afford 25 as a yellow amorphous solid (1.46 g, 98%). M.p. 144.5-145.4°C (hexane and EtOAc); ¹H NMR (400 MHz, CDCl₃): $\delta = 1.44$ (t, J = 7.2 Hz, 3H), 3.66 (s, 3H), 3.87 (s, 3H), 3.98 (s, 3H), 4.00 (s, 3H), 4.44 (q, J=7.2 Hz, 2H), 5.24 (s, 2H), 5.43 (s, 2H), 6.63 (d, J=1.6 Hz, 1H), 7.26 (d, J=1.6 Hz, 1H), 7.37 (t, J=7.5 Hz, 1 H), 7.45 (t, J=7.5 Hz, 2 H), 7.57 (d, J=1.4 Hz, 1 H), 7.62 (d, J=7.5 Hz, 2 H), 8.81 ppm (d, J = 1.4 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 14.9$, 55.8, 57.1, 61.4, 62.9, 64.6, 71.7, 92.5, 96.6, 101.4, 109.3, 116.6, 120.8, 122.0, 126.2, 126.3, 128.1, 128.5, 129.0, 131.4, 137.0, 146.7, 151.8, 153.1, 157.2, 158.9, 167.1 ppm; IR (KBr): $\tilde{\nu} = 2940$, 1700, 1620, 1540, 1410, 1360, 1310, 1260, 1220, 1150, 1110, 1060, 990, 830, 770, 750, 700 cm⁻¹; elemental analysis calcd (%) for C₂₉H₃₀O₈: C 68.76, H 5.77; found: C 68.46, H 5.98.

8-Benzyloxy-6,9,10-trimethoxy-4-(methoxymethoxy)anthracene-2-carboxylic acid (26): A 5M aq. KOH solution (133 mL) was added to a suspension of anthracene 25 (1.30 g, 2.57 mmoL) in EtOH (186 mL) and stirred for 1.5 h under reflux conditions. After cooling to RT, the reaction mixture was diluted with water and washed with $\mathrm{Et_2O}$ (×3). The aqueous layer was acidified with 2 M aq. HCl and extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na_2SO_4) , and concentrated in vacuo. The precipitate was collected by filtration and washed with Et₂O to afford **26** as a vellow powder (1.12 g, 91%). M.p. 198–199°C (hexane and EtOAc); ¹H NMR (400 MHz, CDCl₃): $\delta = 3.66$ (s, 3H), 3.88 (s, 3H), 3.99 (s, 3H), 4.00 (s, 3H), 5.26 (s, 2H), 5.44 (s, 2H), 6.64 (d, J=1.7 Hz, 1 H), 7.27 (d, J=1.7 Hz, 1 H), 7.38 (t, J=7.3 Hz, 1 H), 7.45 (t, J=7.3 Hz, 2H), 7.58 (s, 1H), 7.63 (d, J=7.3 Hz, 2H), 8.93 (s, 1H) ppm; IR (KBr): v=2370, 2340, 1680, 1640, 1560, 1460, 1420, 1360, 1330, 1250, 1160, 1060, 1030, 1000, 830 cm⁻¹; elemental analysis calcd (%) for C₂₇H₂₆O₈: C 67.77, H 5.48; found: C 68.07, H 5.30. This title compound was carried on to the subsequent experiment without further characterization.

2-Iodo-6-methoxycarbonyl-3-(methoxymethoxy)methyl-5-methylphenyl 8benzyloxy-6.9.10-trimethoxy-4-(methoxymethoxy)anthracene-2-carboxylate: Et₃N (140 mg, 1.38 mmol) and 2,4,6-trichlorobenzoyl chloride (459 mg, 1.88 mmol) were added to a solution of carboxylic acid 26 (600 mg, 1.25 mmol) in toluene (10 mL) at RT. After stirring for 3 h at RT, iodophenol 14 (689 mg, 1.88 mmol) and DMAP (460 mg, 3.77 mmol) was added to the reaction mixture and then stirred for 30 min. The reaction was stopped by adding water and the mixture was extracted with CH₂Cl₂ (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=70:30) to afford the title compound as a yellow amorphous solid (974 mg, 94%). ¹H NMR (500 MHz, CDCl₃): $\delta = 2.45$ (s, 3H), 3.45 (s, 3H), 3.67 (s, 3H), 3.71 (s, 3H), 3.90 (s, 3H), 4.00 (s, 3H), 4.03 (s, 3H), 4.64 (s, 2H), 4.80 (s, 2H), 5.26 (s, 2H), 5.46 (s, 2H), 6.65 (d, J=2.2 Hz, 1H), 7.28 (d, J= 2.2 Hz, 1 H), 7.32 (s, 1 H), 7.37 (t, J=7.4 Hz, 1 H), 7.45 (t, J=7.4 Hz, 2 H), 7.62 (d, J=7.4 Hz, 1 H), 7.64 (d, J=1.4 Hz, 1 H), 9.05 ppm (d, J=1.4 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃): $\delta = 20.0$, 52.3, 55.4, 55.7, 56.8, 62.6, 64.3, 71.3, 73.0, 91.9, 92.1, 96.2, 96.3, 101.0, 108.7, 116.3, 120.6, 123.4, 124.1, 125.7, 126.6, 127.3, 127.6, 127.8, 128.1, 128.6, 131.5, 136.4, 138.5,

9808 -

144.0, 146.4, 149.3, 151.8, 152.9, 156.9, 163.8, 166.0 ppm; IR (KBr): $\tilde{\nu}=$ 2930, 1740, 1690, 1650, 1620, 1560, 1540, 1450, 1410, 1360, 1260, 1210, 1170, 1060, 980 cm^{-1}; elemental analysis calcd (%) for $C_{39}H_{39}O_{12}I$: C 56.67, H 4.63; found: C 56.46, H 4.63.

2-Iodo-6-methoxycarbonyl-3-(methoxymethoxy)methyl-5-methylphenyl 8benzyloxy-4-hydroxy-6,9,10-trimethoxyanthracene-2-carboxylate (27): Trifluoroacetic acid (133 mg, 1.17 mmol) was added to a degassed solution 2-iodo-6-(methoxycarbonyl)-3-(methoxymethoxy)methyl-5-methylof phenyl 8-(benzyloxy)-6,9,10-trimethoxy-4-(methoxymethoxy)anthracene-2-carboxylate (320 mg, 0.387 mmol) in CH2Cl2 (6 mL) at 0 °C. After stirring for 2 h at RT, the reaction was stopped by adding saturated aqueous NaHCO3 and water, and then the mixture was extracted with CH2Cl2 (× 3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, benzene/EtOAc=9:1) to afford 27 as a yellow solid (264 mg, 87%). M.p. 187-188°C (hexane and EtOAc); ¹H NMR (500 MHz, CDCl₃): $\delta = 2.44$ (s, 3 H), 3.45 (s, 3 H), 3.70 (s, 3 H), 3.90 (s, 3 H), 4.01 (s, 3 H), 4.10 (s, 3 H), 4.63 (s, 2 H), 4.80 (s, 2 H), 5.26 (s, 2H), 6.63 (d, J=2.2 Hz, 1H), 7.01 (d, J=2.2 Hz, 1H), 7.32 (s, 1H), 7.38 (t, J=7.3 Hz, 1 H), 7.45 (dd, J=7.3, 7.6 Hz, 2 H), 7.53 (d, J=1.5 Hz, 1 H), 7.62 (d, J=7.6 Hz, 2H), 8.85 (d, J=1.5 Hz, 1H), 9.81 ppm (s, 1H; OH); ^{13}C NMR (125 MHz, CDCl₃): $\delta\!=\!20.0,\;52.4,\;55.5,\;55.7,\;63.2,\;64.3,\;71.4,$ 73.1, 90.7, 91.9, 96.3, 100.9, 107.8, 116.4, 118.6, 120.2, 125.3, 125.6, 126.6, 127.7, 127.8, 128.2, 128.7, 128.9, 136.2, 138.6, 144.0, 145.4, 149.4, 152.8, 153.1, 157.3, 159.1, 163.9, 166.1 ppm; IR (KBr): v=3320, 2950, 1740, 1650, 1620, 1560, 1540, 1450, 1410, 1370, 1260, 1190, 1060, 960, 820, 740, 700 cm⁻¹; elemental analysis calcd (%) for $C_{37}H_{35}O_{11}I$: C 56.79, H 4.51; found: C 56.76, H 4.56.

Methyl 9-benzyloxy-14-hydroxy-8,11,13-trimethoxy-1-(methoxymethoxy)methyl-3-methyl-6-oxo-6H-anthra[2,3-c]benzopyran-4-carboxylate (28): PPh₃ (41.4 mg, 0.158 mmol) and 27 (99.8 mg, 0.128 mmol) were added to a suspension of Pd(OCOCF₃)₂ (99.8 mg, 0.128 mmol), sodium pivalate (44.6 mg, 0.359 mmol), and $P(t-Bu)_3$ (35 μ L, 0.15 mmol) in DMA (12.8 mL). After stirring at 110°C for 25 min, the reaction mixture was cooled to 0°C. The reaction was stopped by adding water and saturated aqueous NaHCO₃. The mixture was extracted with EtOAc (×3) and the combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was azeotropically distiled with benzene (100 mL) and purified by flash column chromatography (silica gel, benzene/EtOAc=9:1) to afford 28 as a red amorphous solid (53.6 mg, 64%). ¹H NMR (400 MHz, CDCl₃): δ = 2.38 (s, 3H), 3.22 (s, 3H), 3.84 (s, 3H), 3.93 (s, 3H), 3.95 (s, 3H), 4.05 (s, 3H), 4.53 (s, 2H), 4.78 (s, 2H), 5.19 (s, 2H), 6.57 (d, J=1.7 Hz, 1H), 6.94 (d, J=1.7 Hz, 1H), 7.33 (t, J= 7.3 Hz, 1 H), 7.38(s, 1 H), 7.40 (t, J=7.3 Hz, 2 H), 7.55 (d, J=7.3 Hz, 2 H), 8.84 (s, 1 H), 10.73 ppm (s, 1 H; OH).

2-Chloro-6-hydroxymethyl-4-methoxyphenol: Me₂AlCl (1.0м toluene solution, 24.4 mL, 0.263 mol) and (HCHO)_n (8.67 g, 0.263 mol) were added to a solution of 2-chloro-4-methoxyphenol (29) (27.7 g, 0.175 mol) in CH₂Cl₂ (500 mL) at 0°C. After the addition was complete, the ice bath was removed and the reaction mixture was stirred for 5 h. The reaction was stopped by adding 2M aq. HCl at 0°C and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na2SO4), and concentrated in vacuo. After adding Et2O and hexane to the residue, the precipitate was collected by filtration to afford 2-chloro-6-hydroxymethyl-4-methoxyphenol as a white crystalline solid (25.9 g, 89%). The filtrate was concentrated in vacuo and the residue was purified by flash column chromatography (silica gel, hexane/EtOAc= 6:4) to afford 2-chloro-6-hydroxymethyl-4-methoxyphenol as a white solid (4.94 g, 4%; total yield: 93%). M.p. 70-72°C (hexane and EtOAc); ¹H NMR (400 MHz, CDCl₃): $\delta = 2.51$ (t, J = 5.3 Hz, 1H; OH), 3.75 (s, 3H), 4.73 (d, J=5.3 Hz, 2H), 6.26 (brs, 1H; OH), 6.70 (d, J=3.0 Hz, 1 H), 6.82 ppm (d, J = 3.0 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 55.9$, 62.7, 113.2, 113.6, 120.4, 128.0, 144.1, 153.1 ppm; IR (neat): $\tilde{\nu} = 3510$, 3250, 2930, 2840, 1590, 1480, 1450, 1430, 1380, 1350, 1320, 1290, 1240, 1190, 1170, 1130, 1050, 1040, 980, 930, 860, 830, 790 cm⁻¹; elemental analysis calcd (%) for C₈H₉O₃Cl: C 50.94, H 4.81; found: C 50.77, H 4.95.

3-Chloro-2,5-dimethoxybenzyl alcohol: K_2CO_3 (68.1 g, 492 mmol) and Me_2SO_4 (15.0 mL, 158 mmol) were added to a solution of 2-chloro-6-hy-

droxymethyl-4-methoxyphenol (18.6 g, 98.6 mmol) in acetone (400 mL) at RT. The reaction mixture was heated at reflux for 13 h. After cooling to 0°C, Et₃N (150 mL) was added and the stirring was continued for 1 h. The resulting suspension was filtered through a Celite pad (washed with EtOAc) and the volatile materials were removed in vacuo. The resulting mixture was diluted with EtOAc and water, and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. Recrystallization from benzene afforded 3-chloro-2,5-(dimethoxyphenyl)methanol as colorless needles (1st crop: 11.0 g, 55 %; 2nd crop: 4.57 g, 23 %). The filtrate was concentrated in vacuo and the residue was purified by flash column chromatography (silica gel, hexane/EtOAc=7:3) to afford 3-chloro-2,5-dimethoxyphenylmethanol as a white solid (2.50 g, 13%; total yield: 91%). M.p. 54–55 °C (benzene); ¹H NMR (400 MHz, CDCl₃): $\delta = 2.25$ (t, J =3.9 Hz, 1 H; OH), 3.77 (s, 3 H), 3.83 (s, 3 H), 4.69 (d, J=3.9 Hz, 2 H), 6.84 (d, J=2.9 Hz, 1 H), 6.85 ppm (d, J=2.9 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃): δ = 55.7, 61.1, 61.2, 112.8, 114.7, 127.9, 136.2, 147.4, 156.0 ppm; IR (neat): $\tilde{\nu} = 3420, 2940, 2840, 1600, 1570, 1480, 1430, 1360, 1310, 1270,$ 1230, 1180, 1120, 1050, 1000, 950, 860, 760 cm⁻¹; elemental analysis calcd (%) for C₉H₁₁O₃Cl: C 53.35, H 5.47; found: C 53.46, H 5.49.

3-Chloro-2,5-dimethoxybenzaldehyde (30): MnO2 (33.4 g, 384 mmol) was added to a solution of 3-chloro-2,5-(dimethoxyphenyl)methanol (15.5 g, 76.5 mmol) in CH2Cl2 (260 mL) at RT and the reaction mixture was warmed to 40 °C and stirred. After about every 1 h, MnO₂ (ca. 5 g) was added seven times and then the suspension was stirred further 10 h. The reaction mixture was filtered through a Celite pad (washed with EtOAc) and concentrated in vacuo. Recrystallization from benzene afforded aldehyde 30 as colorless needles (1st crop: 7.92 g, 52 %; 2nd crop: 1.27 g, 8 %; 3rd crop: 2.85 g. 19%). The filtrate was concentrated in vacuo and the residue was purified by flash column chromatography (silica gel, hexane/ EtOAc=9:1) to afford 30 as a white solid (1.72 g, 11%; total yield: 90%). M.p. 69–70 °C (benzene); ¹H NMR (400 MHz, CDCl₃): $\delta = 3.82$ (s, 3H), 3.95 (s, 3H), 7.21 (d, J=3.0 Hz, 1H), 7.23 (d, J=3.0 Hz, 1H), 10.25 ppm (s, 1H); ¹³C NMR (100 MHz, CDCl₃): δ = 56.0, 63.5, 109.4, 123.6, 129.4, 130.6, 153.4, 156.1, 188.9 ppm; IR (KBr): $\tilde{\nu} = 3080$, 3020, 2970, 2940, 2870, 2840, 2750, 1690, 1610, 1470, 1420, 1390, 1330, 1260, 1230, 1210, 1110, 1050, 990, 870, 850, 770 cm⁻¹; elemental analysis calcd (%) for C₉H₉O₃Cl: C 53.88, H 4.52; found: C 53.72, H 4.65.

Ethyl 4-acetoxy-7-chloro-5,8-dimethoxy-2-naphthoate (31): tert-Butyl 3ethoxycarbonyl-3-diethoxyphosphorylpropionate (19) (1.80 g, 5.32 mmol) in THF (10 mL) was added to a suspension of NaH (60% dispersion in mineral oil, washed with hexane (209 mg, 5.22 mmol)) in THF (10 mL) at 0°C and the reaction mixture was stirred for 2 h. A solution of 30 (955 mg, 4.76 mmol) in THF (10 mL) was added at 0°C. The reaction mixture was warmed to RT and the stirring was continued for 1 h. The reaction was stopped by adding water and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with water and brine, dried (Na₂SO₄), and concentrated in vacuo. The resulting mixture was dissolved in trifluoroacetic acid/water (9:1, 34 mL) and the reaction mixture was stirred for 40 min at RT. After adding toluene, azeotropic evaporation was performed three times. NaOAc (3.45 g, 42.0 mmol) and Ac₂O (47.0 mL, 499 mmol) were added to this crude material at RT. The reaction mixture was heated at reflux for 1 h. After cooling to RT, the reaction was stopped by adding water and then the mixture was extracted with EtOAc $(\times 3)$. The combined organic extracts were washed with saturated aqueous NaHCO3 and brine, dried (Na2SO4), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=77:23) to afford 31 as a pale yellow solid (1.24 g, 76%). M.p. 104-105°C (hexane and EtOAc); ¹H NMR (400 MHz, CDCl₃): $\delta = 1.43$ (t, J = 7.3 Hz, 3H), 2.38 (s, 3H), 3.92 (s, 3H), 3.99 (s, 3 H), 6.88 (s, 1 H), 4.44 (q, J = 7.3 Hz, 2 H), 7.68 (d, J = 1.7 Hz, 1 H), 8.71 ppm (d, J = 1.7 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 14.3$, 20.8, 56.6, 61.5, 61.8, 109.9, 119.5, 120.8, 123.0, 124.2, 129.3, 131.0, 146.5, 147.0, 151.6, 165.5, 170.7 ppm; IR (KBr): $\tilde{\nu} = 3411$, 2984, 2942, 2846, 1770, 1760, 1715, 1593, 1478, 1459, 1424, 1387, 1366, 1333, 1271, 1232, 1214, 1160, 1130, 1117, 1029, 939, 914, 872, 840, 814 cm⁻¹; elemental analysis calcd (%) for C₁₇H₁₇O₃Cl: C 57.88, H 4.86; found: C 57.66, H 4.98.

7-*Chloro-4-hydroxy-5,8-dimethoxy-2-naphthoic* acid (**32**): A 3M aq. NaOH solution of (23.0 mL) was added to a suspension of **31** (2.60 g, 7.35 mmol) in THF/EtOH (1:1) (30 mL) and the reaction mixture was heated at 70 °C for 1 h. After cooling to RT, the aqueous layer was washed with Et₂O (×2). The aqueous phase was acidified with 2M aq. HCl. The precipitates were collected by filtration to afford **32** as a white powder (1.81 g, 87%). M.p. > 250 °C; ¹H NMR (400 MHz, [D₆]DMSO): δ = 3.86 (s, 3H), 3.99 (s, 3H), 7.06 (s, 1H), 7.27 (d, *J* = 1.4 Hz, 1H), 8.06 (d, *J* = 1.4 Hz, 1H), 9.65 (s, 1H; OH), 13.0–13.4 ppm (br, 1H); ¹³C NMR (100 MHz, [D₆]DMSO): δ = 56.9, 61.2, 107.7, 110.3, 114.3, 116.5, 123.0, 130.2, 130.7, 145.7, 152.9, 155.1, 167.0 ppm; IR (KBr): $\tilde{\nu}$ = 3380, 2940, 2640, 1680, 1630, 1600, 1510, 1470, 1430, 1380, 1340, 1310, 1280, 1240, 1210, 1120, 1090, 1020, 940, 820, 800, 770 cm⁻¹; elemental analysis calcd (%) for C₁₃H₁₁O₅Cl: C 55.24, H 3.92; found: C 55.24, H 4.15.

2-Iodo-6-methoxycarbonyl-3-(methoxymethoxy)methyl-5-methylphenyl 7chloro-4-hydroxy-5,8-dimethoxy-2-naphthoate (33): 1-Ethyl-3-[3-(dimethylamino)propyllcarbodiimide hvdrochloride (EDCI) (196 mg. 0.981 mmol) and DMAP (125 mg, 1.02 mmol) were added to a mixed suspension of acid 32 (252 mg, 0.892 mmol) and phenol 14 (651 mg, 1.78 mmol) in CH₂Cl₂ (9 mL) at RT. After stirring for 3 h, the reaction was stopped by adding water. The mixture was extracted with EtOAc (× 3) and the combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=7:3) to afford $\mathbf{33}$ as a yellow solid (438 mg, 78%). M.p. 174.5-175.2°C (hexane and EtOAc); ¹H NMR (400 MHz, CDCl₃): $\delta = 2.45$ (s, 3H), 3.45 (s, 3H), 3.70 (s, 3H), 4.00 (s, 3 H), 4.09 (s, 3 H), 4.63 (s, 2 H), 4.81 (s, 2 H), 6.88 (s, 1 H), 7.33 (s, 1H), 7.63 (s, 1H), 8.50 (s, 1H), 9.32 ppm (s, 1H; OH); ¹³C NMR $(100 \text{ MHz}, \text{ CDCl}_3): \delta = 20.1, 52.4, 55.7, 56.8, 61.7, 73.0, 91.7, 96.2, 108.2,$ 111.1, 116.8, 117.1, 123.6, 126.4, 128.0, 129.0, 130.9, 138.7, 144.2, 147.5, 149.2, 152.4, 155.4, 163.6, 165.9 ppm; IR (KBr): $\tilde{\nu}$ = 3390, 2950, 2890, 2850, 1740, 1600, 1520, 1450, 1390, 1340, 1280, 1200, 1180, 1150, 1120, 1070, 1060, 1040, 990, 950, 920, 880, 800, 750 cm⁻¹; elemental analysis calcd (%) for C25H24O9Cl: C 47.60, H 3.83; found: C 47.62, H 4.03. Starting material 14 was also recovered (359 mg, 55 %).

Methyl (\pm) -3-(6-chloro-1-hydroxy-3-hydroxymethyl-5,8-dimethoxynaphthalen-2-yl)-2-hydroxy-4-(methoxymethoxy)methyl-6-methylbenzoate (35): A suspension of ester 33 (307 mg, 0.486 mmol), Pd(OAc)₂ (33.3 mg, 0.148 mmol, 30 mol%), PPh3 (81.1 mg, 0.309 mmol), and sodium pivalate (187 mg, 1.50 mmol) in DMA (33 mL) was heated at 110 °C for 1.5 h. After cooling to RT, the resulting dark brown suspension was filtered through a Celite pad (washed with EtOAc). The filtrate was washed with water and brine, dried (Na₂SO₄), and concentrated in vacuo. NaBH₄ (29.2 mg, 0.772 mmol) was added to a solution of this crude material, which included 34, in THF/MeOH (20:1) (17 mL) at -40 °C. After stirring for 3 h, the reaction was stopped by adding five drops of AcOH at that temperature. After warming to RT, water was added and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with saturated aqueous NaHCO₃ and brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, benzene/EtOAc = 85:15) to afford (\pm)-35 as a yellow solid (211 mg, 86%).

34: M.p. 218.5–219.3 °C (hexane and Et₂O); ¹H NMR (400 MHz, CDCl₃): $\delta = 2.44$ (s, 3H), 3.31 (s, 3H), 3.98 (s, 3H), 4.03 (s, 3H), 4.15 (s, 3H), 4.58 (s, 2H), 4.72 (s, 2H), 6.94 (s, 1H), 7.43 (s, 1H), 8.57 (s, 1H), 10.16 ppm (s, 1H; OH); ¹³C NMR (100 MHz, CDCl₃): $\delta = 19.6$, 52.7, 55.5, 57.1, 61.8, 68.9, 96.2, 109.0, 113.7, 114.3, 116.4, 116.7, 121.2, 123.5, 124.3, 126.0, 129.9, 136.6, 138.9, 146.8, 147.6, 151.4, 152.2, 160.4, 167.0 ppm; IR (KBr): $\tilde{\nu} = 3310, 2950, 1740, 1720, 1610, 1600, 1460, 1440, 1390, 1360, 1330, 1260, 1200, 1150, 1090, 1050, 1040, 1010, 950, 920, 840, 810, 760, 710 cm⁻¹; elemental analysis calcd (%) for C₂₅H₂₃O₉Cl: C 59.71, H 4.61; found: C 59.65, H 4.57.$

(±)-**35**: M.p. 142.5–143.3 °C (hexane and EtOAc); ¹H NMR (400 MHz, CDCl₃): δ =2.64 (s, 3H), 2.72 (t, *J*=6.3 Hz, 1H; OH), 3.13 (s, 3H), 3.979 (s, 3H), 3.981 (s, 3H), 4.00 (s, 3H), 4.27 (d, *J*=12.2 Hz, 1H), 4.34 (d, *J*=12.2 Hz, 1H), 4.42 (d, *J*=6.6 Hz, 1H), 4.46 (d, *J*=6.3 Hz, 2H), 4.53 (d, *J*=6.6 Hz, 1H), 6.74 (s, 1H), 7.03 (s, 1H), 7.80 (s, 1H), 9.45 (s, 1H; OH), 11.77 ppm (s, 1H; OH); ¹³C NMR (100 MHz, CDCl₃): δ =24.3, 52.4, 55.3,

56.6, 61.3, 64.1, 67.4, 96.2, 105.9, 111.9, 113.1, 114.0, 117.2, 121.3, 122.8, 123.3, 131.3, 141.2, 141.4, 143.7, 146.6, 151.3, 152.5, 159.9, 172.4 ppm; IR (KBr): $\bar{\nu}$ =3390, 2940, 1660, 1620, 1600, 1450, 1380, 1360, 1290, 1260, 1200, 1150, 1110, 1050, 1010, 970, 950, 820 cm⁻¹; elemental analysis calcd (%) for C₂₅H₂₇O₉Cl: C 59.23, H 5.37; found: C 59.14, H 5.50.

Methyl (\pm) -3-(6-chloro-1-hydroxy-3-hydroxymethyl-5,8-dimethoxynaphthalen-2-yl)-2-hydroxy-4-hydroxymethyl-6-methylbenzoate (38): А 6м аq. HCl solution (7 mL) was added to a solution of (\pm) -35 (516 mg, 1.02 mmol) in DME (10 mL) at RT and the reaction mixture was heated at 50 °C for 3 h. After the mixture had cooled to RT, water was added and the mixture was extracted with EtOAc (\times 3). The combined organic extracts were washed with water and brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=45:55) to afford (\pm) -38 as a white solid (440 mg, 93%). M.p. 117-120°C (hexane and EtOAc); ¹H NMR $(500 \text{ MHz}, \text{ CDCl}_3): \delta = 2.64 \text{ (s, 3H)}, 2.35-2.70 \text{ (br, 2H; OH)}, 3.98 \text{ (s, })$ 3H), 3.99 (s, 3H), 4.01 (s, 3H), 4.32 (s, 2H), 4.45 (d, J=12.0 Hz, 1H), 4.50 (d, J=12.0 Hz, 1 H), 6.76 (s, 1 H), 7.04 (s, 1 H), 7.79 (s, 1 H), 9.54 (s, 1 H; OH), 11.76 ppm (s, 1 H; OH); 13 C NMR (125 MHz, CDCl₃): $\delta = 24.2$, 52.3, 56.6, 61.3, 63.4, 64.2, 106.2, 111.9, 113.4, 114.1, 117.5, 121.0, 123.0, 123.8, 131.1, 140.8, 141.7, 146.3, 146.7, 151.3, 152.4, 160.6, 172.4 ppm; IR (neat): $\tilde{\nu} = 3390, 2940, 1650, 1620, 1600, 1450, 1390, 1360, 1300, 1260,$ 1200, 1130, 1110, 1070, 1040, 1000, 980, 820, 750 cm⁻¹; elemental analysis calcd (%) for $C_{23}H_{23}O_8Cl$: C 59.68, H 5.01; found: C 59.76, H 5.31.

 $Methyl \quad (\pm)-4-(tert-butyl dimethyl siloxy) methyl-3-[3-(tert-butyl dimethyl siloxy)] \\$ loxy)methyl-6-chloro-1-hydroxy-5,8-dimethoxynaphthalen-2-yl]-2-hydroxy-6-methylbenzoate (36): Imidazole (211 mg, 3.10 mmol) and tert-butylchlorodimethylsilane (355 mg, 2.36 mmol) were added to a solution of (\pm) -38 (358 mg, 0.774 mmol) in DMF (5 mL) at RT. After stirring for 25 min, the reaction was stopped by adding water and the mixture was extracted with Et₂O (×3). The combined organic extracts were washed with water and brine, dried (Na2SO4), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/ EtOAc=85:15) to afford (\pm)-36 as a white solid (451 mg, 84%). M.p. 162.5–163.4°C (hexane and EtOAc); ¹H NMR (500 MHz, CDCl₃): $\delta =$ -0.05 (s, 3H), -0.04 (s, 3H), 0.01 (s, 3H), 0.03 (s, 3H), 0.88 (s, 9H), 0.95 (s, 9H), 2.64 (s, 3H), 3.97 (s, 3H), 3.98 (s, 3H), 3.99 (s, 3H), 4.27 (d, J= 15.0 Hz, 1 H), 4.40 (d, J=16.0 Hz, 1 H), 4.49 (d, J=15.0 Hz, 1 H), 4.59 (d, J=16.0 Hz, 1 H), 6.71 (s, 1 H), 7.11 (s, 1 H), 7.90 (s, 1 H), 9.36 (s, 1 H; OH), 11.55 ppm (s, 1H; OH); 13 C NMR (125 MHz, CDCl₃): $\delta = -5.49$, -5.43, -5.40, -5.38, 18.3 (2C), 24.5, 25.9, 52.1, 56.5, 61.1, 62.5, 62.9, 105.3, 110.0, 110.9, 113.5, 115.5, 118.7, 120.9, 122.4, 130.9, 140.9, 141.8, 146.5, 147.1, 150.8, 152.7, 159.6, 172.5 ppm; IR (neat): $\tilde{\nu} = 3400$, 2960, 2860, 1740, 1660, 1610, 1500, 1460, 1390, 1360, 1290, 1260, 1200, 1120, 1060, 1020, 1010, 990, 960, 950, 840, 780 cm⁻¹; elemental analysis calcd (%) for C₃₅H₅₁O₈ClSi₂: C 60.80, H 7.43; found: C 60.57, H 7.62.

Camphanoyl esters **37a** and **37b**: (1S)-(-)-Camphanic chloride (235 mg, 1.08 mmol) and DMAP (18.0 mg, 0.147 mmol) were added to a solution of (\pm) -**36** (497 mg, 0.719 mmol) in pyridine (3.6 mL) at RT and the reaction mixture was stirred for 20 h. The reaction was stopped by adding water and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with 2 M aq. HCl (×2), saturated aqueous NaHCO₃, and brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by column chromatography (hexane/EtOAc=8:2) to afford the less polar ester **37b** as a white solid (250 mg, 40%) and a more polar mixture that included **37a**. The mixture was further purified by column chromatography (benzene/EtOAc=6:4) to afford the more polar ester **37a** (237 mg, 38%).

37a: R_f =0.34 (hexane/EtOAc=7:3); m.p. 97.0–98.0°C (hexane and EtOAc); $[a]_D^{22} = -60.3$ (c=0.975 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ =-0.07 (brs, 6H), 0.03 (s, 3H), 0.05 (s, 3H), 0.40 (s, 3H), 0.64 (s, 3H), 0.86 (s, 9H), 0.93 (s, 3H), 0.96 (s, 9H), 1.40–1.72 (m, 3H), 1.90–2.02 (m, 1H), 2.51 (s, 3H), 3.85 (s, 3H), 3.92 (s, 3H), 4.01 (s, 3H), 4.33 (d, J= 14.2 Hz, 1 H), 4.35 (d, J=15.4 Hz, 1 H), 4.35 (d, J=15.4 Hz, 1 H), 6.73 (s, 1 H), 7.48 (s, 1 H), 7.87 (s, 1 H), 9.30 ppm (s, 1 H; OH); ¹³C NMR (100 MHz, CDCl₃): δ =-5.53, -5.47, -5.45, -5.3, 9.5, 15.6, 15.9, 18.2, 18.3, 20.8, 25.9 (2C), 28.7, 30.6, 52.3, 53.7, 54.7, 56.7, 61.0, 62.3, 62.5, 90.8, 105.5, 110.2, 113.3, 114.1, 123.0, 124.4, 125.2, 127.2, 130.9,

9810 -

138.0, 141.9, 144.1, 145.7, 146.3, 151.6, 152.5, 165.0, 166.8, 177.8 ppm; IR (neat): $\tilde{\nu}$ =3380, 2950, 2860, 1790, 1730, 1620, 1600, 1500, 1460, 1390, 1360, 1320, 1260, 1210, 1170, 1120, 1060, 1020, 990, 950, 840 cm⁻¹; elemental analysis calcd (%) for C₄₅H₆₃O₁₁ClSi₂: C 62.01, H 7.29; found: C 61.77, H 7.55.

37b: R_f =0.42 (hexane/EtOAc=7:3); $[\alpha]_D^{24}$ =+58.2 (*c*=1.00 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ =-0.07 (s, 3H), -0.06 (s, 3H), 0.04 (s, 3H), 0.06 (s, 3H), 0.24 (s, 3H), 0.61 (s, 3H), 0.86 (s, 9H), 0.92 (s, 3H), 0.96 (s, 9H), 1.45-1.56 (m, 1H), 1.63-1.76 (m, 2H), 1.96-2.05 (m, 1H), 2.51 (s, 3H), 3.85 (s, 3H), 3.93 (s, 3H), 4.00 (s, 3H), 4.33 (d, *J*=14.4 Hz, 1H), 4.35 (d, *J*=15.1 Hz, 1H), 4.45 (d, *J*=14.4 Hz, 1H), 4.57 (d, *J*= 15.1 Hz, 1H), 6.71 (s, 1H), 7.48 (s, 1H), 7.87 (s, 1H), 9.29 ppm (s, 1H; OH); ¹³C NMR (100 MHz, CDCl₃): δ =-5.44, -5.40, -5.37, -5.2, 9.6, 15.4, 16.0, 18.28, 18.34, 20.8, 25.7, 25.9, 28.7, 30.8, 52.2, 53.6, 54.7, 56.7, 61.0, 62.3, 62.4, 90.6, 105.4, 110.0, 113.2, 113.9, 122.9, 124.4, 124.9, 127.0, 130.8, 138.0, 141.7, 143.9, 145.8, 146.0, 151.6, 152.5, 165.0, 166.6, 177.7 ppm; IR (neat): $\tilde{\nu}$ =3380, 3020, 2960, 2930, 2860, 1790, 1730, 1620, 1600, 1500, 1460, 1390, 1360, 1320, 1260, 1210, 1100, 1060, 1030, 990, 950, 840, 760 cm⁻¹; elemental analysis calcd (%) for C₄₅H₆₃O₁₁ClSi₂: C 62.01, H 7.29; found: C 61.73, H 7.39.

Tetrol (M)-38: Two drops of 40% aqueous HF were added to a solution of 37 a (237 mg, 0.272 mmol) in CH₃CN (3 mL) at 0 °C. The reaction mixture was gradually warmed to RT and stirred for 45 min. The reaction was stopped by adding water and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with saturated aqueous NaHCO3 and brine, dried (Na2SO4), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/ EtOAc=35:65) to afford the triol derivative of 37a as a white solid (170 mg, 97%). M.p. 242–243°C; $[\alpha]_D^{26} = -108$ (c = 1.07 in CHCl₃); ¹H NMR (400 Hz, CDCl₃): $\delta = 0.26$ (s, 3H), 0.67 (s, 3H), 0.92 (s, 3H), 1.42-1.78 (m, 3H), 1.95-2.08 (m, 1H), 2.48 (s, 3H), 2.70-3.15 (br, 2H; OH), 3.86 (s, 3H), 3.92 (s, 3H), 4.01 (s, 3H), 4.29 (d, J=12.5 Hz, 1H), 4.34 (d, J=12.5 Hz, 1 H), 4.42 (d, J=12.5 Hz, 1 H), 4.49 (d, J=12.5 Hz, 1H), 6.76 (s, 1H), 7.44 (s, 1H), 7.55 (s, 1H), 9.47 ppm (s, 1H; OH); ¹³C NMR (100 MHz, CDCl₃): $\delta = 9.5$, 15.4, 16.0, 20.4, 28.7, 30.8, 52.4, 53.7, 54.7, 56.8, 61.3, 62.8, 63.1, 90.6, 106.2, 113.2, 113.7, 115.8, 123.4, 126.1, 126.4, 129.7, 131.1, 138.4, 141.1, 143.8, 145.8, 146.4, 152.1, 152.3, 165.2, 166.6, 177.6 ppm; IR (neat): $\tilde{\nu} = 3380$, 2960, 2880, 1770, 1730, 1620, 1600, 1510, 1460, 1390, 1360, 1320, 1260, 1210, 1170, 1130, 1100, 1060, 1040, 990, 940, 870, 810, 800 cm⁻¹; elemental analysis calcd (%) for C33H35O11Cl: C 61.63, H 5.49; found: C 61.35, H 5.69.

K₂CO₃ (367 mg, 2.66 mmol) was added to a solution of the triol derivative of **37a** (170 mg, 0.264 mmol) in MeOH (3 mL). After stirring for 19 h at RT, the reaction was stopped by adding 2 M aq. HCl and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with saturated aqueous NaHCO₃ and brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=4:6) to afford tetrol (*M*)-**38** as a white solid (122 mg, quant). $[a]_D^{28} = -52.7$ (*c*=1.07 in CHCl₃); m.p. 117–119 °C (hexane and EtOAc).

Dialdehyde (M)-39: K₂CO₃ (235 mg, 1.70 mmol) and MeI (0.11 mL, 1.77 mmol) were added to a solution of (M)-38 (78.6 mg, 0.170 mmol) in acetone (3.4 mL) at RT. The reaction mixture was warmed to 40 °C and stirred for 11 h. The reaction was stopped by adding water and the mixture was extracted with EtOAc (\times 3). The combined organic extracts were washed with 2M aq. HCl, saturated aqueous NaHCO₃, and brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (benzene/EtOAc=1:1) to afford the dimethyl ether derivative of (*M*)-**38** as a white solid (75.8 mg, 91 %). M.p. 171.5–172.3 °C; $[\alpha]_{\rm D}^{27} = -106$ $(c=1.10 \text{ in CHCl}_3)$; ¹H NMR (400 MHz, CDCl₃): $\delta = 2.38$ (s, 3H), 3.01 (brs, 1H; OH), 3.23 (brs, 1H; OH), 3.44 (s, 3H), 3.53 (s, 3H), 3.92 (s, 3H), 3.95 (s, 3H), 3.99 (s, 3H), 4.23 (brs, 2H), 4.37 (d, J=12.4 Hz, 1H), 4.45 (d, J=12.4 Hz, 1H), 6.81 (s, 1H), 7.22 (s, 1H), 8.04 ppm (s, 1H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 19.4$, 52.3, 56.6, 61.4 (2C), 61.8, 63.2, 63.7, 107.7, 118.5, 119.4, 123.5, 126.2, 126.6, 127.0, 127.5, 131.8, 137.1, 139.9, 142.5, 145.6, 152.7, 153.8, 154.5, 168.7 ppm; IR (neat): $\tilde{\nu} = 3390$, 3010, 2940, 2840, 1730, 1590, 1570, 1460, 1390, 1340, 1290, 1200, 1140,

1100, 1050, 960, 920, 890, 800, 750 $\rm cm^{-1};$ elemental analysis calcd (%) for $\rm C_{25}H_{27}O_8Cl\colon C$ 61.16, H 5.54; found: C 61.25, H 5.61.

MnO₂ (62.0 mg, 0.713 mmol) was added to a solution of the dimethyl ether derivative of (M)-38 (71.3 mg, 0.145 mmol) in CH₂Cl₂ (3 mL) at RT and the reaction mixture was stirred for 26 h. The reaction was monitored by TLC and MnO2 (62.0 mg, 0.713 mmol) was added about every 4 h. When the reaction was completed, the mixture was filtered through a Celite pad and concentrated in vacuo. The residue was purified by PTLC (benzene/EtOAc=6:4) to afford dialdehyde 39 as a yellow oil (55.4 mg, 79%). $[a]_{D}^{27} = +16.7 (c = 1.08 \text{ in CHCl}_3)$; ¹H NMR (400 MHz, $CDCl_3$): $\delta = 2.46$ (s, 3H), 3.36 (s, 3H), 3.46 (s, 3H), 3.95 (s, 3H), 3.97 (s, 3H), 4.06 (s, 3H), 6.99 (s, 1H), 7.71 (s, 1H), 8.59 (s, 1H), 9.74 (s, 1H), 9.88 ppm (s, 1 H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 19.4$, 52.6, 56.8, 61.4, 61.5, 61.9, 110.8, 122.0, 122.2 (2 C), 123.4, 125.0, 125.5, 128.0, 131.4, 133.4, 133.6, 135.9, 137.5, 146.9, 152.8, 155.2, 167.7, 190.8, 190.9 ppm; IR (neat): $\tilde{\nu} = 3380, 3020, 2940, 2840, 2750, 2000, 1740, 1700, 1600, 1570, 1460, 1390,$ 1360, 1340, 1280, 1240, 1210, 1190, 1140, 1100, 1080, 1050, 1000, 970, 930, 810, 750 cm $^{-1}\!;$ elemental analysis calcd (%) for $C_{25}H_{23}O_8Cl\colon$ C 61.67, H 4.76: found: C 61.77, H 5.02.

The enantiomeric purity of **39** was assessed by HPLC analysis (DAICEL CHIRALCEL OD-H (0.46 cm $\varphi \times 25$ cm), hexane/*i*PrOH=9:1, flow rate = 0.7 mLmin⁻¹, $t_{\rm R}$ =18.3 min for the *M* isomer, 26.4 min for the *P* isomer).

Diol 40: SmI₂ (0.1 M THF solution, 3.5 mL, 0.35 mmol) was added to a solution of (M)-39 (68.5 mg, 0141 mmol) in THF (3.0 mL) at 0°C. After stirring for 10 min at 0 °C, the reaction was stopped by adding water and the mixture was extracted with EtOAc (×3). The combined organic extracts were successively washed with 2M aq. HCl, water, and brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (benzene/EtOAc = 50:50) to afford 40 as a yellow solid (68.9 mg, quant). M.p. 225–226 °C (decomp); $[\alpha]_{\rm D}^{26} = -282$ (c = 1.08 in CHCl₃), $[\alpha]_{\rm D}^{27} = -215$ (c=1.11 in MeOH); ¹H NMR (400 MHz, CDCl₃): $\delta = 2.37$ (s, 3H), 3.11– 3.36 (br, 2H; OH), 3.39 (s, 3H), 3.42 (s, 3H), 3.92 (s, 3H), 3.94 (s, 3H), 3.97 (s, 3 H), 4.36-4.56 (br, 2 H), 6.79 (s, 1 H), 7.30 (br s, 1 H), 8.06 ppm (s, 1 H); ¹³C NMR (100 MHz, CDCl₃): δ = 19.4, 52.3, 56.5, 61.3, 61.5, 62.3, 73.8, 74.0, 107.4, 111.6, 119.2, 119.4, 120.0, 120.3, 123.8, 128.4, 131.3, 136.5, 137.7, 140.7, 145.3, 153.6, 154.8, 155.1, 169.1 ppm; IR (neat): $\tilde{\nu} =$ 3453, 3009, 2936, 2837, 1728, 1619, 1589, 1574, 1454, 1421, 1343, 1285, 1216, 1120, 1096, 1044, 1002, 969, 926, 892, 816, 798, 755 cm⁻¹; UV (MeOH): λ_{max} =357 (ϵ =8500), 339 (7800), 323 (9500), 309 (10000), 278 (34000), 270 (34000), 229 nm (34000 M^{-1} cm⁻¹); CD (MeOH): $\lambda_{ext} = 354$ $(\Delta \varepsilon = -2.8)$, 321 (-8.0), 278 (+14), 246 (-32), 224.5 nm (+35 M⁻¹ cm⁻¹); elemental analysis calcd (%) for C25H25O8Cl: C 61.42, H 5.15; found: C 61.25, H 5.35.

The enantiomeric purity of **40** was assessed by HPLC analysis (DAICEL CHIRALCEL OD-H (0.46 cm $\varphi \times 25$ cm), hexane/*i*PrOH=9:1, flow rate = 1.0 mLmin⁻¹, $t_{\rm R}$ = 10.9 min for the *R*,*R* isomer, 15.0 min for the *S*,*S* isomer).

Diacetate 41: Ac₂O (0.050 mL, 0.53 mmol) and a catalytic amount of DMAP were added to a solution of 40 (63.5 mg, 0.131 mmol) in pyridine (2.6 mL) at RT. After stirring for 20 min, the reaction was stopped by adding water and the mixture was extracted with EtOAc (×2). The combined organic extracts were successively washed with 1 M aq. HCl, water, saturated aqueous NaHCO3, and brine, dried (Na2SO4), and concentrated in vacuo. The residue was purified by PTLC (benzene/EtOAc=85:15) to afford 41 as a white amorphous solid (72.9 mg, 98%). M.p. 98-100°C; $[\alpha]_{D}^{26} = +106 \ (c = 1.15 \ \text{in CHCl}_3); {}^{1}\text{H NMR} \ (400 \ \text{MHz}, \ \text{CDCl}_3 \ \text{at } 300 \ \text{K}):$ $\delta = 1.70-2.45$ (br, 6H), 2.37 (s, 3H), 3.43 (s, 3H), 3.46 (s, 3H), 3.94 (s, 3H), 3.98 (brs, 6H), 5.58-6.20 (br, 2H), 6.84 (brs, 1H), 7.05-7.30 (br, 1 H), 7.55–8.15 ppm (br, 1 H); ¹H NMR (400 MHz, [D₆]DMSO, at 333 K): $\delta = 1.80 - 2.20$ (br, 6H), 2.28 (s, 3H), 3.321 (s, 3H), 3.324 (s, 3H), 3.88 (s, 3H), 3H), 3.90 (s, 3H), 3.96 (s, 3H), 5.91 (d, J=7.1 Hz, 1H), 6.00 (d, J= 7.1 Hz, 1H), 7.02 (s, 1H), 7.06–7.14 (br, 1H), 7.60–7.92 ppm (br, 1H); ^{13}C NMR (100 MHz, CDCl₃): $\delta\!=\!18.3,\ 20.2,\ 20.3,\ 51.8,\ 56.8,\ 60.4,\ 60.9,$ 61.2, 70.1, 70.3, 108.5, 119.5, 120.4, 123.1, 129.0, 130.0, 132.3, 135.3, 135.4, 144.7, 153.1, 155.0, 155.1, 167.5, 169.3 ppm (signals for four sp² carbon atoms were not detected presumably owing to peak-broadening ascribed to the slow conformational interconversion, diaxial diequatorial); IR

(neat): $\bar{\nu}$ =3010, 2940, 2840, 1760, 1730, 1620, 1580, 1460, 1370, 1340, 1290, 1220, 1140, 1100, 1030, 970, 940, 920, 900, 890, 850, 810, 760 cm⁻¹; elemental analysis calcd (%) for C₂₉H₂₉O₁₀Cl: C 60.79, H 5.10; found: C 60.69, H 5.38.

Quinone 42: A solution of CAN (299 mg, 0.545 mmol) in water (2 mL) was added to a solution of 41 (126 mg, 0.220 mmol) in CH₃CN (3 mL) at 0°C. After stirring for 10 min at 0°C, the reaction was stopped by adding water and the mixture was extracted with EtOAc (\times 3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=4:6) to afford 42 as a yellow solid (120 mg, quant). M.p. 187–188 °C; $[\alpha]_D^{25} = +98.9$ (c=1.04 in CHCl₃); ¹H NMR (400 MHz, $[D_6]$ DMSO, 333 K): $\delta = 1.92-2.26$ (br, 6H), 2.30 (s, 3H), 3.39 (s, 3H), 3.49 (s, 3H), 3.89 (s, 3H), 5.94 (d, J=7.3 Hz, 1H), 6.00 (d, J= 7.3 Hz, 1H), 7.06–7.20 (br, 1H), 7.36 (s, 1H), 7.77–7.94 pppm (br, 1H); ¹³C NMR (100 MHz, [D₆]DMSO): δ =18.5, 20.17, 20.22, 52.0, 60.9, 61.5, 69.6 (br, 2C), 78.9, 119.0 (br), 120.7, 123.6 (br), 128.8, 131.2, 132.2, 135.2 (br), 137.4, 137.5, 138.8 (br), 142.4, 155.2, 157.9, 167.1, 169.3 (2 C), 177.0, 180.7 ppm; IR (neat): $\tilde{v} = 3470$, 3020, 2930, 2850, 1740, 1680, 1660, 1610, 1580, 1460, 1370, 1300, 1260, 1230, 1140, 1080, 1040, 980, 940, 900, 860, 750 cm⁻¹; elemental analysis calcd (%) for C₂₇H₂₃O₁₀Cl: C 59.73, H 4.27; found: C 59.49, H 4.48.

Anthraquinone 43: A solution of siloxydiene 21 (94.5 mg, 0.467 mmol) in THF (2 mL) was added to a solution of 42 (120 mg, 0.220 mmol) in THF (3 mL) at 0 °C. Immediately, the ice bath was removed and the reaction mixture was stirred for 2 h at RT. Acidic SiO₂ (pH 6) (8 g) was added to the reaction mixture and the solvent removed in vacuo. After standing for 12 h, the SiO₂ was placed on a Celite pad, washed with EtOAc, and then the filtrate was concentrated in vacuo. K₂CO₃ (348 mg, 2.52 mmol) was added to the solution of this crude material in THF/CH₂Cl₂ (1:3) (10 mL) at RT. After stirring for 1.5 h, K₂CO₃ (324 mg, 2.34 mmol) was added to the reaction mixture and stirred for a further 1 h. The reaction was stopped by adding 1 M aq. HCl and the mixture was extracted with CH₂Cl₂ (×3). The combined organic extracts were washed with water and brine, dried (Na₂SO₄), and concentrated in vacuo. After adding Et₂O to the residue, the precipitate was collected by filtration to afford quinone 43 as a yellow powder (101 mg, 76%). The filtrate was concentrated in vacuo and the residue was purified by PTLC (CHCl₃/MeOH=95:5) to afford 43 as a vellow solid (18.8 mg, 14%; total yield: 90%). M.p. > 220 °C; $[\alpha]_{D}^{26} = +76.8$ (c = 0.800 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): $\delta = 1.70-2.37$ (br, 6H), 2.38 (s, 3H), 3.51 (s, 3H), 3.64 (s, 3H), 3.95 (s, 3H), 3.96 (s, 3H), 5.90-6.16 (br, 2H), 6.68 (d, J=2.5 Hz, 1H), 6.80-7.30 (br, 1H), 7.34 (d, J=2.5 Hz, 1H), 7.80-8.40 (br, 1H), 12.71 ppm (s, 1H; OH); ¹³C NMR (100 MHz, CDCl₃, 323 K): $\delta = 19.4$, 20.8, 29.7, 52.4, 56.1, 61.8, 62.2, 70.7 (br), 106.1, 107.6, 110.6, 126.3 (br), 129.8 (br), 131.7 (br), 135.0, 135.6 (br), 136.8, 138.3, 139.9 (br), 156.3, 159.5, 165.1, 166.8, 168.2, 169.8, 181.1, 185.7 ppm (signals for four sp² carbon atoms were not detected presumably owing to peak-broadening ascribed to the slow conformational interconversion, diaxial \rightleftharpoons diequatorial); IR (KBr): $\tilde{\nu} = 3450$, 3020, 2950, 1740, 1680, 1630, 1610, 1590, 1440, 1370, 1310, 1260, 1230, 1160, 1120, 1070, 1040, 970, 750 cm⁻¹; elemental analysis calcd (%) for C₃₂H₂₈O₁₂: C 63.57, H 4.67; found: C 63.37, H 4.86.

Phenol 44: BCl₃ (1.0 M in hexane, 0.50 mL, 0.50 mmol) was added to a solution of 43 (31.9 mg, 52.8 µmol) in CH₂Cl₂ (2 mL) at -10 °C. The reaction mixture was stirred for 30 min and then the reaction was stopped by adding saturated aqueous NaHCO3. After adding 1M aq. HCl, the mixture was extracted with CH_2Cl_2 (×3). The combined organic extracts were washed with brine, dried (Na2SO4), and concentrated in vacuo. After adding Et₂O to the residue, the precipitate was collected by filtration to afford quinone 44 as an orange powder (28.1 mg, 92%). The filtrate was concentrated in vacuo and the residue was purified by PTLC (CHCl₃/MeOH=97:3) to afford 44 as a red powder (2.0 mg, 7%; total yield: 99%). M.p.>220°C; $[\alpha]_D^{31} = +127$ (c=0.500 in CHCl₃); ¹H NMR (400 MHz, CDCl₃, 323 K): $\delta = 2.08$ (s, 3H), 2.13 (s, 3H), 2.48 (s, 3H), 3.94 (s, 3H), 3.99 (s, 3H), 5.99 (d, J=7.8 Hz, 1H), 6.05 (d, J=7.8 Hz, 1H), 6.72 (d, J=2.5 Hz, 1H), 6.85 (s, 1H), 7.40 (d, J=2.5 Hz, 1H), 7.90 (s, 1H), 10.18 (brs, 1H; OH), 12.78 (s, 1H; OH), 14.27 ppm (brs, 1H; OH); ¹³C NMR (100 MHz, CDCl₃, 323 K): $\delta = 20.8$, 21.7, 52.3, 56.2, 70.6,

70.8, 107.5, 108.5, 110.7, 116.2, 116.8, 118.2 (br), 121.0 (br), 127.3, 132.5, 134.5, 136.8, 140.9, 142.4, 156.0, 158.2, 165.8, 166.6, 169.7, 169.8, 170.2, 185.0, 188.7 ppm (signals for four sp² carbon atoms were not detected presumably owing to the peak-broadening ascribed to the slow conformational interconversion, diaxial⇔diequatorial); IR (KBr): $\tilde{\nu}$ =2950, 1760, 1660, 1610, 1490, 1450, 1420, 1370, 1350, 1290, 1260, 1230, 1180, 1160, 1120, 1080, 1030, 970, 920, 810, 750 cm⁻¹; elemental analysis calcd (%) for C₃₀H₂₄O₁₂: C 62.50, H 4.20; found: C 62.80, H 4.47.

Benanomicinone methyl ester (**46**): A suspension of **44** (9.9 mg, 17 µmol) in 2 M aq. NaOH (5 mL) was heated at 70 °C for 2.5 h. After cooling to RT, the reaction mixture was acidified with 2 M aq. HCl. The precipitates were collected by filtration to afford crude material of carboxylic acid **45** as a deep-red powder (8.5 mg). D-Ala-OMe-HCl (12.4 mg, 88.8 µmol), BOP (112 mg, 25.3 µmol), and Et₃N (17 µL, 120 µmol) were added to a solution of crude **45** (8.5 mg) in DMF (0.4 mL) at RT. After stirring for 1.5 h, the reaction was stopped by adding 2 M aq. HCl and the reaction mixture was diluted with EtOAc. After separating the two layers, the organic layer was washed with 2 M aq. HCl and brine, dried (Na₂SO₄), and concentrated in vacuo. After adding Et₂O to the residue, the precipitate was collected by filtration to afford **46** as a red powder (6.1 mg, 63 %). The filtrate was concentrated in vacuo and the residue was purified by column chromatography (CHCl₃/MeOH=93:7) to afford **46** (2.8 mg, 17%; total yield: 80%).

45: M.p. > 230 °C; ¹H NMR (400 MHz, [D₆]DMSO, 313 K): δ =2.48 (s, 3 H), 3.94 (s, 3 H), 4.25 (d, *J*=10.5 Hz, 1 H), 4.32 (d, *J*=10.5 Hz, 1 H), 6.90 (d, *J*=2.3 Hz, 1 H), 7.08 (s, 1 H), 7.27 (d, *J*=2.3 Hz, 1 H), 8.08 (s, 1 H), 12.86 ppm (s, 1 H; OH); ¹³C NMR (100 MHz, [D₆]DMSO, 313 K): δ =21.7, 56.3, 71.6, 72.2, 106.6, 107.4, 110.1, 114.3, 115.1, 115.4, 117.8, 118.1, 125.7, 131.4, 134.6, 139.7, 143.6, 149.4, 155.7, 157.7, 164.6, 165.9, 171.4, 185.3, 187.0 ppm; IR (KBr): $\tilde{\nu}$ =3396, 2930, 2850, 1715, 1605, 1488, 1446, 1386, 1296, 1255, 1188, 1162, 1119, 1083, 1064, 1033, 983, 967, 822 cm⁻¹.

46: M.p. > 230 °C; ¹H NMR (400 MHz, [D₆]DMSO, 313 K): $\delta = 1.32$ (d, J = 7.3 Hz, 3H), 2.32 (s, 3H), 3.67 (s, 3H), 3.91 (s, 3H), 4.23 (d, J =10.0 Hz, 1 H), 4.25 (d, J=10.0 Hz, 1 H), 4.46 (qd, J=6.7, 7.3 Hz, 1 H), 5.95 (brs, 1H; OH), 6.15 (brs, 1H; OH), 6.86 (d, J=2.4 Hz, 1H), 7.07 (s, 1H), 7.19 (d, J = 2.4 Hz, 1H), 8.08 (s, 1H), 8.46 (br s, 1H), 8.68 (d, J =6.7 Hz, 1H; NH), 12.81 (s, 1H; OH), 13.84 ppm (brs, 1H; OH); ¹³C NMR (100 MHz, [D₆]DMSO, 313 K): $\delta = 16.7$, 19.0, 47.8, 51.7, 56.4, 71.4, 72.3, 106.8, 107.6, 110.0, 113.7, 115.3, 115.6, 117.5, 125.8, 127.1, 131.2, 134.2, 137.4, 141.0, 149.9, 150.8, 156.6, 164.8, 165.9, 173.0, 185.1, 187.4 ppm (signals for four sp² carbon atoms were not detected presumably owing to peak-broadening ascribed to the slow conformational interconversion, diaxial \rightleftharpoons diequatorial); IR (neat): $\tilde{\nu} = 3490, 3250, 1740, 1640,$ 1600, 1450, 1430, 1400, 1380, 1340, 1290, 1240, 1210, 1160, 1140, 1110, 1080, 1070, 1030, 1000, 980, 960, 920, 900, 880, 840, 810, 800, 790 cm⁻¹; elemental analysis calcd (%) for $C_{29}H_{25}NO_{11}$: C 61.81, H 4.47, N 2.49; found: C 61.51, H 4.71, N 2.29.

Benzo[a]naphthacene 48: 2-Methoxypropene (264 mg, 3.65 mmol) and TsOH-H₂O (21.5 mg, 0.113 mmol) were added to a solution of 46(205 mg, 0.364 mmol) in DMF (3 mL) at RT. After stirring 18 h at RT, the reaction was stopped by adding water and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with water (×3) and brine, dried (Na₂SO₄), and concentrated in vacuo. Ac₂O (0.8 mL) and zinc powder (40 mg, 0.764 mmol) were added to the solution of this crude material in pyridine (6 mL) at RT. After stirring for 10 h at this temperature, the reaction was stopped by adding MeOH and the mixture was extracted with EtOAc (×3). The combined organic extracts were successively washed with 2M aq. HCl (×3), water (×1), saturated aqueous NaHCO₃ (×1), and brine (×2), dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (benzene/acetone = 75:25) to afford pentacycle 47 as a yellow solid (218 mg, 73 %, two steps) contaminated with a trace amount of impurity (<1%, ¹H NMR) arising from the air sensitivity of this compound. This material was employed in the next experiment without further purification. ¹H NMR (400 MHz, CDCl₃): $\delta = 1.46$ (d, J = 7.1 Hz, 3H), 1.60 (s, 3H), 1.63 (s, 3H), 2.17 (s, 3H), 2.28 (s, 3H), 2.44 (s, 3H), 2.46 (s, 3H), 2.57 (s, 3H), 2.60 (s, 3 H), 3.78 (s, 3 H), 3.93 (s, 3 H), 4.54 (d, J = 10.7 Hz, 1 H), 4.60 (d, J =

9812

10.7 Hz, 1 H), 4.79 (qd, J = 7.1, 7.8 Hz, 1 H), 6.25 (d, J = 7.8 Hz, 1 H; NH), 6.93 (d, J = 2.4 Hz, 1 H), 7.09 (d, J = 2.4 Hz, 1 H), 7.29 (s, 1 H), 7.86 ppm (s, 1 H); ¹³C NMR (100 MHz, CDCl₃): δ = 18.3, 19.1, 20.7, 21.06, 21.14, 21.2, 27.28, 27.31, 48.0, 52.6, 55.5, 79.2, 79.7, 96.8, 112.5, 115.4, 116.0, 116.7, 120.8, 122.3, 124.6, 127.8, 131.0, 133.3, 138.1, 138.5, 138.7, 139.3, 141.3, 143.8, 146.2, 150.7, 157.5, 166.1, 168.3, 168.7, 169.0, 169.2, 169.8, 172.6 ppm.

TsOH·H₂O (36.5 mg, 0.192 mmol) was added to a solution of 47 (63.6 mg, 0.0780 mmol) in CH₃CN/H₂O (4:1) (3 mL) at RT. After stirring for 2 h at RT, the reaction was stopped by adding water, the mixture was extracted with EtOAc (×2), and washed with saturated aqueous NaHCO3 and water. The combined organic extracts were washed with brine, dried (Na2SO4), and concentrated in vacuo. The residue was purified by PTLC (benzene/acetone=1:1) to afford 48 as a yellow solid (56.5 mg, 93%). M.p. 190-191°C (hexane and EtOAc); ¹H NMR (400 MHz, $[D_6]$ acetone): $\delta = 1.46$ (d, J = 7.2 Hz, 3H), 2.21 (s, 3H), 2.31 (s, 3H), 2.44 (s, 3H), 2.46 (s, 3H), 2.61 (s, 3H), 2.71 (s, 3H), 3.76 (s, 3H), 3.99 (s, 3H), 4.36 (dd, J=4.8, 10.8 Hz, 1H), 4.44 (ddd, J=0.8 4.4, 10.8 Hz, 1H), 4.68 (qd, J=7.2, 7.2 Hz, 1H), 5.13(brs, 1H; OH), 5.26 (d, J=4.8 Hz, 1H; OH), 7.05 (d, J=0.8 Hz, 1H), 7.27 (s, 1H), 7.55 (s, 1H), 7.62 (d, J = 7.2 Hz, 1H; NH), 8.25 ppm (s, 1H); ¹³C NMR (100 MHz, $[D_6]$ acetone): $\delta = 17.6, 19.3, 20.7, 21.0, 21.16, 21.21, 21.3, 48.9, 52.3, 56.1,$ 73.6, 74.2, 97.2, 115.2, 116.5, 117.4, 120.9, 121.6, 123.9, 124.3, 125.3, 128.5, 132.2, 138.2, 138.4, 138.8, 140.2, 141.2, 142.5, 144.5, 147.7, 158.5, 166.8, 168.9, 169.1, 169.8, 169.9, 170.1, 173.5 ppm; IR (ATR): v=3490, 3340, 2940, 2860, 1760, 1640, 1530, 1450, 1430, 1420, 1360, 1240, 1180, 1120, 1020, 880 cm⁻¹; elemental analysis calcd (%) for $C_{39}H_{37}NO_{16}$: C 60.39, H 4.81, N 1.81; found: C 60.61, H 5.01, N 1.72.

β-Glycosides **49** and **50**: MS4A (300 mg) was placed in a two-necked round-bottomed flask and flame dried. The promoter was prepared in situ by stirring a mixture of Cp₂HfCl₂ (31.5 mg, 0.0830 mmol) and AgClO₄ (32.0 mg, 0.154 mmol) in CH₂Cl₂ (1.0 mL) for 10 min at RT. A solution of **48** (20.3 mg, 0.0261 mmol) and glycosyl fluoride **51** (28.3 mg, 0.0530 mmol) in CH₂Cl₂ (3.0 mL) was added to this suspension at -78° C. The reaction mixture was warmed to -12° C, stirred for 25 min, and the reaction was stopped by adding saturated aqueous NaHCO₃. The mixture was filtered through a Celite pad and extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (acetone/benzene=3:7) to afford β-glycoside **49** as a yellow amorphous solid (9.1 mg, 27%) and its regioisomeric β-glycoside **50** also as a yellow amorphous solid (5.1 mg, 17%).

49: M.p. 198.2–199.1 °C; $[\alpha]_D^{25} = -75.2$ (c = 0.874 in CHCl₃); ¹H NMR (500 MHz, CDCl₃): $\delta = 1.34$ (d, J = 6.3 Hz, 3H), 1.44 (d, J = 7.2 Hz, 3H), 1.62 (s, 3H), 1.97 (s, 3H), 2.01 (s, 3H), 2.11 (s, 3H), 2.26 (s, 3H), 2.42 (s, 6H), 2.49 (s, 3H), 2.57 (s, 3H), 3.33 (dd, J=9.3, 11.6 Hz, 1H), 3.66-3.72 (m, 1H), 3.78 (s, 3H), 3.89 (dd, J = 0.8, 3.6 Hz, 1H), 3.92 (s, 3H), 4.08 (dd, J=3.6, 10.0 Hz, 1 H), 4.14 (dd, J=5.2, 11.6 Hz, 1 H), 4.49 (br d, 11.2 Hz, 1H), 4.66 (d, J=7.2 Hz, 1H), 4.77 (qd, J=7.2, 7.7 Hz, 1H), 4.9016 (dd, J=7.2, 9.2 Hz, 1 H), 4.9025 (d, J=8.2 Hz, 1 H), 4.94 (ddd, J= 5.2, 9.1, 9.3 Hz, 1 H), 5.05 (dd, J=9.1, 9.2 Hz, 1 H), 5.68 (dd, J=8.2, 10.0 Hz, 1 H), 6.16 (d, J=7.7 Hz, 1 H; NH), 6.91 (d, J=2.3 Hz, 1 H), 7.05 (s, 1H), 7.50-7.67 (m, 4H), 7.98 (s, 1H), 8.08-8.13 ppm (m, 2H); ¹³C NMR (125 MHz, CDCl₃): $\delta = 17.2$, 18.2, 19.3, 19.9, 20.6, 20.7 (2 C), 20.98, 21.03, 21.1, 21.2, 48.0, 52.5, 55.5, 62.4, 65.6, 68.9, 69.7, 71.0, 71.3, 71.6, 72.8, 80.3, 81.7, 96.7, 102.2, 102.3, 114.8, 115.9, 116.5, 119.9, 120.3, 121.9, 124.5, 125.6, 127.8, 128.9, 129.2, 129.7, 131.1, 133.8, 134.6, 137.8, 138.1, 139.2, 139.5, 140.4, 143.1, 146.3, 157.5, 164.7, 166.2, 168.5, 168.6-169.3 (3C), 169.1, 169.6, 169.8, 170.2, 172.6; IR (ATR): $\tilde{v} = 3488$, 3369, 2941, 2111, 1742, 1667, 1638, 1520, 1450, 1433, 1361, 1247, 1175, 1116, 1025, 876, 713 cm⁻¹; elemental analysis calcd (%) for $C_{63}H_{64}N_4O_{27}$: C 57.80, H 4.93, N 4.28; found: C 57.60, H 5.21, N 4.05.

50: M.p. 191.5–192.5 °C; $[a]_D^{26} = -192$ (c = 0.705 in CHCl₃); ¹H NMR (500 MHz, CDCl₃): $\delta = 1.32$ (brs, 3H), 1.45 (d, J = 7.2 Hz, 3H), 1.66 (s, 3H), 1.97 (s, 3H), 2.01 (s, 3H), 2.16 (s, 3H), 2.20 (s, 3H), 2.39 (s, 3H), 2.49 (s, 3H), 2.56 (s, 3H), 2.77 (s, 3H), 3.32 (dd, J = 10.0, 11.2 Hz, 1H), 3.68 (brs, 1H), 3.77 (s, 3H), 3.91 (s, 3H), 3.94 (d, J = 3.6 Hz, 1H), 4.10 (dd, J = 3.6, 10.0 Hz, 1H), 4.13 (dd, J = 6.7, 11.2 Hz, 1H), 4.41–4.52 (m,

2 H), 4.64 (d, J=7.4 Hz, 1 H), 4.77 (qd, J=7.2, 7.5 Hz, 1 H), 4.86 (d, J=8.1 Hz, 1 H), 4.93 (dd, J=7.4, 9.2 Hz, 1 H), 4.95 (ddd, J=6.7, 9.2, 10.0 Hz, 1H), 5.06 (dd, J=9.2, 9.2 Hz, 1H), 5.70 (brs, 1H), 6.22 (d, J=7.5 Hz, 1H; NH), 6.92 (s, 1H), 7.03 (s, 1H), 7.38 (s, 1H), 7.53 (dd, J=7.4, 7.8 Hz, 2H), 7.65 (t, J=7.4 Hz, 1H), 8.09 (d, J=7.8 Hz, 2H), 8.22 ppm (brs, 1 H); ¹³C NMR (125 MHz, CDCl₃): $\delta = 17.2$, 18.3 (2 C), 19.2, 19.7, 20.0, 20.6, 20.7, 20.8, 21.0, 21.1, 21.2, 48.0, 52.5, 55.4, 62.6, 65.9, 69.0, 69.5, 71.1, 71.2, 71.7, 72.4, 72.8, 80.4, 96.6, 102.4, 103.0, 115.8, 116.7, 117.1, 119.9, 120.6, 121.6, 123.9, 124.6, 127.8, 129.0, 129.6, 131.0, 133.4, 133.9, 137.6, 137.9, 139.9, 140.0, 140.5, 143.2, 146.5, 157.4, 164.6, 166.1, 168.1, 168.9 (br), 169.0, 169.2, 169.7, 169.8, 170.0, 170.2, 172.7 ppm (signals for four sp² carbon atoms were not detected presumably owing to peak-broadening ascribed to the slow conformational interconversion, diaxial diequatorial); IR (ATR): v=3528, 3376, 2941, 2109, 1741, 1667, 1638, 1513, 1432, 1361, 1247, 1184, 1113, 1025, 876, 712 cm⁻¹; elemental analysis calcd (%) for C63H64N4O27: C 57.80, H 4.93, N 4.28; found: C 58.01, H 5.17. N 4.01.

Benanomicin B hydrochloride (2 b·HCl): A 2 M aq. HCl solution (100 μL) and 5% Pd/C (30 mg) were added to a solution of 49 (37.0 mg, 0.0283 mmol) in MeOH (5.0 mL). After stirring under H₂ (1 atm) at RT for 6 h, the reaction mixture was filtered though a Celite pad (washed with MeOH) and the filtrate was concentrated in vacuo. A 1 M aq. NaOH solution (5 mL) was added to a solution of this crude material (36.0 mg) in MeOH (5.0 mL). After stirring for 2 h, the reaction was stopped by adding 2M aq. HCl (ca. 3.0 mL) and the mixture concentrated in vacuo (azeotropic evaporation ×5). The residue was purified by Sephadex LH-20 (DMF) to afford a dimethyformamide solvate of benanomicin B which was dissolved in MeOH and then 2M aq. HCl was added until pH 3.5. After removal of the volatile material in vacuo, the residue was dissolved in DMSO (0.05 mL) and CHCl₃ (ca. 6.0 mL) was added. The precipitate was collected by filtration to afford benanomicin B hydrochloride (16.7 mg, 68%67% given in legend to Scheme 12.) as a red amorphous powder. M.p. 213-215 °C (decomp) (lit.:^[1b] > 220 °C (decomp)); $[\alpha]_{D}^{22} = +353$ (c=0.0501 in H₂O) (lit.:^[1b] +360 (c=0.05, H₂O)); ¹H NMR (500 MHz, [D₆]DMSO, 313 K): $\delta = 1.18$ (d, J = 6.5 Hz, 3H), 1.34 (d, J=7.3 Hz, 3H), 2.33 (s, 3H), 3.07 (dd, J=10.7, 10.9 Hz, 1H), 3.16-3.18 (m, 3H), 3.29-3.34 (m, 1H), 3.39-3.44 (m, 1H), 3.60 (br, 1 H), 3.72 (dd, J=5.3, 11.3 Hz, 1 H), 3.83 (br q, J=6.5, 1 H), 3.91–3.94 (m, 1 H), 3.95 (s, 3 H), 4.42 (dq, J=7.1, 7.3 Hz, 1 H), 4.53–4.56 (m, 2 H), 4.58– 4.65 (m, 1H), 4.74 (d, J=7.6 Hz, 1H), 6.94 (d, J=2.4 Hz, 1H), 7.23 (brs, 1 H), 7.31 (d, J=2.4 Hz, 1 H), 8.06 (s, 1 H), 8.43 (d, J=7.1 Hz, 1 H), 12.8 ppm (s, 1 H); ¹³C NMR (125 MHz, $[D_6]$ DMSO, 313 K): $\delta = 16.3$, 16.9, 19.1, 47.6, 54.2, 56.4, 65.7, 66.9, 69.4, 69.7, 73.3, 75.9, 77.4, 80.9, 104.0, 104.5, 106.9, 107.6, 110.1, 113.7, 115.6, 125.7, 127.5, 131.3, 134.3, 137.3, 137.8, 147.9, 151.0, 156.9, 164.7, 166.0, 166.8, 173.9, 185.0, 187.5 ppm; IR (KBr): $\tilde{\nu} = 3360$, 1730, 1610, 1300, 1160, 1080, 1040 cm⁻¹; LRMS (MALDI-TOF, DHBA matrix): m/z: calcd for C₃₉H₄₂O₁₈N₂Na ([M+Na]⁺): 849.2; found: 849.2; HRMS (FAB): m/z: calcd for C₃₉H₄₃O₁₈N₂ ([M+H]⁺): 827.2511; found: 827.2506.

Second-generation synthesis

 $\label{eq:2-methoxy} 2-methoxy-6-methyl-4-[(triisopropylsiloxy)methyl] ben-$ Methvl zoate: Imidazole (17.3 g, 0.254 mol) and triisopropylsilyl chloride (28.2 mL, 0.133 mol) were added to a solution of 11 (30.5 g, 0.127 mol) in DMF (130 mL) at 0°C. After stirring for 2 h at RT, the reaction was stopped by adding water and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=4:1 to 3:1, gradient elution) to afford methyl 2-methoxymethoxy-6-methyl-4-[(triisopropylsiloxy)methyl]benzoate as a colorless oil (50.5 g, quant). ¹H NMR (400 MHz, CDCl₃): $\delta = 1.05-1.25$ (m, 21 H), 2.29 (s, 3 H), 3.46 (s, 3 H), 3.90 (s, 3 H), 4.79 (s, 2H), 5.16 (s, 2H), 6.83 (s, 1H), 7.04 ppm (s, 1H); ¹³C NMR (100 MHz, $CDCl_3$): $\delta = 12.1$, 18.1, 19.5, 52.1, 56.0, 64.5, 94.7, 109.6, 120.4, 123.0, 136.2, 144.4, 154.1, 168.6 ppm; IR (neat): $\tilde{\nu} = 3000$, 2890, 2870, 1730, 1610, 1590, 1460, 1440, 1270, 1160, 1110, 1090, 1050, 1000, 930, 880, 810 cm⁻¹; elemental analysis calcd (%) for C₂₁H₃₆O₅Si: C 63.60, H 9.15; found: C 63.50, H 9.08.

Methyl 2-hydroxy-6-methyl-4-[(triisopropylsiloxy)methyl]benzoate: Tri-fluoroacetic acid (8.8 mL, 0.12 mol) was added to a solution of methyl 2-methoxymethoxy-6-methyl-4-[(triisopropylsiloxy)methyl]benzoate

(30.4 g, 76.5 mmol) in CH₂Cl₂ (300 mL) at 0 °C. After stirring for 3.5 h at 0°C and then for 2.5 h at RT, the reaction was stopped by adding saturated aqueous NaHCO₃ and the mixture was extracted with EtOAc (\times 3). The combined organic extracts were washed with brine, dried (Na_2SO_4) , and concentrated in vacuo. After adding Et₂O and hexane to the residue, the precipitate was collected by filtration to afford methyl 2-hydroxy-6methyl-4-[(triisopropylsiloxy)methyl]benzoate (1st crop: 22.8 g, 85 %; 2nd crop: 1.50 g, 5.5%) as a white solid. The filtrate was concentrated in vacuo and the residue was purified by flash column chromatography (silica gel, hexane/EtOAc=95:5) to afford methyl 2-hydroxy-6-methyl-4-[(triisopropylsiloxy)methyl]benzoate as a white solid (1.40 g, 5.2%; total yield: 96%). M.p. 56.5-57.2°C (hexane and Et₂O); ¹H NMR (400 MHz, CDCl₃): $\delta = 1.06 - 1.27$ (m, 21 H), 2.53 (s, 3 H), 3.95 (s, 3 H), 4.76 (s, 2 H), 6.67 (s, 1 H), 6.87 (s, 1 H), 11.37 ppm (s, 1 H; OH); ¹³C NMR (100 MHz, $CDCl_3$): $\delta = 12.1, 18.1, 24.3, 52.0, 64.3, 110.5, 112.2, 119.9, 141.0, 148.6,$ 163.0, 172.1; IR (ATR): $\tilde{\nu}$ =2950, 2890, 2860, 1660, 1630, 1580, 1450, 1420, 1370, 1320, 1280, 1220, 1160, 1110, 1060, 1010, 990, 950, 880, 850, 800 cm⁻¹; elemental analysis calcd (%) for $C_{19}H_{32}O_4Si$: C 64.73, H 9.15; found: C 64.93, H 9.39.

Methyl 2-hydroxy-3-iodo-6-methyl-4-[(triisopropylsiloxy)methyl]benzoate (53): NaHCO₃ (2.39 g, 28.4 mmol), BnMe₃N⁺ICl₂⁻ (4.99 g, 14.3 mmol), and MeOH (35 mL) were added to a solution of methyl 2-hydroxy-6methyl-4-[(triisopropylsiloxy)methyl]benzoate (5.00 g, 14.2 mmol) in CH₂Cl₂ (90 mL). After stirring for 4 h at RT, the reaction was stopped by adding aqueous $Na_2S_2O_3$ and the mixture was extracted with EtOAc (× 3). The combined organic extracts were washed with brine, dried (Na_2SO_4) , and concentrated in vacuo. The residue was passed through a silica gel column (hexane/EtOAc=95:5) to afford somewhat impure 53 with trace amounts of by-products. The mixture was further purified by washing with MeOH to give a pure form of 53 as a white amorphous solid (5.77 g, 85%). The filtrate was concentrated in vacuo and the residue was further purified by flash column chromatography (silica gel, hexane/benzene=1:1) to afford an additional product of 53 as a white amorphous solid (0.650 g, 9.6%; total yield: 95%). M.p. 33.9-34.1°C (hexane); ¹H NMR (400 MHz, CDCl₃): $\delta = 1.06-1.24$ (m, 21 H), 2.53 (s, 3H), 3.96 (s, 3H), 4.68 (s, 2H), 7.05 (s, 1H), 12.3 (s, 1H); ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3): \delta = 12.0, 18.1, 24.2, 52.5, 70.0, 83.4, 110.8, 121.6, 141.2,$ 149.9, 160.4, 171.8 ppm; IR (KBr): $\tilde{\nu} = 2940$, 2890, 2870, 1660, 1600, 1540, 1470, 1450, 1390, 1360, 1340, 1280, 1240, 1200, 1180, 1120, 1060, 980, 950, 880, 820 cm⁻¹; elemental analysis calcd (%) for $C_{19}H_{31}IO_4Si$: C 47.70, H 6.53; found: C 47.70, H 6.56.

Ethyl 7-chloro-4-hydroxy-5,8-dimethoxy-2-naphthoate: K₂CO₃ (33.9 g, 0.245 mol) was added to a solution of **31** (28.8 g, 81.7 mmol) in EtOH (350 mL) at RT. The reaction mixture was warmed to 60 °C and heated at this temperature with stirring for 1.5 h. After cooling to 0°C, the reaction mixture was diluted with water and then the reaction was stopped by adding 6M aq. HCl. The mixture was filtered through a Celite pad (washed with EtOAc) and extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. After adding Et₂O and hexane to the residue, the precipitate was collected by filtration to afford the title compound as a white crystalline solid (22.2 g, 88%). The filtrate was concentrated in vacuo and the residue was purified by flash column chromatography (silica gel, hexane/ EtOAc=7:3) to afford further ethyl 7-chloro-4-hydroxy-5,8-dimethoxy-2naphthoate as a white crystalline solid (2.84 g, 11%; total yield: 99%). M.p. 130–131 °C (hexane and EtOAc); ¹H NMR (400 MHz, CDCl₃): $\delta =$ 1.43 (t, J=7.1 Hz, 3H), 3.97 (s, 3H), 4.06 (s, 3H), 4.43 (q, J=7.1 Hz, 2H), 6.82 (s, 1H), 7.49 (d, J=1.7 Hz, 1H), 8.29 (d, J=1.7 Hz, 1H), 9.22 ppm (s, 1H; OH); ¹³C NMR (100 MHz, CDCl₃): $\delta = 14.3$, 56.7, 61.3, 61.5, 107.6, 110.9, 115.5, 116.6, 123.2, 130.6, 130.8, 147.4, 152.3, 155.0, 166.3 ppm; IR (KBr): \tilde{v} = 3560, 3100, 3030, 2990, 2950, 2900, 2850, 1720, 1600, 1520, 1460, 1390, 1340, 1310, 1270, 1230, 1210, 1160, 1120, 1090, 1030, 1020, 970, 940, 870, 840, 810, 770 cm⁻¹; elemental analysis calcd (%) for C₁₅H₁₅O₅Cl: C 57.98, H 4.87; found: C 58.0, H 5.03.

Ethyl 7-chloro-5,8-dimethoxy-4-methoxymethoxy-2-naphthoate: 7-Chloro-4-hydroxy-5,8-dimethoxy-2-naphthoate (25.7 g, 82.8 mmol) and MOMCl (9.43 mL, 0.124 mmol) were added to a suspension of NaH (55% dispersion in mineral oil, washed with hexane (4.69 g, 0.108 mol)) in DMF (830 mL) at 0°C. After stirring for 1 h, the reaction was stopped by adding water and the resulting mixture was extracted with Et_2O (×3). The combined organic extracts were washed with brine, dried (MgSO₄), and concentrated in vacuo. After adding Et₂O and hexane to the residue, the precipitate was collected by filtration to afford the title compound as a white powder (1st crop: 14.8 g, 51 %; 2nd crop: 8.94 g, 30 %; 3rd crop: 4.39 g, 15%; 4th crop: 1.17 g, 4%; total yield: quant). M.p. 119.7-120.0°C (hexane and EtOAc); ¹H NMR (400 MHz, CDCl₃): $\delta = 1.44$ (t, J = 8.1 Hz, 3 H), 3.60 (s, 3 H), 3.95 (s, 3 H), 3.98 (s, 3 H), 4.44 (q, J=8.1 Hz, 2 H), 5.31 (s, 2H), 6.88 (s, 1H), 7.62 (d, J=1.6 Hz, 1H), 8.49 ppm (d, J=1.6 Hz, 1 H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 14.4$, 56.6, 56.7, 61.3, 61.6, 96.4, 109.6, 112.0, 118.8, 120.1, 123.9, 129.3, 131.3, 146.3, 153.4, 154.5, 166.2 ppm; IR (KBr): $\tilde{\nu}$ = 3120, 3070, 2980, 2830, 1730, 1590, 1510, 1460, 1390, 1330, 1270, 1230, 1180, 1150, 1140, 1050, 1030, 930, 820, 770 cm⁻¹; elemental analysis calcd (%) for C₁₇H₁₉O₆Cl: C 57.55, H 5.40; found: C 57.35, H 5.55.

7-Chloro-5,8-dimethoxy-4-methoxymethoxy-2-naphthoic acid (52): А 6м aq. NaOH solution (330 mL) was added to a suspension of ethyl 7chloro-5,8-dimethoxy-4-methoxymethoxy-2-naphthoate (35.0 g, 98.5 mmol) in THF (450 mL) and EtOH (180 mL) and the reaction mixture was heated at 70 °C for 1 h. After cooling to RT, the reaction mixture was diluted with water and then the aqueous layer was washed with Et_2O (×3). The aqueous phase was acidified with 6M aq. HCl. The crystalline precipitates were collected by filtration and dried to afford 52 as a white powder (32.1 g, quant). M.p. 192-193 °C (Et₂O and EtOAc); ¹H NMR (400 MHz, [D₆]DMSO): $\delta = 3.32$ (s, 3H), 3.87 (s, 3H), 3.99 (s, 3H), 5.29 (s, 2H), 7.08 (s, 1H), 7.53 (d, J=1.5 Hz, 1H), 8.28 (d, J= 1.5 Hz, 1H), 13.3 ppm (brs, 1H); 13 C NMR (100 MHz, [D₆]DMSO): $\delta =$ 56.0, 56.7, 61.3, 95.7, 109.3, 111.1, 117.3, 119.1, 123.4, 129.8, 130.5, 145.2, 153.4, 154.6, 166.8 ppm; IR (ATR): $\tilde{\nu} = 2960, 2930, 2910, 2840, 1640, 2540,$ 1700, 1590, 1510, 1460, 1440, 1370, 1330, 1280, 1230, 1160, 1130, 1090, 1030, 990, 970, 920, 910, 890, 800, 770, 710 cm⁻¹; elemental analysis calcd (%) for C₁₅H₁₅O₆Cl: C 55.14, H 4.63; found: C 54.91, H 4.86.

2-Iodo-6-methoxycarbonyl-5-methyl-3-[(triisopropylsiloxy)methyl]phenyl 7-chloro-5,8-dimethoxy-4-methoxymethoxy-2-naphthoate: Et₃N (8.45 mL, 60.6 mmol) and 2,4,6-trichlorobenzoyl chloride (9.47 mL, 60.6 mmol) were added to a solution of carboxylic acid 52 (18.0 g, 55.1 mmol) in toluene (540 mL) at RT. After stirring for 3 h at RT, iodophenol 53 (27.7 g, 57.9 mmol) and DMAP (10.2 g, 82.7 mmol) were added to the reaction mixture which was then stirred for 20 min. The reaction was stopped by adding water and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. After adding Et₂O and hexane to the residue, the precipitate was collected by filtration to afford the title compound as a yellow solid (40.1 g, 95%). The filtrate was concentrated in vacuo and the residue was purified by flash column chromatography (silica gel, hexane/EtOAc=9:1 to 7:3, gradient elution) to afford further 2-iodo-6methoxycarbonyl-5-methyl-3-(triisopropylsiloxymethyl)phenyl 7-chloro-5,8-dimethoxy-4-methoxymethoxy-2-naphthoate as a yellow solid (1.67 g, 4%; total yield: 99%). M.p. 156–157°C (hexane and Et_2O); ¹H NMR (400 MHz, CDCl₃): $\delta = 1.08-1.28$ (m, 21 H), 2.47 (s, 3 H), 3.62 (s, 3 H), 3.70 (s, 3H), 3.97 (s, 3H), 4.00 (s, 3H), 4.75 (s, 2H), 5.34 (s, 2H), 6.93 (s, 1H), 7.46 (s, 1H), 7.75 (s, 1H), 8.72 (s, 1H); ¹³C NMR (100 MHz, $CDCl_3$): $\delta = 12.1, 18.1, 20.4, 52.3, 56.7, 56.8, 61.7, 69.7, 89.6, 96.4, 110.2,$ 112.0, 120.0, 120.5, 124.2, 125.6, 126.9, 127.7, 131.3, 138.5, 146.3, 146.8, 148.5, 153.4, 154.8, 163.4, 166.0 ppm; IR (KBr): $\tilde{\nu} = 2940$, 2890, 2860, $1730,\,1590,\,1460,\,1370,\,1340,\,1260,\,1190,\,1150,\,1120,\,1080,\,1020,\,980,\,920,$ 880, 820, 800, 750 cm⁻¹; elemental analysis calcd (%) for $C_{34}H_{44}ClO_9Si: C$ 51.88, H 5.63; found: C 52.08, H 5.64.

2-Iodo-6-methoxycarbonyl-5-methyl-3-[(triisopropylsiloxy)methyl]phenyl 7-chloro-4-hydroxy-5,8-dimethoxy-2-naphthoate (54): Trifluoroacetic acid (9.4 mL, 0.122 mol) was added to a solution of 2-iodo-6-methoxycarbonyl-5-methyl-3-[(triisopropylsiloxy)methyl]phenyl 7-chloro-5,8-dimethoxy-4-methoxymethoxy-2-naphthoate (32.0 g, 40.7 mmol) in CH_2Cl_2 (410 mL)

9814 -

at 0°C. After stirring for 3.5 h at RT, the reaction was stopped by adding saturated aqueous NaHCO3 and water, and then the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na2SO4), and concentrated in vacuo. After adding Et2O and hexane to the residue, the precipitate was collected by filtration to afford phenol 54 as a white crystalline (21.2 g; 70%). The filtrate was concentrated in vacuo, and the residue was purified by flash column chromatography (silica gel, hexane/EtOAc=4:1) to afford crude material that included 54. Addition of Et₂O and hexane allowed the formation of a precipitate that was collected by filtration to afford 54 as a white solid (1st crop; 4.39 g, 15%; 2nd crop; 4.32 g, 14%; total yield: 99%). M.p. 139.5-140.0 °C (hexane and Et₂O); ¹H NMR (400 MHz, CDCl₃): $\delta = 1.09-1.30$ (m, 21 H), 2.46 (s, 3 H), 3.70 (s, 3 H), 4.00 (s, 3 H), 4.08 (s, 3 H), 4.75 (s, 2H), 6.87 (s, 1H), 7.45 (s, 1H), 7.63 (s, 1H), 8.50 (s, 1H), 9.30 ppm (s, 1H; OH); 13 C NMR (100 MHz, CDCl₃): $\delta = 12.1$, 18.1, 20.4, 52.3, 56.8, 61.6, 69.7, 89.6, 108.1, 111.0, 116.7, 117.0, 123.5, 125.6, 126.9, 129.0, 130.8, 138.5, 146.8, 147.3, 148.6, 152.2, 155.2, 163.5, 166.0 ppm; IR (KBr): $\tilde{\nu} =$ 3394, 2943, 2866, 1736, 1603, 1514, 1448, 1387, 1340, 1265, 1205, 1151, 1119, 1078, 1026, 978, 951, 922, 883, 845, 820, 798, 756 cm⁻¹; elemental analysis calcd (%) for $C_{32}H_{40}CIO_8Si$: C 51.72, H 5.43; found: C 51.44, H 5.31.

Biaryl lactone 55: A suspension of ester 54 (1.50 g, 2.02 mmol), Pd(OAc)₂ (136 mg, 0.604 mmol), PPh₃ (317 mg, 1.21 mmol), and sodium pivalate (751 mg, 6.05 mmol) in DMA (100 mL) was heated at 110 °C for 20 min. After cooling the mixture to 0°C, the reaction was stopped by adding water. The crystalline precipitates were collected by filtration and washed with Et_2O . The resulting powder was dissolved in CH_2Cl_2 and filtered through a Celite pad. The filtrate was concentrated in vacuo to afford 55 as yellow solid (747 mg, 60 %). M.p. 233.5–234.2 $^{\circ}\mathrm{C}$ (CH_2Cl_2 and hexane); ¹H NMR (500 MHz, CDCl₃): $\delta = 0.91-1.08$ (m, 21 H), 2.45 (s, 3H), 4.01 (s, 3H), 4.02 (s, 3H), 4.14 (s, 3H), 4.93 (s, 2H), 6.96 (s, 1H), 7.51 (s, 1 H), 8.61 (s, 1 H), 10.2 ppm (s, 1 H); ¹³C NMR (125 MHz, CDCl₃): $\delta = 11.9, 17.9, 19.6, 52.6, 57.1, 61.9, 65.1, 109.1, 113.0, 114.8, 116.3, 116.8,$ 120.9, 123.6, 124.1, 125.6, 129.9, 136.5, 142.2, 146.8, 147.8, 151.4, 152.1, 160.5, 167.2 ppm; IR (KBr): $\tilde{\nu} = 3310$, 2960, 2860, 1750, 1610, 1460, 1390, 1330, 1260, 1210, 1150, 1090, 1060, 1040, 950, 880, 830, 810 cm⁻¹; elemental analysis calcd (%) for $C_{32}H_{39}ClO_8Si$: C 62.48, H 6.39; found: C 62.21, H 6.26; HRMS (FAB): m/z: calcd for $C_{32}H_{40}O_8ClSi$ ([*M*+H]⁺): 615.2181; found: 615.2199.

Amide alcohol **57c**: (*R*)-Valinol (0.875 g, 8.48 mmol) was added to a solution of **55** (2.01 g, 3.27 mmol) in CH₂Cl₂ (20 mL) at 0 °C. After stirring for 43 h at 0 °C, the reaction was stopped by adding water and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=4:6) to afford a diastereomeric mixture of **57c** as a yellow solid (2.11 g, 90%, R,M:R,P=10:1). Recrystallization from hexane/EtOAc gave pure (R,M)-**57c** (1.85 g, 79%) as a yellow powder. The filtrate was concentrated in vacuo and the residue was further purified by PTLC (hexane/EtOAc=4:6) to afford (R,M)-**57c** (0.187 g, 8%) as a yellow amorphous solid.

(*R*,*M*)-**57 c**: M.p. 184–185 °C (hexane and EtOAc); $[a]_{D}^{25}$ =+58.0 (*c*= 0.994 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ =0.63 (d, *J*=6.8 Hz, 3H), 0.67 (d, *J*=6.8 Hz, 3H), 0.78–1.12 (m, 21 H), 1.59 (dsept, *J*=6.8, 6.8 Hz, 1H), 2.47 (t, *J*=5.6 Hz, 1H; OH), 2.62 (s, 3H), 3.50–3.55 (m, 2H), 3.61–3.72 (m, 1H), 3.96 (s, 3H), 3.99 (s, 3H), 4.01 (s, 3H), 4.44 (d, *J*=14.4 Hz, 1H), 4.58 (d, *J*=14.4 Hz, 1H), 6.35 (d, *J*=8.0 Hz, 1H; NH), 6.79 (s, 1H), 7.18 (s, 1H), 7.99 (s, 1H), 9.55 (s, 1H; OH), 11.76 ppm (s, 1H; OH); ¹³C NMR (100 MHz, CDCl₃): δ =12.0, 18.0, 18.4, 18.9, 24.4, 28.9, 52.3, 56.7, 57.9, 61.5, 63.1, 64.1, 106.9, 111.1, 113.6, 114.5, 114.8, 119.7, 121.5, 123.2, 130.6, 138.0, 141.3, 146.9, 147.3, 151.5, 152.2, 159.4, 169.2, 172.2 ppm; IR (KBr): $\tilde{\nu}$ =3360, 2940, 2890, 2870, 1660, 1620, 1600, 1520, 1460, 1400, 1360, 1290, 1270, 1200, 1180, 1130, 1110, 1040, 1000, 950, 880, 810 cm⁻¹; elemental analysis calcd (%) for C₃₇H₃₂ClNO₉Si: C 61.86, H 7.30, N 1.95; found: C 62.14, H 7.45, N 1.86; HRMS (FAB): m/z: calcd for C₃₇H₃₃O₉NClSi ([*M*+H]⁺): 718.3179; found: 718.3164.

FULL PAPER

(*R*,*P*)-**57 c**: $[a]_{27}^{27}$ = −26.8 (*c* = 1.09 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ = 0.80 (d, *J* = 6.8 Hz, 3H), 0.85 (d, *J* = 6.8 Hz, 3H), 0.92–1.08 (m, 21H), 1.54–1.62 (m, 1H), 1.92 (brs, 1H; OH), 2.64 (s, 3H), 3.12–3.29 (m, 2H), 3.68–3.73 (m, 1H), 3.97 (s, 3H), 3.99 (s, 3H), 4.01 (s, 3H), 4.42 (d, *J* = 15.2 Hz, 1H), 4.55 (d, *J* = 15.2 Hz, 1H), 6.35 (d, *J* = 8.8 Hz, 1H; NH), 6.80 (s, 1H), 7.23 (s, 1H), 7.95 (s, 1H), 9.56 (s, 1H; OH), 11.83 ppm (s, 1H; OH); ¹³C NMR (100 MHz, CDCl₃): δ = 11.9, 17.9, 18.2, 19.3, 24.4, 28.7, 52.3, 56.7, 57.6, 61.4, 62.8, 63.8, 106.8, 110.8, 113.0, 114.3, 114.7, 119.5, 121.4, 123.2, 130.6, 138.1, 141.3, 146.8, 147.7, 151.6, 152.2, 159.1, 169.1, 172.4 ppm; IR (neat): $\tilde{\nu}$ = 3380, 2960, 2940, 2870, 1660, 1610, 1600, 1520, 1450, 1410, 1360, 1290, 1260, 1200, 1130, 1100, 1040, 990, 950, 880, 810, 750 cm⁻¹; elemental analysis calcd (%) for C₃₇H₅₂ClNO₉Si: C 61.86, H 7.30, N 1.95; found: C 61.58, H 7.20, N 1.67; HRMS (FAB): m/z: calcd for C₃₇H₅₃O₉NClSi ([*M*+H]⁺): 718.3179; found: 718.3184.

Benzyl ether (R,M)-65: BnBr (0.59 mL, 4.93 mmol) and Cs₂CO₃ (2.66 g, 8.15 mmol) were added to a solution of (R,M)-57 c (1.48 g, 2.06 mmol) in DMF (15 mL) at 0 °C. After stirring for 1 h, the reaction was stopped by adding water and N,N-dimethylpropanediamine (0.5 mL), and the mixture was extracted with Et₂O (×3). The combined organic extracts were washed with brine, dried (Na2SO4), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/ EtOAc=65:35) to afford (R,M)-65 as a yellow oil (1.79 g, 97%). $[\alpha]_{D}^{27}$ = -80.7 (c = 0.976 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): $\delta = 0.69$ (d, J = 6.8 Hz, 3H), 0.73 (d, J=6.8 Hz, 3H), 0.88-1.05 (m, 21H), 1.64-1.70 (m, 1H), 2.41 (s, 3H), 2.62 (brs, 1H; OH), 3.40-3.50 (m, 2H), 3.59-3.63 (m, 1H), 3.75 (s, 3H), 3.76 (s, 3H), 4.00 (s, 3H), 4.59-4.77 (m, 6H), 6.23 (d, J=6.8 Hz, 1 H; NH), 6.85 (s, 1 H), 6.96–7.00 (m, 4 H), 7.10–7.11 (m, 3 H), 7.25–7.26 (m, 3 H), 7.32 (s, 1 H), 8.47 ppm (s, 1 H); $^{13}\mathrm{C}\,\mathrm{NMR}$ (100 MHz, $CDCl_3$): $\delta = 11.9, 17.87, 17.93, 18.8, 19.0, 19.6, 28.9, 52.3, 56.4, 58.5, 61.7,$ 63.7, 63.8, 75.8, 75.9, 109.0, 119.8, 120.9, 123.9, 124.0, 125.1, 126.2, 127.2, 127.3, 127.4, 127.5, 127.8, 128.0, 131.2, 136.6, 136.9, 137.1, 137.5, 143.0, 146.1, 152.5, 152.7, 153.6, 168.2, 168.3 ppm; IR (neat): $\tilde{\nu} = 3420$, 2940, 2870, 1730, 1660, 1600, 1570, 1520, 1460, 1340, 1280, 1060, 1040, 740, 700 cm⁻¹; elemental analysis calcd (%) for C₅₁H₆₄ClNO₉Si: C 68.17, H 7.18, N 1.56; found: C 68.17, H 7.33, N 1.36; HRMS (FAB): m/z: calcd for C₅₁H₆₅O₉NClSi ([*M*+H]⁺): 898.4117; found: 898.4126.

Oxazoline (R,M)-66: I₂ (1.17 g, 4.62 mmol) was added to a solution of (R,M)-65 (2.06 g, 2.30 mmol), imidazole (390 mg, 5.73 mmol), and PPh₃ (1.53 g, 5.83 mmol) in CH₂Cl₂ (80 mL). After stirring for 20 min, the reaction was stopped by adding saturated aqueous Na2S2O3 and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na2SO4), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/ $Et_2O/CHCl_3 = 70:15:15$) to afford oxazoline (R,M)-66 as a yellow oil (2.02 g, quant). $[\alpha]_D^{28} = -58.6$ (c = 1.06 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): $\delta = 0.76$ (d, J = 6.8 Hz, 3 H), 0.76 (d, J = 6.8 Hz, 3 H), 0.93–1.05 (m, 21 H), 1.52-1.56 (m, 1 H), 2.39 (s, 3 H), 3.72 (s, 3 H), 3.76 (s, 3 H), 3.80-3.87 (m, 2H), 3.99 (s, 3H), 4.18-4.25 (m, 1H), 4.54 (d, 14.4 Hz, 1H), 4.58 (d, 14.4 Hz, 1 H), 4.59 (d, J=11.2 Hz, 1 H), 4.63 (d, J=10.0 Hz, 1 H) 4.69 (d, J=11.2 Hz, 1 H), 4.73 (d, J=10.0 Hz, 1 H), 6.83-6.85 (m, 3 H), 6.96-7.05 (m, 5H), 7.17-7.24 (m, 3H), 7.34 (s, 1H), 8.44 ppm (s, 1H); ¹³C NMR (100 MHz, CDCl₃): $\delta = 12.0$, 18.0, 18.6, 18.9, 19.8, 33.3, 51.9, 56.6, 61.3, 63.1, 70.2, 73.5, 75.6, 75.9, 109.1, 120.0, 121.4, 122.6, 123.5, 126.0, 126.3, 126.7, 127.0, 127.1, 127.2, 127.7, 127.9, 128.0, 128.5, 130.7, 135.5, 137.6, 137.7, 143.8, 145.9, 152.7, 153.3, 153.6, 161.8, 169.1 ppm; IR (neat): $\tilde{\nu} = 2940, 2870, 1730, 1660, 1600, 1570, 1450, 1400, 1370, 1340,$ 1280, 1230, 1140, 1100, 1060, 1040, 950, 880, 810, 750, 700 cm⁻¹; elemental analysis calcd (%) for C51H62CINO8Si: C 69.56, H 7.10, N 1.59; found: C 69.66, H 7.37, N 1.53; HRMS (FAB): m/z: calcd for C₅₁H₆₃O₈NClSi ([M+H]⁺): 880.4011; found: 880.4003.

Aldehyde (M)-67: 2,6-Di-tert-butylpyridine ($325 \ \mu$ L, 1.48 mmol) and MeOTf ($160 \ \mu$ L, 1.46 mmol) were added to a solution of (*R*,*M*)-66 (434 mg, 0.493 mmol) in CH₂Cl₂ (4.5 mL) at RT. The reaction mixture was stirred for 1 h at RT before L-Selectride ($1.0 \ m$ in THF, 1.97 mL, 1.97 mmol) was added at 0 °C and the mixture was then stirred for an additional 20 min at 0 °C. The reaction was stopped by adding water and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in

Chem. Eur. J. 2007, 13, 9791-9823

© 2007 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

vacuo. Acidic silica gel (5 g, pH 6, Kanto Chemical) was added to the residue and organic solvents were removed by evaporation. After kept standing for 17 h, the SiO2 was placed on a glass filter and washed with EtOAc, and then the filtrate was concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc= 7:3) to afford aldehyde (M)-67 as a yellow powder (379 mg, 96%). M.p. 144–145 °C (hexane and EtOAc); $[\alpha]_{D}^{29} = -92.2$ (c=1.07 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): $\delta = 0.76 - 1.01$ (m, 21 H), 2.46 (s, 3 H), 3.79 (s, 3H), 3.81 (s, 3H), 4.03 (s, 3H), 4.40 (d, J=13.2 Hz, 1H), 4.57 (d, J= 10.8 Hz, 1 H), 4.62 (d, $J\!=\!13.2$ Hz, 1 H), 4.65 (d, $J\!=\!10.8$ Hz, 1 H), 4.72 (s, 2H), 6.81-6.85 (m, 2H), 6.95 (s, 1H), 7.01-7.12 (m, 6H), 7.26-7.30 (m, 2 H), 7.38 (s, 1 H), 8.57 (s, 1 H), 9.82 ppm (s, 1 H); $^{13}\mathrm{C}\,\mathrm{NMR}$ (100 MHz, $CDCl_3$): $\delta = 11.9, 17.9, 19.8, 52.2, 56.6, 61.9, 63.5, 75.8, 76.2, 110.5, 119.4,$ 123.0, 123.2, 124.1, 124.8, 127.1, 127.2, 127.4, 127.48, 127.51, 127.8, 127.9, 128.1, 130.9, 133.5, 136.7, 137.2, 137.3, 143.3, 147.0, 152.7, 153.7, 153.9, 168.5, 191.4 ppm; IR (KBr): $\tilde{\nu} = 2950$, 2940, 2860, 1730, 1700, 1600, 1570, 1460, 1400, 1360, 1340, 1280, 1210, 1100, 1060, 990, 970, 940, 910, 880, 810, 740, 700 cm⁻¹; elemental analysis calcd (%) for $C_{46}H_{53}ClO_8Si$: C 69.28, H 6.70; found: C 69.09, H 6.95; HRMS (FAB): m/z: calcd for C₄₆H₅₄O₈ClSi ([*M*+H]⁺): 797.3277; found: 797.3257.

Acetal aldehyde (M)-59: Benzyloxytrimethylsilane (3.5 mL, 18 mmol) was added to a solution of (M)-67 (2.13 g, 2.67 mmol) in toluene (21 mL) at RT. TMSOTf (5.7 mg, 0.026 mmol) was added to the reaction mixture at -78°C and stirred for 4 h at -15°C. The reaction was stopped by adding Et₃N (0.1 mL) and saturated aqueous NaHCO₃, and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/ EtOAc=95:5 to 80:20, gradient elution) to afford the acetal derivative of (M)-67 as a white amorphous solid (2.66 g, quant). $[\alpha]_{D}^{29} = -64.2$ (c=1.22 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): $\delta = 0.89-1.02$ (m, 21 H), 2.43 (s, 3H), 3.70 (s, 3H), 3.80 (s, 3H), 3.96 (s, 3H), 4.24 (d, J=11.6 Hz, 1H), 4.30-4.42 (m, 4H), 4.57 (d, J=10.4 Hz, 1H), 4.67-4.74 (m, 3H), 4.79 (d, J=10.4 Hz, 1 H), 5.52 (s, 1 H), 6.75 (d, J=7.2 Hz, 2 H), 6.82 (s, 1 H), 7.09 (m, 7H), 7.20–7.26 (m, 11H), 7.39 (s, 1H), 8.42 ppm (s, 1H); ¹³C NMR $(100 \text{ MHz}, \text{ CDCl}_3): \delta = 12.0, 18.0, 20.0, 52.0, 56.6, 61.5, 62.8, 68.6, 68.7,$ 75.5, 75.6, 99.4, 108.4, 116.3, 120.4, 123.2, 123.7, 124.3, 126.4, 126.7, 127.1, 127.2, 127.26, 127.29, 127.4, 127.5, 127.78, 127.80, 127.85, 127.87, 128.1 (2C), 131.2, 136.3, 137.2, 137.3, 137.7, 137.8, 137.9, 144.2, 146.0, 152.7, 152.79, 152.80, 168.8 ppm; IR (neat): $\tilde{v} = 3430$, 2940, 1730, 1650, 1600, 1570, 1500, 1450, 1410, 1340, 1280, 1220, 1190, 1120, 1060, 1000, 910, 880, 810, 740, 700 cm⁻¹; elemental analysis calcd (%) for $C_{60}H_{67}ClO_9Si$: C 72.37, H 6.78; found: C 72.20, H 6.83.

nBu₄NF (1.0м in THF, 1.28 mL, 1.28 mmol) was added to a solution of the acetal (1.07 g, 1.01 mmol) in THF (10 mL) at 0°C. After stirring for 1 h at 0°C, the reaction was stopped by adding water and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na2SO4), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/ EtOAc = 7:3) to afford the acetal alcohol derivative of (M)-67 as a white amorphous solid (0.898 g, quant). $[\alpha]_{D}^{30} = -111$ (c = 0.924 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): $\delta = 2.42$ (s, 3H), 2.45 (t, J = 6.6, OH), 3.77 (s, 3 H), 3.82 (s, 3 H), 3.97 (s, 3 H), 4.15 (d, J=11.2 Hz, 1 H), 4.15-4.26 (m, 2H), 4.34 (d, J=11.2 Hz, 1H), 4.11 (s, 2H), 4.50 (d, J=10.0 Hz, 1H), 4.63 (d, J=10.0 Hz, 1 H), 4.72 (d, J=10.0 Hz, 1 H), 4.76 (d, J=10.0 Hz, 1H), 5.59 (s, 1H), 6.77 (d, J = 7.6 Hz, 2H), 6.85 (s, 1H), 6.97–7.29 (m, 19H), 8.44 ppm (s, 1H); ¹³C NMR (100 MHz, CDCl₃): δ = 19.6, 52.2, 56.4, 61.5, 63.5, 68.3, 69.4, 75.7, 75.8, 99.4, 108.3, 116.5, 120.1, 123.6, 125.9, 126.8, 126.9, 127.48, 127.50, 127.57, 127.59 (2C), 127.63, 127.7, 127.8, 128.0, 128.1, 128.16, 128.20, 128.3, 131.2, 136.7, 136.8 (2 C), 137.2, 137.4, 137.6, 143.5, 145.9, 152.57, 152.59, 152.7, 168.5 ppm; IR (neat): $\tilde{\nu} = 3500$, 3090, 3060, 3030, 3010, 2950, 2930, 2870, 1730, 1600, 1590, 1570, 1500, 1450, 1440, 1400, 1370, 1340, 1280, 1220, 1200, 1140, 1100, 1050, 1030, 910, 810, 750, 740, 700 cm⁻¹; elemental analysis calcd (%) for C₅₁H₄₇ClO₉: C 72.98, H 5.64; found: C 72.92, H 5.85.

 MnO_2 (8.67 g, 99.6 mmol) was added to a solution of the acetal alcohol derivative of (*M*)-67 (4.18 g, 4.98 mmol) in CH₂Cl₂ (100 mL). After stirring for 15 h at RT, the reaction mixture was filtered through a Celite

pad and the filtrate was concentrated in vacuo to afford analytically pure (M)-59 (4.11 g, 99%) as a white amorphous solid. $[\alpha]_{D}^{30} = -59.1$ (c=1.07 in CHCl₃); ¹H NMR (400 MHz, CDCl₃): $\delta = 2.38$ (s, 3H), 3.71 (s, 3H), 3.85 (s, 3 H), 3.98 (s, 3 H) 4.27 (d, J=11.6 Hz, 1 H), 4.35 (d, 11.6 Hz, 1 H), 4.41 (s, 2 H), 4.54 (d, J=10.4 Hz, 1 H), 4.60 (d, J=10.4 Hz, 1 H), 4.68 (d, J = 10.4 Hz, 1 H), 4.76 (d, J = 10.4 Hz, 1 H), 5.49 (s, 1 H), 6.76 (d, J = 10.4 Hz, 1 H), 5.49 (s, 1 H), 6.76 (d, J = 10.4 Hz, 1 H), 5.49 (s, 1 H), 6.76 (d, J = 10.4 Hz, 1 H), 5.49 (s, 1 H), 6.76 (d, J = 10.4 Hz, 1 H), 5.49 (s, 1 H), 6.76 (d, J = 10.4 Hz, 1 H), 5.49 (s, 1 H), 6.76 (d, J = 10.4 Hz, 1 H), 5.49 (s, 1 H), 6.76 (d, J = 10.4 Hz, 1 H), 5.49 (s, 1 H), 7.6 Hz, 2H), 6.83 (s, 1H), 6.91-6.95 (m, 2H), 6.95-6.99 (m, 2H), 7.04 (t, J = 7.6 Hz, 2 H), 7.09–7.13 (m, 1 H), 7.16–7.32 (m, 11 H), 7.48 (s, 1 H), 8.45 (s, 1 H), 9.66 ppm (s, 1 H); 13 C NMR (100 MHz, CDCl₃): $\delta = 9.4$, 52.4, 56.3, 61.5, 68.1, 69.1, 75.6, 76.2, 99.0, 108.3, 116.5, 119.9, 124.0, 124.1, 124.5, 126.9, 127.3, 127.4, 127.5, 127.69, 127.72, 127.9, 127.96, 128.05, 128.1, 128.2, 130.6, 131.5, 133.6, 133.8, 136.0, 136.4, 136.6, 136.7, 137.3, 137.35, 137.42, 145.9, 152.6, 153.40, 153.45, 167.7, 191.0 ppm; IR (KBr): $\tilde{\nu} = 3030, 3000, 2950, 2930, 2870, 2840, 1730, 1700, 1590, 1570, 1500, 1450,$ 1340, 1280, 1220, 1100, 1040, 910, 850, 810, 740, 700 cm⁻¹; elemental analysis calcd (%) for C₅₁H₄₅ClO₉: C 73.15, H 5.42; found: C 73.44, H 5.62; HRMS (FAB): m/z: calcd for $C_{51}H_{46}O_9Cl$ ([*M*+H]⁺): m/z: 837.2831; found: 837.2803.

The enantiomeric purity of (*M*)-**59** was assessed by HPLC analysis (DAICEL CHIRALPAK AD-H (0.46 cm $\varphi \times 25$ cm), hexane/*i*PrOH= 9:1, flow rate = 1.0 mLmin⁻¹, $t_{\rm R} = 5.4$ min for the *M* isomer, 8.0 min for the *P* isomer).

Tetracycle (*S*,*S*)-60: BF₃-OEt₂ (21.5 μ L, 0.174 mmol) and a solution of MeOH (0.57 μ in THF, 100 μ L, 0.057 mmol) were added to a solution of (*M*)-59 (48.4 mg, 0.0578 mmol) in THF (3.0 mL) at -78° C. Samarium diiodide (0.1 μ in THF, 1.5 mL, 0.15 mmol) was added to the mixture at -78° C and the reaction temperature was immediately raised to 0°C. After stirring for 30 min at 0°C, the reaction was stopped by adding 5% aqueous K₂CO₃ and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (hexane/EtOAc=7:3 and benzene/EtOAc=8:2) to afford tetracycle (*S*,*S*)-60 as a white amorphous solid (40.2 mg, 95%).

Experimental procedure for the preparative-scale synthesis of (S,S)-60: $BF_3\text{-}OEt_2~(88\,\mu\text{L},~0.72\,\text{mmol})$ and MeOH (19 $\mu\text{L},~0.0567\,\text{mmol})$ was added to a solution of (M)-59 (200 mg, 0.239 mmol) in THF (12 mL) at -78°C. Samarium diiodide (0.1 m in THF, 1.5 mL, 0.15 mmol) was added to the reaction mixture at -78°C and the reaction temperature was immediately raised to -30 °C. The reaction temperature was then gradually raised from -30°C to -15°C over 30 min. The reaction was stopped by adding 5% aqueous K2CO3 and the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (benzene/EtOAc = 9:1) to afford tetracycle (S,S)-60 as a white amorphous solid (155 mg, 89%). Compound (S,S)-60 was formed as a mixture of two conformers: diaxial/diequatorial = 1:3.5). $[\alpha]_{D}^{29} = -30.8$ (c = 0.866 in CHCl₃); ¹H NMR (500 MHz, CDCl₃, the minor conformer is indicated by an asterisk): $\delta = 0.42^{*}$ (d, J = 6.5 Hz, OH), 2.36^{*} (s, 3H), 2.38 (s, 3H), 2.88 (d, J=1.8 Hz, 1 H; OH), 3.84* (s, 3 H) 3.90 (s, 3 H), 3.94 (s, 3 H), 3.95 (s, 3H), 3.98 (dd, J=1.8, 10.6 Hz, 1H), 4.00* (s, 3H), 4.01* (s, 3H), 4.05 (d, J=10.6 Hz, 1 H), 4.36 (d, J=11.4 Hz, 1 H), 4.41* (d, J=12.4 Hz, 1 H), 4.476* (d, J = 12.1 Hz, 1H), 4.480* (s, 2H), 4.52* (d, J = 3.3 Hz, 1H), 4.55^* (dd, J=3.3, 6.5 Hz, 1H), 4.59 (d, J=12.1 Hz, 1H), 4.68 (d, J=12.1 Hz, 1H), 4.6811.2 Hz, 1 H), 4.81 (d, J=11.2 Hz, 1 H), 4.89 (d, J=11.4 Hz, 1 H), 5.00 (d, $J = 11.2 \text{ Hz}, 1 \text{ H}), 5.11^* (d, J = 12.1 \text{ Hz}, 1 \text{ H}), 6.75-6.79^* (m, 2 \text{ H}), 6.81-$ 6.85 (m, 3H), 6.87* (s, 1H), 6.89* (s, 1H), 6.93* (t, J=7.6 Hz, 2H), 6.96-7.16 (m, 8H), 6.96–7.16* (m, 9H), 7.20–7.27* (m, 2H), 7.24 (s, 1H), 7.36– 7.41 (m, 1H), 7.43-7.47 (m, 2H), 7.54-7.58 (m, 2H), 7.72* (s, 1H), 7.92 ppm (s, J = 1.4 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃, the minor conformer is indicated by an asterisk): $\delta = 19.2^{*}$, 19.5, 52.1^{*}, 52.2, 56.4, 56.5^{*}, 61.2, 61.4*, 69.9*, 70.8*, 72.3, 74.9, 76.4, 76.7*, 77.1*, 77.2, 78.0*, 82.4, 107.3, 108.1*, 111.6, 119.5, 120.0, 120.2*, 120.5*, 120.9, 122.5*, 122.6, 122.8*, 123.5, 123.8*, 126.5*, 127.4, 127.48, 127.515, 127.517*, 127.598*, 127.60, 127.7, 127.9, 128.2, 128.36, 128.38, 128.6*, 128.81, 128.83, 129.2, 130.5*, 131.0*, 131.4, 132.5*, 136.09, 136.10*, 136.5, 136.8, 137.0*, 137.4*, 137.6, 137.9*, 138.8*, 139.8, 145.5, 145.6*, 152.2, 153.0, 153.1*, 153.2*, 153.4, 154.9, 168.6*, 169.0 ppm; IR (neat): $\tilde{\nu} = 3480$, 2950, 2870, 1730, 1590, 1570, 1450, 1340, 1280, 1220, 1130, 1100, 1050, 810, 750, 700 cm^{-1} ;

elemental analysis calcd (%) for $C_{44}H_{39}ClO_8$: C 72.27, H 5.38; found: C 72.27, H 5.50; HRMS (FAB): m/z: calcd for $C_{44}H_{40}O_8Cl$ ([*M*+H]⁺): 731.2411; found: 731.2377.

The enantiomeric purity of (*S*,*S*)-**60** was analyzed by HPLC analysis (DAICEL CHIRALPAK AD-H (0.46 cm $\varphi \times 25$ cm), hexane/EtOH=8:2, flow rate=1.0 mLmin⁻¹, $t_{\rm R}$ =6.6 min for the *S*,*S* isomer, 9.3 min for the *R*,*R* isomer; Figure 1).

Amide 68: A 5M aq. KOH solution (10 mL) was added to a solution of (S,S)-60 (199 mg, 0.272 mmol) in EtOH (18 mL). The mixture was sealed in a thick-walled glass tube and heated at 100 °C. After stirring for 3 h at 100 °C, the mixture was diluted with Et₂O and the phases were separated. The organic layer was extracted with water (×2). The combined water layers were acidified with 2M aq. HCl and extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was dissolved in DMF (6 mL), and BOP (172 mg, 0.388 mmol), D-Ala-OMe+HCl (185 mg, 1.33 mmol), and Et₃N (250 µL, 1.79 mmol) were added to the resulting solution. After stirring for 1 h at RT, the reaction was stopped by adding 2M aq. HCl and then the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, hexane/EtOAc=1:1) to afford amide 68 as a white solid (184 mg, 84%). Mixture of two conformers: diaxial/diequatorial=1:4.5; m.p. 163.6–164.3 °C (hexane and Et_2O); $[\alpha]_{\text{D}}^{29} = -25.3$ (c=0.872 in CHCl₃); ¹H NMR (500 MHz, CDCl₃, the minor conformer is indicated by an asterisk): $\delta = 0.99^{*}$ (d, J = 6.2 Hz, 1H; OH), 1.23^{*} (d, J = 7.2 Hz, 3H), 1.39 (d, J = 7.2 Hz, 3H), 2.42* (s, 3H), 2.44 (s, 3H), 2.94 (d, J = 2.3 Hz, 1H; OH), 3.67 (s, 3H), 3.73* (s, 3H), 3.94 (s, 3H), 3.95 (s, 3H), 3.96* (s, 3H), 3.97 (dd, J=1.4, 11.4 Hz, 1H; 6-H), 4.00^* (s, 3H), 4.24 (d, J= 11.4 Hz, 1H), 4.25 (d, J=11.4 Hz, 1H; C-5), 4.37* (d, J=11.6 Hz, 1H), 4.43* (d, J=12.3 Hz, 1 H), 4.45* (d, J=11.0 Hz, 1 H), 4.51* (d, J=12.3 Hz, 1H), 4.54* (d, J=11.0 Hz, 1H), 4.578 (d, J=11.5 Hz, 1H), 4.585* (d, J = 4.3 Hz, 1H; 6-H), 4.66* (dq, J = 7.3, 7.3 Hz, 1H), 4.67 (dq, J=7.1, 7.1 Hz, 1 H), 4.70* (d, J=4.3 Hz, 1 H; 5-H),4.76 (d, J=11.5 Hz, 1 H), 4.82 (d, J=11.2 Hz, 1 H), 4.91 (d, J=11.4 Hz, 1 H), 5.01 (d, J=11.4 H 11.2 Hz, 1H), 6.06* (d, J=7.2 Hz, 1H; NH), 6.35 (d, J=7.2 Hz, NH), 6.82 (s, 1H), 6.87* (s, 1H), 6.92-6.99 (m, 5H), 7.02-7.19 (m, 6H), 7.23-7.30 (m, 1H), 7.37–7.41* (m, 2H), 7.44–7.48 (m, 2H), 7,57–7.59 (m, 2H), 7.93 ppm (d, J=1.4 Hz, 1 H); ¹³C NMR (125 MHz, CDCl₃, the minor conformer is indicated by an asterisk): $\delta = 17.8^{*}$, 18.1, 19.2*, 19.8, 48.2*, 48.4, 52.25, 52.31*, 56.5, 61.2, 61.4*, 69.9*, 71.1*, 72.4, 74.9, 76.1*, 76.6, 76.8* (overlapped by a signal of CDCl₃), 77.2, 78.0*, 82.4, 107.3, 108.1*, 111.7, 119.5, 120.2*, 120.6*, 120.7, 121.0, 122.1*, 122.6, 122.9*, 123.5, 123.8*, 126.9*, 127.39*, 127.43, 127.47, 127.50*, 127.6, 127.7*, 127.8*, 127.88, 127.89*, 128.1*, 128.18, 128.22, 128.37, 128.40*, 128.7, 128.8, 136.3, 136.8, 137.3, 137.5*, 137.6, 137.7*, 137.9*, 138.1, 138.9*, 139.6, 145.47, 145.54*, $152.5, \ 152.9, \ 153.2^*, \ 153.3, \ 153.5^*, \ 154.7^*, \ 167.2^*, \ 167.4, \ 173.1,$ 173.2* ppm; IR (neat): $\tilde{\nu} = 3420, 3060, 3030, 3000, 2940, 2870, 2840, 2360,$ 1740, 1660, 1650, 1590, 1570, 1500, 1450, 1340, 1210, 1150, 1130, 1110, 1050, 750, 700 cm⁻¹; elemental analysis calcd (%) for $C_{47}H_{44}CINO_9$: C 70.36, H 5.53, N 1.75; found: C 70.11, H 5.44, N 1.70; HRMS (FAB): m/z: calcd for C₄₇H₄₅O₉NCl ([M+H]⁺): 802.2782; found: 802.2809.

Glycoside **69**: MS4A (320 mg) was placed in a two-necked round-bottomed flask and flame dried. The promoter was prepared in situ by stirring a mixture of Cp₂HfCl₂ (62.8 mg, 0.165 mmol) and AgOTf (84.2 mg, 0.328 mmol) in CH₂Cl₂ (0.8 mL) for 10 min at RT. A solution of **68** (122 mg, 0.152 mmol) and **51** (100 mg, 0.181 mmol) in CH₂Cl₂ (2.8 mL) were added to this suspension at -78 °C. After stirring for 11 h at -36 °C, the reaction was stopped by adding saturated aqueous NaHCO₃. The mixture was filtered through a Celite pad and extracted with EtOAc (× 3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (hexane/Et₂O/CH₃CN=3:4.5:1.5) to afford β-glycoside **69** as a white amorphous solid (145 mg, 72%) and α-glycoside **69** also as a white amorphous solid (18.4 mg, 9%).

β-69 (a mixture of two conformers: diaxial/diequatorial=50:50): M.p. 114–116 °C; $[a]_D^{22} = +20.0$ (c=0.880 in CHCl₃); ¹H NMR (500 MHz, CDCl₃, without distinction of the two conformers): $\delta=1.07$ (d, J=7.2 Hz,

3H), 1.35 (d, J=7.3 Hz, 3H), 1.362 (d, J=6.6 Hz, 3H), 1.37 (s, 3H), 1.38 (d, J=6.6 Hz, 3 H), 1.57 (s, 3 H), 1.89 (s, 3 H), 1.96 (s, 3 H), 1.99 (s, 3 H), 2.02 (s, 3H), 2.34 (s, 3H), 2.45 (s, 3H), 3.24 (dd, J=9.1, 11.7 Hz, 1H), 3.35 (dd, J=9.2, 11.6, 1H), 3.62 (dq, J=1.3, 6.6 Hz, 1H), 3.65 (s, 3H), 3.70-3.74 (m, 1H), 3.72 (s, 3H), 3.79 (s, 3H), 3.80 (dd, J=1.3, 3.7 Hz, 1H), 3.84 (s, 3H), 3.85 (s, 3H), 3.88 (dd, J=1.1, 3.8 Hz, 1H), 3.89 (s, 3 H), 3.92 (dd, J=3.7, 9.9 Hz, 1 H), 3.99 (d, J=11.3 Hz, 1 H), 4.04 (dd, J= 1.4, 10.8 Hz, 1 H), 4.071 (dd, J = 5.0, 11.7 Hz, 1 H), 4.072 (dd, J = 3.8, 10.1 Hz, 1 H), 4.16 (dd, J=5.1, 11.6 Hz, 1 H), 4.25 (d, J=11.3 Hz, 1 H), 4.38 (s, 2 H), 4.39 (d, J=12.1 Hz, 1 H), 4.46 (d, J=12.3 Hz, 1 H), 4.451 (d, J = 7.1 Hz, 1H), 4.46–4.50 (m, 3H), 4.53 (d, J = 8.0 Hz, 1H), 4.55 (d, J = 6.0 Hz, 1H), 4.55 (d, {J = 6.0 10.8 Hz, 1 H), 4.58 (dq, J=7.2, 7.3 Hz, 1 H), 4.63 (d, J=11.6 Hz, 1 H), 4.65 (d, J = 3.2 Hz, 1H), 4,655 (qd, J = 7.2, 7.9 Hz, 1H), 4.660 (d, J =7.3 Hz, 1 H), 4.71 (d, J=12.3 Hz, 1 H), 4.73 (dd, J=7.3, 8.9 Hz, 1 H), 4.85 (ddd, J = 5.1, 8.9, 9.1 Hz, 1 H), 4.87-4.92 (m, 3 H), 4.93 (ddd, J = 5.0, 8.9, 1 Hz, 1 H)9.2 Hz, 1 H), 5.02 (d, J=3.2 Hz, 1 H), 5.03 (dd, J=8.9, 8.9 Hz, 1 H), 5.15 (dd, J=8.0, 10.1 Hz, 1H), 5.16 (d, J=7.9 Hz, 1H), 5.54 (d, J=7.9 Hz, 1H; NH), 5.69 (dd, J=7.9, 9.9 Hz, 1H), 6.23 (d, J=7.2 Hz, 1H; NH), 6.65 (s, 1H), 6.76 (s, 1H), 6.80-6.89 (m, 3H), 6.92-6.99 (m, 4H), 7.03-7.16 (m, 5H), 7.17-7.37 (m, 23H), 7.39-7.44 (m, 4H), 7.46 (s, 1H), 7.48-7.54 (m, 1H), 7.81 (d, J=1.4 Hz, 1H), 7.95–7.99 ppm (m, 1H); ¹³C NMR (125 MHz, CDCl₃, without distinction of the two conformers): $\delta = 17.29$, 17.30, 17.4, 18.1, 19.0, 19.6, 19.8, 19.9, 20.5, 20.60, 20.65, 20.69, 47.9, 48.4, 52.2, 56.47, 56.52, 61.1 (2C), 62.3, 62.4, 65.5, 65.8, 68.9, 69.0, 69.2, 69.4, 69.9, 70.6, 70.8, 70.9, 71.3, 71.5, 71.6, 74.0, 74.3, 74.4, 75.9, 76.50, 76.53, 77.1, 77.2 (overlapped by a signal of CDCl₃), 77.4, 79.8, 80.4, 81.5, 97.8, 100.8, 102.1, 102.4, 107.3, 107.9, 112.3, 118.9, 119.6, 120.9, 122.5, 122.8, 123.1, 123.23, 123.25, 127.1, 127.2, 127.3, 127.47, 127.52, 127.54, 127.59, 127.63, 127.67, 127.73, 127.8, 127.9, 128.0, 128.1, 128.3, 128.4, 128.5, 128.6, 128.7, 129.7 (2C), 130.3, 131.2, 132.7, 133.0, 133.3, 136.5, 136.6, 136.9, 137.2, 137.3, 137.5, 137.89, 138.0, 138.2, 138.8, 145.1, 145.5, 152.3, 152.7, 153.3, 153.4, 154.3, 154.4, 163.8, 164.7, 167.4, 168.9, 169.1, 169.70, 169.73, 170.05, 170.14, 172.9, 173.1 ppm; IR (KBr): v=2100, 1760, 1750, 1660, 1590, 1510, 1450, 1370, 1340, 1270, 1220, 1060, 750, 710, 700 cm⁻¹; elemental analysis calcd (%) for C71H71ClN4O20: C 63.84, H 5.36, N 4.19; found: C 63.56, H 5.32, N 3.91.

 α -69 (mixture of two conformers: diaxial/diequatorial=75:25): M.p. 95-97°C; $[\alpha]_D^{28} = +111$ (c=0.960 in CHCl₃); ¹H NMR (500 MHz, CDCl₃, the minor conformer is indicated by an asterisk): $\delta = 0.84^*$ (d, J = 6.4 Hz, 3 H), 1.05 (d, J=7.2 Hz, 3 H), 1.23 (s, 3 H), 1.36* (d, J=7.2 Hz, 3 H), 1.38 (d, J=6.4 Hz, 3H), 1.48* (s, 3H), 1.62* (s, 3H), 1.89 (s, 3H), 1.98* (s, 3H), 2.01 (s, 3H), 2.07* (s, 3H), 2.29 (s, 3H), 3.24 (dd, J=9.2, 11.8 Hz, 1 H), 3.52^* (dd, J=9.0, 11.8 Hz, 1 H), 3.57-3.59 (m, 1 H), 3.60 (d, J=11.3 Hz, 1 H), 3.63* (s, 3 H), 3.67 (s, 3 H), 3.69 (s, 3 H), 3.85 (d, J =11.3 Hz, 1H), 3.87 (dd, J=1.3, 3.6 Hz, 1H), 3.91-3.96 (m, 1H), 3.96* (s, 3 H), 3.99* (s, 3 H), 4.05 (dd, J=5.2, 11.8 Hz, 1 H), 4.06 (s, 3 H), 4.09 (dd, J=3.6, 10.2 Hz, 1 H), 4.21^* (dd, J=1.3, 11.0 Hz, 1 H), 4.21^* (d, J=1.3, 111.0 Hz, 1 H), 4.257 (dd, J = 5.0, 11.8 Hz, 1 H), 4.258* (d, J = 11.5 Hz, 1 H), 4.35 (d, J=11.1 Hz, 1 H), 4.43* (d, J=11.4 Hz, 1 H), 4.45 12.4 Hz, 1 H), 4.45–4.49* (m, 1 H), 4.46 (d, J = 11.1 Hz, 1 H), 4.48 (d, J = 11.1 Hz, 1 H Hz, 1 H), 4.48 (d, J = 11 7.3 Hz, 1H), 4.52 (d, J=12.4 Hz, 1H), 4.55–4.60* (m, 1H), 4.58 (qd, J= 7.2, 7.9 Hz, 1 H), 4.59* (dq, J=7.0, 7.9 Hz, 1 H), 4.71* (d, J=11.4 Hz, 1 H), 4.73 (dd, J=7.3, 9.1 Hz, 1 H), 4.75 (d, J=3.3 Hz, 1 H), 4.76 (d, J= 3.3 Hz, 1 H), 4.84* (d, J=10.6 Hz, 1 H), 4.85 (ddd, J=5.2, 9.2, 9.2 Hz, 1 H), 4.88* (d, J = 11.5 Hz, 1 H), 4.89–4.94* (m, 2 H), 4.90–4.95 (m, 1 H), 4.99* (ddd, J=5.0, 8.6, 9.0 Hz, 1 H), 5.03* (d, J=10.6 Hz, 1 H), 5.07 (d, J=4.0 Hz, 1 H), 5.14* (dd, J=8.6, 9.0 Hz, 1 H), 5.18* (d, J=3.6 Hz, 1 H), 5.19 (dd, J=4.0, 10.1 Hz, 1 H), 5.53 (d, J=7.9 Hz, 1 H; NH), 5.56* (dd, J=3.6, 10.5 Hz, 1H), 6.34* (d, J=7.0 Hz, 1H; NH), 6.64* (s, 1H), 6.68 (s, 1H), 6.82-6.86 (m, 1H), 6.89-7.05 (m, 4H), 7.07-7.13 (m, 4H), 7.15-7.36 (m, 10H), 7.39–7.66 (m, 2H), 7.81 (s, 1H), 8.01* (d, J=1.3 Hz, 1H), 8.19-8.23 ppm* (m, 2H); ¹³C NMR (125 MHz, CDCl₃, without distinction of the two conformers): $\delta = 16.5$, 17.3, 17.4, 18.1, 18.7, 19.0, 19.5, 19.8, 20.55, 20.64, 20.68, 20.75, 47.9, 48.5, 52.2, 52.3, 56.2, 56.5, 61.2, 61.5, 62.3, 62.4, 64.75, 64.85, 66.1, 66.6, 68.95, 69.02, 69.1, 70.0, 70.1, 71.0, 71.1, 71.5, 71.6, 74.6, 75.0, 75.8, 75.9, 76.2, 76.5, 77.21, 77.16 (overlapped by a signal of CDCl₃), 77.5, 77.68, 80.0, 93.4, 96.1, 100.9, 101.0, 102.1, 102.3, 107.3, 107.9, 112.1, 119.3, 119.5, 119.8, 120.8, 122.0, 122.3, 122.7, 122.9, 123.6, 126.8, 127.1, 127.2, 127.4, 127.6, 127.68, 127.73, 127.76, 127.80, 127.82,

128.0, 128.1, 128.3, 128.5, 128.6, 128.7, 128.9, 128.99, 129.01, 129.2, 129.3, 130.1, 130.3, 130.5, 131.4, 132.9, 133.0, 133.9, 133.98, 134.0, 136.8, 137.0, 137.1, 137.77, 137.82, 137.84, 137.9, 138.7, 145.3, 145.5, 152.3, 153.0, 153.4, 153.8, 154.6, 164.7, 165.7, 167.2, 167.3, 168.9, 169.0, 169.75, 169.83, 170.0, 170.1, 172.8, 173.0 ppm; IR (KBr): $\tilde{\nu}$ = 3060, 3030, 2940, 2110, 1760, 1660, 1590, 1500, 1450, 1370, 1340, 1270, 1250, 1220, 1160, 1110, 1070, 1050, 950, 900, 880, 820, 750, 740, 710, 700 cm⁻¹; elemental analysis calcd (%) for C₇₁H₇₁ClN₄O₂₀: C 63.84, H 5.36, N 4.19; found: C 63.59, H 5.49, N 3.94.

Quinone 70: A solution of CAN (158 mg, 0.288 mmol) in water (0.32 mL) was added to a solution of glycoside 69 (165 mg, 0.123 mmol) in CH₃CN (0.9 mL) at RT. After stirring for 20 min, water (15 mL) was added to the reaction and a precipitate was formed which was collected by filtration as a red powder (154 mg). The powder was directly added to a solution of diene 21 (250 mg, 0.123 mmol) in THF (2.5 mL) at RT. Mixture allowed to stand for 20 min, acidic silica gel (pH 6, Kanto chemical) was added, and the organic solvent was removed in vacuo. After standing for 8 h, the silica gel was placed on a glass filter and washed with EtOAc. Then the filtrate was concentrated in vacuo. K₂CO₃ (176 mg) was added to the solution of this crude material (227 mg) in THF/CH₂Cl₂ (1:3, 8 mL) at RT. The mixture was heated at 40 °C for 3 h, and the reaction was stopped by adding 2M aq. HCl and then the mixture was extracted with CH_2Cl_2 (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (hexane/Et₂O/CH₃CN=2:3:1) and HPLC (Cica-reagent, Myghtysil Si60, 250–20 (5 μ m), hexane/EtOAc=1:2, 3.5 mLmin⁻¹) to afford quinone 70 as a yellow solid (124 mg, 74%). Mixture of two conformers: diaxial/diequatorial=78:22; m.p. 126.0-126.8 °C (hexane and EtOAc); $[\alpha]_{D}^{20} = +111$ (c=1.17 in CHCl₃); ¹H NMR (500 MHz, CDCl₃, the minor conformer is indicated by an asterisk): $\delta = 1.08$ (d, J = 7.2 Hz, 3H), 1.26 (s, 3H), 1.35* (d, J=6.3 Hz, 3H), 1.37* (d, J=7.2 Hz, 3H), 1.45 (d, J=6.3 Hz, 3H), 1.87 (s, 3H), 1.96* (s, 3H), 1.98 (s, 3H), 2.02* (s, 3H), 2.04* (s, 3H), 2.36 (s, 3H), 2.48* (s, 3H), 3.26 (dd, J=8.9, 11.8 Hz, 1H), 3.36* (dd, J=9.3, 11.6 Hz, 1H), 3.63* (s, 3H), 3.69-3.73 (m, 1H), 3.70-3.74* (m, 1H), 3.75 (s, 3H), 3.85 (dd, J=1.4, 3.9 Hz, 1H), 3.89* $(brd, J=4.1 Hz, 1H), 3.91^* (dd, J=1.1, 11.0 Hz, 1H), 3.946^* (s, 3H),$ 3.947 (dd, J=3.9, 9.9 Hz, 1 H), 4.00 (s, 3 H), 4.06* (dd, J=4.1, 9.9 Hz, 1H), 4.08 (dd, J=5.2, 11.8 Hz, 1H), 4.12 (d, J=11.0 Hz, 1H), 4.17* (dd, J=5.2, 11.6 Hz, 1 H), 4.30* (d, J=11.2 Hz, 1 H), 4.32 (d, J=11.5 Hz, 1 H), 4.41 (d, J=12.1 Hz, 1 H), 4.42* (d, J=11.6 Hz, 1 H), 4.441 (d, J=12.1 Hz, 1H), 4.442 (d, J=11.5 Hz, 1H), 4.47* (d, J=11.4 Hz, 1H), 4.48 (d, J=11.4 Hz, 1 7.3 Hz, 1 H), 4.51 (d, J=7.8 Hz, 1 H), 4.52* (d, J=11.0 Hz, 1 H), 4.55 (qd, J=7.2, 7.9 Hz, 1 H), 4.57 (d, J=3.0 Hz, 1 H), 4.64* (qd, J=7.2, 7.2 Hz, 1H), 4.65* (d, J=7.3 Hz, 1H), 4.69* (d, J=11.4 Hz, 1H), 4.71 (dd, J= 7.3, 8.7 Hz, 1 H), 4.77* (d, J=11.6 Hz, 1 H), 4.85 (ddd, J=5.2, 8.9, 9.8 Hz, 1H), 4.90 (dd, J=8.7, 8.9 Hz, 1H), 4.91* (dd, J=7.3, 9.0 Hz, 1H), 4.91-4.95* (m, 1H), 4.96 (d, J=3.0 Hz, 1H), 5.03* (dd, 1H; J=9.0, 9.0 Hz, 1H), 5.12* (d, J=8.0 Hz, 1H), 5.14 (d, J=11.0 Hz, 1H), 5.15 (dd, J=7.8, 9.9 Hz, 1H), 5.18* (d, J=11.2 Hz, 1H), 5.37 (d, J=7.9 Hz, 1H; NH), 5.69* (dd, J=8.0, 9.9 Hz, 1H), 6.24* (d, J=7.2 Hz, 1H; NH), 6.69-6.80 (m, 3H), 6.95-7.14 (m, 11H), 7.19-7.40 (m, 7H), 7.51-7.59 (m, 2H), 7.60 (s, 1H), 7.95* (s, 1H), 7.97* (s, 1H), 8.08* (d, J=1.1 Hz, 1H), 12.7 (s, 1H; OH), 12.9 ppm* (s, 1H; OH); ¹³C NMR (125 MHz, CDCl₃, without distinction of the two conformers): $\delta = 17.25, 17.3, 17.35, 18.0, 19.0, 19.5,$ 19.9, 20.1, 20.5, 20.60, 20.64, 20.7, 47.8, 48.4, 52.2, 52.3, 56.0, 56.1, 62.4, 62.5, 65.4, 65.7, 68.9, 69.0, 69.4, 69.5, 70.6, 70.8, 70.9, 71.1, 71.2, 71.5, 71.6, 73.2, 74.1, 74.6, 76.7, 76.8 (overlapped by a signal of CDCl₃), 77.1, 77.3 (2 C, overlapped by a signal of CDCl₃), 79.8, 80.3, 80.5, 98.1, 100.6, 102.2, 102.4, 105.8, 106.0, 107.0, 107.3, 110.4, 110.8, 118.2, 120.3, 121.5, 123.1, 123.2, 125.1, 126.1, 127.3, 127.55, 127.58, 127.64, 127.66, 127.8, 127.9, 128.1, 128.26, 128.31, 128.4, 128.49, 128.50, 128.6, 128.67, 128.68, 128.8, 129.0, 129.6, 129.7, 130.2, 132.4, 132.9, 133.1, 133.3, 133.4, 134.4, 135.3, 136.1, 136.3, 136.5, 136.6, 136.7, 137.27, 137.31, 137.4, 137.8, 138.2, 139.1, 139.9, 140.1, 144.1, 152.9, 154.8, 156.5, 158.5, 163.5, 164.6, 164.7, 164.9, 166.51, 166.54, 166.99, 167.02, 168.8, 169.1, 169.70, 169.75, 170.0, 170.1, 172.8, 172.9, 180.5, 181.0, 185.5, 186.0 ppm; IR (KBr): $\tilde{\nu}$ =2960, 2920, 2850, 2110, 1740, 1670, 1630, 1600, 1580, 1450, 1380, 1310, 1260, 1220, 1100, 1070, 1030, 800, 710 cm⁻¹; elemental analysis calcd (%) for C₇₄H₇₀N₄O₂₂: C 65.00, H 5.16, N 4.10; found: C 65.00, H 5.29, N 4.08.

Benanomicin B hydrochloride (2b·HCl): А 2м aq. NaOH solution (0.25 mL) was added to a solution of 70 (59.3 mg, 0.0434 mmol) in MeOH (5.0 mL). After stirring for 8.5 h, the reaction was stopped by adding 2M aq. HCl (0.55 mL) and the mixture was concentrated in vacuo (azeotropic evaporation ×5). The residue was purified by reversed-phase silica gel column chromatography (Cosmosil: MeOH/H2O=9:1) and Sephadex (LH-20: MeOH) to afford the crude product of 71. A 1 M aq. HCl solution (100 µL) and 5% Pd/C (82.3 mg) were added to a solution of this crude material (46.5 mg) in MeOH (4.5 mL) and DMF (1.5 mL). After stirring under H₂ (1 atm) at RT for 2 h, the reaction mixture was filtered through a Hyflo Super-Cel pad (washed with DMF) and concentrated in vacuo. The residue was purified by reversed-phase silica gel column chromatography (Cosmosil: CH₃CN/H₂O=3:7, pH 3) to give red precipitates, which were further purified by Sephadex LH-20 (DMF) (twice) to afford benanomicin B dimethylformamide solvate which was dissolved in MeOH and then 2M aq. HCl was added to adjust the pH value to 3.5. After removal of the solvent in vacuo, the residue was dissolved in DMSO (0.1 mL), and CHCl₃ (6.0 mL) was added. The precipitates were collected by filtration to afford benanomicin B hydrochloride (2b·HCl) as a red amorphous powder (19.7 mg, 53%). M.p. 213-215°C (decomp) (lit.:^[1b] >220 °C (decomp)); $[a]_{D}^{22} = +353$ (c=0.0501, H₂O) $(lit.:^{[1b]} + 360 (c = 0.05, H_2O)); {}^{1}H NMR (500 MHz, [D_6]DMSO, 313 K):$ $\delta = 1.18$ (d, J = 6.5 Hz, 3H), 1.34 (d, J = 7.3 Hz, 3H), 2.33 (s, 3H), 3.07 (dd, J=10.7, 10.9 Hz, 1 H), 3.16-3.18 (m, 3 H), 3.29-3.34 (m, 1 H), 3.39-3.44 (m, 1 H), 3.60 (br, 1 H), 3.72 (dd, J=5.3, 11.3 Hz, 1 H), 3.83 (br q, J= 6.5, 1 H), 3.91–3.94 (m, 1 H), 3.95 (s, 3 H), 4.42 (dq, J=7.1, 7.3 Hz, 1 H), 4.53-4.56 (m, 2H), 4.58-4.65 (m, 1H), 4.74 (d, J=7.6 Hz, 1H), 6.94 (d, J=2.4 Hz, 1 H), 7.23 (brs, 1 H), 7.31 (d, J=2.4 Hz, 1 H), 8.06 (s, 1 H), 8.43 (d, J=7.1 Hz, 1 H), 12.8 ppm (s, 1 H); ¹³C NMR (125 MHz, [D₆]DMSO, 313 K): $\delta = 16.3$, 16.9, 19.1, 47.6, 54.2, 56.4, 65.7, 66.9, 69.4, 69.7, 73.3, 75.9, 77.4, 80.9, 104.0, 104.5, 106.9, 107.6, 110.1, 113.7, 115.6, 125.7, 127.5, 131.3, 134.3, 137.3, 137.8, 147.9, 151.0, 156.9, 164.7, 166.0, 166.8, 173.9, 185.0, 187.5 ppm; IR (KBr): $\tilde{\nu}$ = 3360, 1730, 1610, 1300, 1160, 1080, 1040 cm⁻¹; LRMS (MALDI-TOF, DHBA matrix): m/z: calcd for C₃₉H₄₂O₁₈N₂Na ([*M*+Na]⁺): 849.2; found: 849.2; HRMS (FAB): m/z: calcd for C₃₉H₄₃O₁₈N₂ ([M+H]⁺): 827.2511; found: 827.2506.

N-Monomethylamine 73: A solution of Me₃P in toluene (0.91 M, 50 µL, 0.048 mmol) was added to a solution of 69 (53.5 mg, 0.0401 mmol) in CH_2Cl_2 (0.67 mL) at 0°C. After stirring for 1.5 h at RT, (HCHO)_n (6.50 mg, 0.201 mmol) was added. The reaction mixture was stirred for 11 h at RT before treatment with MeOH (0.50 mL) and NaBH₄ (10.6 mg, 0.281 mmol) at 0°C. After stirring for 10 min, the reaction was stopped by adding AcOH and the mixture was extracted with EtOAc (\times 3). The combined organic extracts were washed with saturated aqueous NaHCO3 and brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (hexane/Et₂O/CH₃CN/MeOH=22:55:17:6) to afford N-monomethylamine 73 (39.9 mg, 75%) as a white amorphous solid. Mixture of two conformers: diaxial/diequatorial=57:43); m.p. 95-97 °C; $[\alpha]_{D}^{30} = +42.6 \ (c = 1.14 \text{ in CHCl}_{3}); {}^{1}\text{H NMR} \ (500 \text{ MHz}, \text{CDCl}_{3}, \text{ the minor}$ conformer is indicated by an asterisk): $\delta = 1.06$ (d, J = 7.3 Hz, 3 H), 1.338* (d, J=6.0 Hz, 3 H), 1.342 (s, 3 H), 1.36* (d, J=7.3 Hz, 1 H), 1.37 (d, J= 6.0 Hz, 3 H), 1.57* (s, 3 H), 1.88 (s, 3 H), 1.96* (s, 3 H), 1.99 (s, 3 H), 2.04* (s, 3H), 2.35 (s, 3H), 2.44* (s, 3H), 2.51 (s, 3H), 2.65* (s, 3H), 2.75 (brd, J = 4.3 Hz, 1 H), 2.86* (brd, J = 4.3 Hz, 1 H), 3.24 (dd, J = 8.7, 11.8 Hz, 1H), 3.36* (dd, J=8.9, 11.6 Hz, 1H), 3.54–3.60 (m, 1H), 3.65* (s, 3H), 3.67-3.72* (m, 1H), 3.72 (s, 3H), 3.76 (s, 3H), 3.79* (s, 3H), 3.83 (s, 3H), 3.85 (s, 3H), 3.91* (s, 3H), 3.94 (d, J=11.3 Hz, 1H), 3.95* (dd, J=4.3, 10.0 Hz, 1 H), 4.00* (dd, J=1.3, 10.6 Hz, 1 H), 4.06* (dd, J=5.0, 11.6 Hz, 1 H), 4.15* (dd, J = 5.1, 11.8 Hz, 1 H), 4.25 (d, J = 11.4 Hz, 1 H), 4.38 (s, 2H), 4.39* (d, J=7.0 Hz, 1H), 4.41* (s, 2H), 4.43 (d, J=11.6 Hz, 1H), 4.46* (d, J=11.3 Hz, 1 H), 4.47* (d, J=12.0 Hz, 1 H), 4.49 (d, J=7.8 Hz, 1 H), 4.56* (d, J=10.6 Hz, 1 H), 4.57 (dq, J=6.8, 7.3 Hz, 1 H), 4.62 (d, J= 7.1 Hz, 1 H), 4.63* (d, J=11.3 Hz, 1 H), 4.66* (dq, J=7.2, 7.3 Hz, 1 H), 4.67 (d, J=3.1 Hz, 1 H), 4.71 (dd, J=7.1, 8.7 Hz, 1 H), 4.72* (d, J=12.0 Hz, 1 H), 4.84 (ddd, J=5.1, 8.7, 8.7 Hz, 1 H), 4.87 (dd, J=8.7, 8.7 Hz, 1 H), 4.88* (dd, J=7.0, 8.9 Hz, 1 H), 4.90 (d, J=11.4 Hz, 1 H), 4.93* (ddd, J=5.0, 8.9, 8.9 Hz, 1 H), 5.00* (dd, J=8.9, 8.9 Hz, 1 H), 5.04 (dd, J=7.8, 10.3 Hz, 1 H), 5.07 (d, J=3.1 Hz, 1 H), 5.16* (d, J=7.6 Hz, 1 H), 5.54 (d, J=6.8 Hz, 1H; NH), 5.59* (dd, J=7.6, 10.0 Hz, 1H), 6.24* (d, J=7.2 Hz,

Chem. Eur. J. 2007, 13, 9791-9823

1H; NH), 6.65 (s, 1H), 6.77* (s, 1H), 6.78-6.83 (m, 1H), 6.83-6.87* (m, 1H), 6.90-6.93 (m, 2H), 6.94-6.97* (m, 2H), 7.05-7.37 (m, 16H), 7.34-7.42 (m, 2H), 7.47 (s, 1H), 7.42–7.52* (m, 1H), 7.80* (d, J=1.3 Hz, 1H), 7.93-7.97 ppm* (m, 2H); 13C NMR (125 MHz, CDCl₃, without distinction of the two conformers): $\delta = 17.2$, 17.4 (2 C), 18.1, 19.0, 19.7, 19.8, 19.9, 20.55, 20.61, 20.69, 20.73, 38.1, 38.2, 47.9, 48.4, 52.19, 52.22, 56.4, 56.5, 61.08, 61.10, 62.1, 62.2, 63.9, 64.0, 69.0, 69.1, 69.9, 70.6, 70.8, 71.2, 71.5, 71.6, 71.8, 72.1, 73.7, 74.2, 74.3, 75.9, 76.5, 76.6, 77.3 (overlapped by CDCl₃), 80.6, 81.2, 81.6, 97.9, 101.0, 101.8, 102.1, 107.2, 107.8, 112.3, 118.9, 119.5, 120.8, 121.0, 122.5, 122.7, 122.8, 123.1, 123.20, 123.22, 127.05, 127.06, 127.2, 127.3, 127.50, 127.51, 127.56, 127.63, 127.66, 127.3, 127.8, 127.89, 127.94, 128.0, 128.1, 128.3, 128.4, 128.49, 128.52, 128.7, 128.9, 129.6, 129.8, 130.16, 130.23, 131.2, 132.2, 132.6, 133.1, 133.2, 136.6, 136.8, 137.17, 137.22, 137.7, 137.89, 137.93, 138.2, 138.8, 139.0, 145.1, 145.4, 152.2, 152.6, 153.2, 153.3, 154.2, 154.3, 163.9, 165.0, 167.5, 169.2, 169.4, 169.8, 169.9, 169.96, 170.04, 172.9, 173.1 ppm; IR (KBr): $\tilde{\nu}\!=\!3640,\;3410,$ 2940, 2870, 1750, 1660, 1590, 1590, 1500, 1450, 1370, 1340, 1270, 1220, 1110, 1050, 910, 820, 750, 700 cm^{-1} ; elemental analysis calcd (%) for C₇₂H₇₅ClN₂O₂₀: C 65.32, H 5.71, N 2.12; found: C 65.05, H 5.97, N 1.96. Trifluoroacetoamide β-76: Pyridine (0.1 mL, 1.25 mmol) and trifluoroacetic anhydride (0.01 mL, 0.625 mmol) were added to a solution of 73 (82.7 mg, 0.0625 mmol) in CH₂Cl₂ (0.8 mL) at 0°C. After stirring for 20 min, the reaction was stopped by adding saturated aqueous NaHCO₃ and then the mixture was extracted with EtOAc (\times 3). The combined organic extracts were washed with 2 M aq. HCl and brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (hexane/ EtOAc=4:6) to afford trifluoroacetoamide β -76 as a white solid (79.9 mg, 90%). Mixture of two conformers: diaxial/diequatorial = 57:43); m.p. 110–113 °C (Et₂O and hexane); $[\alpha]_D^{30} = +32.2$ (c=1.13 in CHCl₃); ¹H NMR (500 MHz, CDCl₃, the minor conformer is indicated by an asterisk): $\delta = 1.09$ (d, J = 7.2 Hz, 3H), 1.25^* (d, J = 6.4 Hz, 3H), 1.27(d, J=6.4 Hz, 3H), 1.35* (d, J=7.2 Hz, 3H), 1.45 (s, 3H), 1.65* (s, 3H), 1.88 (s, 3H), 1.95* (s, 3H), 1.96 (s, 3H), 2.00* (s, 3H), 2.36 (s, 3H), 2.43* (s, 3H), 3.19 (dd, J=7.9, 12.0 Hz, 1H), 3.25 (s, 2H), 3.32* (dd, J=8.1, 12.0 Hz, 1H), 3.57* (s, 3H), 3.66* (s, 3H), 3.71 (s, 3H), 3.78* (s, 3H), 3.80-3.84 (m, 1H), 3.82 (s, 3H), 3.85 (s, 3H), 3.90* (s, 3H), 3.93 (dd, J= 5.0, 12.0 Hz, 1 H), 3.93–3.98* (m, 1 H), 4.03* (dd, J = 5.3, 12.0 Hz, 1 H), 4.04 (dd, J=7.3, 10.3 Hz, 1H), 4.05* (dd, J=1.5, 10.7 Hz, 1H), 4.06 (d, J=11.3 Hz, 1H), 4.24 (d, J=11.4 Hz, 1H), 4.26* (dd, J=7.2, 10.3 Hz, 1H), 4.35 (d, J=6.9 Hz, 1H), 4.403* (d, J=12.4 Hz, 1H), 4.405 (s, 2H), 4.457* (d, J = 10.8 Hz, 1 H), 4.460* (d, J = 12.3 Hz, 1 H), 4.47 (d, J = 12.3 Hz, 11.3 Hz, 1 H), 4.48* (d, J=12.4 Hz, 1 H), 4.57* (d, J=10.7 Hz, 1 H), 4.561 (d, J=8.0 Hz, 1H), 4.562* (d, J=6.8 Hz, 1H), 4.60 (qd, J=7.2, 7.8 Hz, 1H), 4.63 (dd, J=6.9, 8.8 Hz, 1H), 4.64* (d, J=10.8 Hz, 1H), 4.65 (d, J= 3.1 Hz, 1 H), 4.67* (qd, J=7.2, 7.3 Hz, 1 H), 4.69* (d, J=12.3 Hz, 1 H), 4.79 (ddd, J=5.0, 7.9, 8.5 Hz, 1 H), 4.80* (dd, J=6.8, 8.4 Hz, 1 H), 4.83 (dd, J = 8.5, 8.8 Hz, 1 H), 4.87 (d, J = 11.4 Hz, 1 H), 4.89* (ddd, J = 5.3, 8.1, 1 H)8.4 Hz, 1 H), 4.96* (dd, J=8.4, 8.4 Hz, 1 H), 4.99 (dd, J=3.1, 7.3 Hz, 1 H), 5.01 (d, J=3.1 Hz, 1H), 5.11* (dd, J=7.2, 10.3 Hz, 1H), 5.24* (d, J=8.0 Hz, 1 H), 5.29 (dd, J=8.0, 10.3 Hz, 1 H), 5.67 (d, J=7.8 Hz, 1 H; NH), 5.88* (dd, J=8.0, 10.3 Hz, 1H), 6.29* (d, J=7.3 Hz, 1H; NH), 6.64 (s, 1H), 6.77* (s, 1H), 6.81-6.85 (m, 2H), 6.86-6.90* (m, 1H), 6.91-6.95* (m, 2H), 6.98-7.02* (m, 2H), 7.05-7.18 (m, 7H), 7.18-7.40 (m, 10H), 7.41-7.46 (m, 2H), 7.48 (s, 1H), 7.51-7.56 (m, 1H), 7.81 (d, J=1.6 Hz, 1H), 7.95-7.99 ppm (m, 2H); ¹³C NMR (125 MHz, CDCl₃, without distinction of the two conformers): $\delta = 16.0 (2 \text{ C}), 17.5, 18.1, 19.1, 19.8, 20.0,$ 20.1, 20.5, 20.57, 20.64, 20.7, 34.23, 34.24, 47.9, 48.3, 52.2, 56.2, 56.4, 56.5, 56.6, 61.1, 62.2, 62.4, 68.9, 69.1, 70.3, 70.4, 70.8, 71.0, 71.3, 71.4, 71.5, 72.4, 74.31, 74.34, 74.5, 76.0, 76.4, 76.5, 76.6, 77.1, 77.2, 77.6, 81.4, 98.6, 100.8, 101.4, 101.8, 102.1, 107.4, 108.0, 112.4, 116.7 (q, ${}^{1}J_{CF}=286$ Hz), 116.8 (q, ¹*J*_{CF}=286 Hz), 119.0, 119.6, 120.8, 121.1, 122.8, 122.9, 123.4, 123.5, 127.1, 127.2, 127.4, 127.5, 127.6, 127.7, 127.8, 127.9, 128.01, 128.03, 128.2, 128.4, 128.5, 128.7, 128.8, 129.3, 129.7, 130.30, 130.32, 131.2, 132.5, 132.8, 133.0, 133.6, 136.3, 136.7, 136.8, 137.3, 137.4, 137.8, 137.8, 138.0, 138.7, 138.8, 145.2, 145.5, 152.4, 153.0, 153.2, 153.3, 154.35, 154.39, 158.76 (q, ${}^{2}J_{CF} =$ 35 Hz), 158.83 (q, ${}^{2}J_{C,F}$ =35 Hz), 164.0, 164.9, 167.30, 167.33, 168.9, 169.1, 169.56, 169.59, 169.9, 170.0, 172.9, 173.1 ppm; IR (KBr): $\tilde{\nu} = 3410$, 2940, 1750, 1700, 1660, 1580, 1510, 1460, 1370, 1340, 1250, 1220, 1150, 1070,

FULL PAPER

900, 820, 750, 700 cm⁻¹; elemental analysis calcd (%) for C₇₄H₇₄ClF₃N₂O₂₁: C 62.60, H 5.25, N 1.97; found: C 62.33, H 5.28, N 1.87. Anthraquinone 77: A solution of CAN (77.8 mg, 0.142 mmol) in water (0.18 mL) was added to a solution of β -76 (96.0 mg, 0.0676 mmol) in CH₃CN (0.53 mL) at 0°C. After stirring for 20 min at RT, the reaction was stopped by adding water. The precipitates were collected by filtration to afford the chloroquinone (71.3 mg) as a red powder. The filtrate was extracted with EtOAc (×3) and the combined organic extracts were washed with brine, dried (Na2SO4), and concentrated in vacuo. A solution of diene 21 (150 mg, 0.744 mmol) in THF (1.4 mL) was added to the combined crude material (99.6 mg) at RT and stirred for 20 min. Acidic silica gel (pH 6, Kanto chemical) was added and the organic solvent was removed in vacuo. After standing for 14 h, the silica gel was placed on a glass filter (washed with CHCl₃/MeOH=9:1) and then the filtrate was concentrated in vacuo. K₂CO₃ (251 mg) was added to the solution of this crude material in THF/CH2Cl2 (1:3, 6.8 mL) at RT. After stirring for 3 h at 40 °C, the reaction was stopped by adding 2 M aq. HCl and then the mixture was extracted with CH2Cl2 (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (hexane/Et₂O/CH₃CN=2:3:1) to afford anthraquinone 77 as a yellow solid (73.9 mg, 75%). Mixture of two conformers: diaxial/diequatorial=75:25; m.p. 123-126°C (Et₂O and hexane); $[a]_{D}^{30} = +110$ (c=1.10 in CHCl₃); ¹H NMR (500 MHz, CDCl₃, the minor conformer is indicated by an asterisk): $\delta = 1.12$ (d, J = 7.3 Hz, 3H), 1.25* (d, J=6.4 Hz, 3H), 1.35 (d, J=6.4 Hz, 3H), 1.366* (d, J= 7.2 Hz, 3 H), 1.371 (s, 3 H), 1.64* (s, 3 H), 1.85 (s, 3 H), 1.95* (s, 3 H), 1.96 (s, 3H), 2.00* (s, 3H), 2.38 (s, 3H), 2.47* (s, 3H), 3.22 (dd, J=8.0, 12.2 Hz, 1 H), 3.27 (s, 3 H), 3.32* (dd, J=8.1, 12.2 Hz, 1 H), 3.56* (s, 3 H), 3.69* (s, 3H), 3.75 (s, 3H), 3.88-3.93* (m, 1H), 3.92-3.96 (m, 1H), 3.938 (dd, J = 5.0, 12.1 Hz, 1H), 3.946* (dd, J = 0.6, 12.4 Hz, 1H), 3.949* (s, 3H), 4.00 (s, 3H), 4.03* (dd, J=4.9, 12.2 Hz, 1H), 4.09 (dd, J=7.1, 10.3 Hz, 1 H), 4.20 (d, J=11.1 Hz, 1 H), 4.24* (dd, J=7.2, 10.3 Hz, 1 H), 4.29* (d, J=11.2 Hz, 1 H), 4.34 (d, J=11.5 Hz, 1 H), 4.38 (d, J=6.8 Hz, 1 H), 4.42 (d, J=12.1 Hz, 1 H), 4.43* (d, J=11.7 Hz, 1 H), 4.46 (d, J= 12.1 Hz, 1 H), 4.465 (d, J=11.5 Hz, 1 H), 4.471* (d, J=12.1 Hz, 1 H), 4.54* (d, J=12.4 Hz, 1 H), 4.55 (d, J=7.9 Hz, 1 H), 4.57 (d, J=3.3 Hz, 1H), 4.54–4.59 (m, 1H), 4.61 (dd, J=6.8, 8.8 Hz, 1H), 4.61–4.67* (m, 1H), 4.69* (d, J=12.1 Hz, 1H), 4.76-4.81 (m, 1H), 4.78-4.82* (m, 3H), 4.82 (dd, J=8.8, 8.8 Hz, 1H), 4.89* (ddd, J=4.9, 8.1, 8.2 Hz, 1H), 4.95 (d, J=3.3 Hz, 1H), 4.96^* (dd, J=8.2, 9.2 Hz, 1H), 5.04 (dd, J=3.2, 7.1 Hz, 1 H), 5.11 (d, J=11.1 Hz, 1 H), 5.10-5.14* (m, 1 H), 5.17* (d, J= 11.2 Hz, 1 H), 5.21* (d, J=7.9 Hz, 1 H) 5.30* (dd, J=7.9, 10.3 Hz, 1 H), 5.52* (d, J=7.9 Hz, 1 H; NH), 5.87 (dd, J=7.9, 10.3 Hz, 1 H), 6.29 (d, J= 7.2 Hz, 1H; NH), 6.70* (d, J=2.5 Hz, 1H), 6.72 (d, J=2.5 Hz, 1H), 6.72-6.81 (m, 1H), 6.98-7.18 (m, 10H), 7.20-7.25 (m, 2H), 7.25-7.40 (m, 7H), 7.53-7.62 (m, 1H), 7.62 (s, 1H), 7.96-7.98 (m, 1H), 8.08* (d, J=0.6 Hz, 1 H), 12.6 (s, 1 H; OH), 12.8 ppm (s, 1 H; OH); $^{13}\!C\,NMR$ (125 MHz, CDCl₃, without distinction of the two conformers): $\delta = 16.0$, 16.1, 17.5, 18.0, 19.2, 19.7, 20.0, 20.4, 20.5, 20.57, 20.63, 20.7, 29.7, 34.2, 47.9, 48.4, 52.26, 52.31, 56.06, 56.11, 56.2, 56.6, 62.2, 62.4, 68.9, 69.1, 70.2, 70.4, 71.0, 71.1, 71.16, 71.17, 71.2, 71.4, 71.5, 72.3, 73.4, 74.0, 74.6, 76.4, 76.8, 76.9 (2C), 77.2, 77.3, 80.4, 98.7, 101.3, 101.9, 102.1, 105.8, 106.0, 107.1, 107.4, 110.4, 110.7, 116.6 (q, ${}^{1}J_{C,F}$ =286 Hz), 118.3, 120.5, 121.7, 122.5, 123.2, $125.2,\ 126.2,\ 127.3,\ 127.4,\ 127.5,\ 127.6,\ 127.68,\ 127.72,\ 127.9,\ 128.05,$ 128.12, 128.2, 128.3, 128.4, 128.45, 128.48, 128.56, 128.64, 128.8, 129.0, 129.1, 129.7, 130.3, 132.6, 132.8, 133.1, 133.5, 133.7, 134.6, 135.1, 136.0, 136.3, 136.4, 136.5, 136.8, 137.1, 137.2, 138.0, 138.1, 139.2, 139.8, 140.0, 144.0, 153.1, 154.9, 156.5, 158.4, 158.8 (q, ${}^{2}J_{C,F}=35$ Hz), 163.7, 164.6, 164.9, 166.6, 166.9, 168.9, 169.1, 169.55, 169.59, 169.9, 170.0, 172.86, 172.90, 180.5, 181.1 ppm; IR (KBr): $\tilde{\nu} = 3410$, 2960, 1750, 1700, 1670, 1630, 1610, 1580, 1540, 1500, 1450, 1380, 1320, 1260, 1220, 1150, 1070, 1030, 900 870, 800, 740, 700 cm $^{-1}$; elemental analysis calcd (%) for $C_{77}H_{73}F_3N_2O_{23}{:}\ C$ 63.72, H 5.07, N 1.93; found: C 64.00, H 5.21, N 1.76. Pradimicin A hydrochloride (1 a·HCl): 5% Pd/C (27.8 mg) was added to a solution of 77 (21.0 mg, 0.0145 mmol) in MeOH (1.6 mL) and DMF (0.5 mL). After stirring under H₂ (1 atm) at RT for 30 min, the reaction mixture was filtered though a Hyflo Super-Cel pad (washed with CHCl₃/ MeOH=9:1) and concentrated in vacuo. The residue was purified by PTLC (CHCl₃/MeOH=9:1) to afford the corresponding tetrol (13.5 mg)

Chem. Eur. J. 2007, 13, 9791-9823

© 2007 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim

as a red amorphous solid. A 5 M aq. NaOH solution (0.07 mL) was added to the solution of this crude material in H₂O (0.71 mL) at 0°C. After stirring for 24 h at RT, the reaction was stopped by adding 2M aq. HCl (pH 3.5) and the mixture concentrated in vacuo. The residue was purified by reversed-phase silica gel column chromatography (Cosmosil: CH₃CN/ $H_2O = 3.7$, pH 3) to give red precipitates that were further purified by Sephadex LH-20 (DMF) to afford pradimicin A (1a) dimethylformamide solvate, which was dissolved in MeOH and then 2M aq. HCl was added until a pH value of 3.5 was obtained. After removal of the solvent in vacuo, the residue was dissolved in DMSO (25 µL) and CHCl3 was added. The precipitates were collected by filtration to afford 1a·HCl as a red amorphous powder (4.3 mg, 61%). M.p. 194-197 °C (decomp) (lit.^[1d] 193–195°C (decomp)); $[\alpha]_{D}^{26} = +280$ (c=0.100 in 0.1 M aq. HCl) (authentic: +287 (c=0.100 in 0.1 M aq. HCl)); ¹H NMR (500 MHz, [D₆]DMSO, 313 K): $\delta = 1.25$ (d, J = 6.2 Hz, 3H), 1.35 (d, J = 7.0 Hz, 3H), 2.29 (s, 3H), 2.61 (s, 3 H), 3.74 (dd, J=5.3, 11.2 Hz, 1 H), 3.84 (q, J=6.2 Hz, 1 H), 4.39 (q, J=7.0 Hz, 1 H), 4.44 (d, J=7.3 Hz, 1 H), 4.46 (d, J=10.3 Hz, 1 H),4.49 (d, J=10.3 Hz, 1H), 4.73 (d, J=7.7 Hz, 1H), 6.71 (d, J=2.4 Hz, 1H), 6.85 (s, 1H), 7.11 (d, J=2.4 Hz, 1H), 7.69 (s, 1H), 8.78 ppm (d, J= 7.0 Hz, 1 H); $^{13}{\rm C}$ NMR (125 MHz, [D₆]DMSO, 313 K): $\delta\!=\!16.6,\,17.5,\,19.7,$ 36.8, 48.2, 57.0, 63.8, 66.4, 68.0, 70.0, 70.6, 74.1, 76.5, 79.7, 81.5, 104.6, 105.3, 107.5, 108.2, 110.7, 114.4, 116.2, 126.4, 128.2, 131.9, 134.9, 137.9, 138.3, 148.5, 151.7, 157.5, 165.3, 166.6, 167.4, 174.5, 185.6, 188.1 ppm; IR (KBr): $\tilde{v} = 3380, 2980, 2880, 1720, 1610, 1450, 1390, 1330, 1300, 1260,$ 1200, 1160, 1060, 970, 910, 880, 790, 750, 660, 650 cm⁻¹; LRMS (MALDI-TOF, DHBA matrix): m/z: calcd for $C_{40}H_{44}O_{18}N_2Na$ ([*M*+Na]⁺): 863.2; found: 863.1; HRMS (FAB): m/z: calcd for $C_{40}H_{45}O_{18}N_2$ ([*M*+H]⁺): 841.2667; found: 841.2679.

Glycoside **78**: MS4A (537 mg) was placed in a two-necked round-bottomed flask and flame dried. The promoter was prepared in situ by stirring a mixture of Cp₂HfCl₂ (104 mg, 0.274 mmol) and AgOTf (141 mg, 0.548 mmol) in CH₂Cl₂ (1.9 mL) for 10 min at RT. A solution of **68** (200 mg, 0.249 mmol) and **80** (189 mg, 0.299 mmol) in CH₂Cl₂ (4.0 mL) was added to this suspension at -78 °C. After stirring for 17 h at -30 °C, the reaction was stopped by adding saturated aqueous NaHCO₃. The mixture was filtered through a Celite pad and extracted with EtOAc (× 3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (hexane/Et₂O/CH₃CN = 2:3:1) to afford β-glycoside **78** as a white amorphous (261 mg, 74%), along with some mixed fractions including α-glycoside **78**. Further purification by PTLC (hexane/EtOAc = 1:1) gave pure α-glycoside **78** (26.2 mg, 7.4%) as a white amorphous solid.

 β -78 (a mixture of two conformers: diaxial/diequatorial = 54:46): M.p. 118–119°C (Et₂O and hexane); $[\alpha]_D^{24} = +54.0$ (c=1.00 in CHCl₃); ¹H NMR (500 MHz, CDCl₃, the minor conformer is indicated by an asterisk): $\delta = 1.10$ (d, J = 7.2 Hz, 3H), 1.25^{2*} (d, J = 6.4 Hz, 3H), 1.29 (d, J =6.4 Hz, 3H), 1.340 (s, 3H), 1.341* (d, J=7.2 Hz, 3H), 1.51* (s, 3H), 1.65 (s, 3H), 1.76* (s, 3H), 1.97 (s, 3H), 2.02* (s, 3H), 2.35 (s, 3H), 2.47* (s, 3H), 3.31* (dd, J=6.5, 12.4 Hz, 1H), 3.44* (dd, J=6.3, 12.4 Hz, 3H), 3.65* (s, 3H), 3.71 (s, 3H), 3.78-3.84 (m, 1H), 3.81* (s, 3H), 3.82 (s, 3H), 3.89* (s, 3H), 3.92* (dq, J=1.0, 6.4 Hz, 1H), 3.98 (dd, J=3.7, 10.3 Hz, 1 H), 4.03 (dd, J = 4.3, 12.4 Hz, 1 H), 4.10* (dd, J = 1.6, 10.7 Hz, 1 H), 4.13 (d, J=11.3 Hz, 1 H), 4.16* (dd, J=3.6, 10.1 Hz, 1 H), 4.17* (dd, J=4.4, 10.2 Hz, 1 H), 4.25^* (d, J=11.3 Hz, 1 H), 4.41 (s, 2 H), 4.42^* (d, J=11.9 Hz, 1H), 4.47 (d, J = 11.8 Hz, 1H), 4.49* (d, J = 11.9 Hz, 1H), 4.50 (d, J=11.3 Hz, 1 H), 4.5098 (dd, J=7.4, 7.9 Hz, 1 H), 4.510* (d, J=12.4 Hz, 1 H), 4.60 (qd, J=7.2, 8.4 Hz, 1 H), 4.64* (d, J=10.7 Hz, 1 H), 4.650 (d, J=11.8 Hz, 1 H), 4.652* (qd, J=7.2, 7.2 Hz, 1 H), 4.67-4.70* (m, 1H), 4.68* (d, J = 7.9 Hz, 1H), 4.69 (d, J = 7.9 Hz, 1H), 4.70–4.73 (m, 1H), 4.71 (d, J=3.3 Hz, 1H), 4.71-4.74* (m, 1H), 4.75* (d, J=12.4 Hz, 1 H), 4.76 (dd, J=7.1, 7.4 Hz, 1 H), 4.81–4.84* (m, 1 H), 4.87* (d, J=11.3 Hz, 1H), 4.90* (dd, J=7.2, 7.6 Hz, 1H), 5.08 (d, J=3.3 Hz, 1H), 5.29* (dd, J=7.9, 10.2 Hz, 1 H), 5.30 (d, J=7.9 Hz, 1 H), 5.51 (dd, J=1.1, 3.7 Hz, 1H), 5.62* (dd, J = 1.0, 4.4 Hz, 1H), 5.63 (d, J = 8.4 Hz, 1H), 5.87 (dd, J=7.9, 10.2 Hz, 1 H), 6.27* (d, J=7.2 Hz, 1 H), 6.65 (s, 1 H), 6.77* (s, 1H), 6.82-6.91 (m, 4H), 6.94-7.03 (m, 6H), 7.05-7.46 (m, 12H), 7.48-7.58 (m, 3H), 7.63–7.67* (m, 1H), 7.86* (d, J=1.6 Hz, 1H), 7.97–8.01* (m, 1H), 8.01–8.05 (m, 1H), 8.21–8.25 ppm* (m, 2H); ¹³C NMR (125 MHz, CDCl₃, without distinction of the two conformers): $\delta = 16.38$,

16.42, 17.5, 18.1, 19.0, 19.6, 19.8, 20.0, 20.2, 20.4, 20.7, 20.8, 47.9, 48.4, 52.2, 56.4, 56.5, 61.08, 61.10, 61.4, 61.5, 68.7, 68.8, 69.5, 69.7, 69.9 (2C), 70.1, 70.18, 70.22, 71.4, 72.2, 72.8, 73.2, 74.37, 74.43, 75.4, 76.0, 76.47, 76.50, 77.0 (overlapped by a signal of CDCl₃), 77.1, 77.3, 77.6, 78.0, 81.3, 98.1, 100.9, 101.0, 101.4, 107.3, 107.9, 112.5, 119.1, 119.6, 120.9, 121.1, 122.4, 122.97, 123.04, 123.29, 123.31, 127.0, 127.1, 127.25, 127.31, 127.4, 127.55, 127.64, 127.65, 127.72, 127.81, 127.82, 127.9, 127.98, 128.00, 128.1, 128.3, 128.40, 128.44, 128.5, 128.57, 128.62, 128.7, 128.8, 129.5, 129.6, 129.7, 129.8, 130.07, 130.13, 130.3, 131.2, 132.4, 132.7, 133.0, 133.2, 133.3, 133.4, 136.5, 136.8, 136.9, 137.3, 137.4, 137.7, 137.87, 137.91, 138.2, 138.8, 139.1, 145.2, 145.5, 152.4, 152.8, 153.26, 153.33, 154.3, 154.4, 164.0, 164.9, 166.15, 166.24, 167.39, 167.41, 168.5, 168.7, 169.6, 169.69, 169.72, 169.8, 172.9, 173.1 ppm; IR (KBr): v=2940, 1730, 1660, 1590, 1570, 1560, 1510, 1460, 1370, 1340, 1270, 1220, 1160, 1110, 1100, 1070, 1030, 910, 810, 740, 710 cm⁻¹; elemental analysis calcd (%) for C₇₈H₇₆ClNO₂₂: C 66.21, H 5.41, N 0.99; found: C 65.99, H 5.43, N 0.78.

 α -78 (a mixture of two conformers: diaxial/diequatorial=82:18): M.p. 105–107 °C (Et₂O and hexane); $[\alpha]_{D}^{23} = +143$ (c=0.990 in CHCl₃); ¹H NMR (500 MHz, CDCl₃, the minor conformer is indicated by an asterisk): $\delta = 0.78^*$ (d, J = 6.6 Hz, 3H), 1.06 (d, J = 7.2 Hz, 3H), 1.25 (s, 3H), 1.26 (d, J=6.6 Hz, 3H), 1.36* (d, J=7.2 Hz, 3H), 1.55* (s, 3H), 1.657 (s, 3H), 1.662* (s, 3H), 1.69* (s, 3H), 1.99 (s, 3H), 2.07* (s, 3H), 2.31 (s, 3H), 3.33 (dd, J=6.2, 12.5 Hz, 1H), 3.62* (dd, J=5.7, 12.8 Hz, 1H), 3.63* (s, 3H), 3.68 (s, 3H), 3.697 (s, 3H), 3.698 (d, J=11.2 Hz, 1H), 3.93 (d, J=11.2 Hz, 1 H), 3.95^* (s, 3 H), 4.00^* (s, 3 H), 4.05 (dd, J=4.1, 12.5 Hz, 1 H), 4.068 (s, 3 H), 4.070 (br q, J = 6.6 Hz, 1 H), 4.17* (dd, J =0.9, 11.2 Hz, 1 H), 4.19 (dd, J=3.5, 10.3 Hz, 1 H), 4.27* (d, J=11.3 Hz, 1 H), 4.28* (dd, J=4.5, 12.8 Hz, 1 H), 4.30* (d, J=11.2 Hz, 1 H), 4.40 (d, J=11.0 Hz, 1 H), 4.46* (d, J=11.4 Hz, 1 H), 4.48 (d, J=12.3 Hz, 1 H), 4.493 (d, J = 11.0 Hz, 1 H), 4.50 (dd, J = 5.5, 7.5 Hz, 1 H), 4.54 (d, J =5.5 Hz, 1 H), 4.55 (d, J=12.3 Hz, 1 H), 4.55 (d, J=12.3 Hz, 1 H), 4.57-4.63* (m, 2H), 4.60 (qd, J=7.2, 7.9 Hz, 1H), 4.69-4.74* (m, 3H), 4.70-4.75 (m, 1H), 4.78 (dd, J=7.5, 7.5 Hz, 1H), 4.82 (s, 2H), 4.83-4.87* (m, H), 4.90* (d, J=11.1 Hz, 1 H), 4.93* (d, J=11.3 Hz, 1 H), 4.96* (dd, J= 6.7, 6.7 Hz, 1 H), 5.01* (d, J=4.9 Hz, 1 H), 5.05* (d, J=11.1 Hz, 1 H), 5.22 (d, J=4.0 Hz, 1 H), 5.33* (dd, J=1.0, 3.0 Hz, 1 H), 5.34 (dd, J=4.0, 10.3 Hz, 1 H), 5.36* (d, J=3.6 Hz, 1 H), 5.54 (dd, J=1.0, 4.6 Hz, 1 H), 5.58 (d, J = 7.9 Hz, 1H), 5.74* (dd, J = 3.6, 10.7 Hz, 1H), 6.34* (d, J =7.1 Hz, 1 H), 6.68 (s, 1 H), 6.74* (s, 1 H), 6.83* (s, 1 H), 6.87-7.00 (m, 3 H), 7.02-7.08 (m, 1H), 7.09-7.14 (m, 2H), 7.16-7.36 (m, 8H), 7.42-7.65 (m, 5H), 7.84 (s, 1H), 8.04* (d, J=1.0 Hz, 1H), 8.04-8.15 (m, 2H), 8.20-8.23 ppm* (m, 2H); ¹³C NMR (125 MHz, CDCl₃, without distinction of the two conformers): $\delta = 15.6$, 16.4, 17.5, 18.1, 18.8, 19.1, 19.6, 20.0, 20.25, 20.32, 20.8, 20.9, 47.9, 48.5, 52.20, 52.24, 56.2, 56.5, 61.1, 61.2, 61.3, 61.4, 65.7, 65.8, 68.6, 68.8, 69.4, 69.5, 69.7, 70.09 (2 C), 70.14, 71.0, 73.3, 73.46, 73.49, 73.9, 75.0, 75.2, 75.8, 75.9, 76.3, 76.7, 77.2, 77.3 (overlapped by a signal of CDCl₃), 80.1, 93.4, 96.1, 100.9, 101.0, 107.3, 107.9, 112.1, 119.4, 119.6, 119.9, 120.7, 122.0, 122.3, 122.8, 122.9, 123.6, 123.7, 126.8, 127.0, 127.2, 127.4, 127.6, 127.7, 127.8, 127.9, 128.0, 128.1, 128.2, 128.4, 128.46, 128.47, 128.52, 128.6, 128.8, 128.9, 129.0, 129.16, 129.24, 129.6, 129.77, 129.80, 130.0, 130.09, 130.13, 130.4, 130.5, 131.4, 132.9, 133.0, 133.18, 133.23, 133.3, 133.9, 134.0, 134.3, 137.1, 137.2, 137.3, 137.79, 137.80, 137.82, 137.83, 138.7, 138.9, 145.4, 145.5, 152.3, 153.0 (2C), 153.4, 153.8, 154.7, 165.0, 166.0, 166.11, 166.14, 167.2, 167.3, 168.5, 168.6, 169.692, 169.694, 169.73, 169.8, 172.8, 173.0 ppm; IR (neat): $\tilde{\nu} = 3400$, 2930, 1760, 1740, 1720, 1660, 1600, 1580, 1570, 1550, 1500, 1450, 1360, 1340, 1260, 1240, 1220, 1160, 1100, 1070, 1050, 900, 810, 720, 710 cm⁻¹; elemental analysis calcd (%) for C78H76CINO22: C 66.21, H 5.41, N 0.99; found: C 65.97, H 5.18, N 0.69.

Anthraquinone **79**: A solution of CAN (85.9 mg, 0.157 mmol) in water (0.19 mL) was added to a solution of **78** (106 mg, 0.0746 mmol) in CH₃CN (0.57 mL) at 0°C. After stirring for 20 min at RT, a further solution of CAN (22.5 mg, 0.0411 mmol) in water (0.05 mL) was added and the reaction mixture was stirred for a further 10 min. The reaction was stopped by adding water and then the precipitates were collected by filtration to afford the chloroquinone as a red powder (79.7 mg). The filtrate was extracted with EtOAc (×3) and the combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. A solution of diene **21** (166 mg, 0.821 mmol) in THF (1.5 mL) was added to

9820 -

a solution of the combined crude material (110 mg) at RT. After stirring for 30 min, the reaction was stopped by adding acidic silica gel and concentrated in vacuo. After standing for 14 h, the silica gel was placed on a glass filter, washed with EtOAc, and then the filtrate was concentrated in vacuo. K₂CO₃ (276 mg) was added to the solution of this crude material in CH_2Cl_2 (5.7 mL) and THF (1.9 mL) at RT. After stirring for 2 h at 40°C, the reaction was stopped by adding 2M aq. HCl and then the mixture was extracted with EtOAc (×3). The combined organic extracts were washed with brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by PTLC (hexane/Et₂O/CH₃CN=40:45:15 and CHCl₃/EtOAc=8:2) to afford 79 as a yellow solid (71.7 mg, 66%). Mixture of two conformers: diaxial/diequatorial=75:25; m.p. 124-125°C (Et₂O and hexane); $[\alpha]_D^{26} = +137$ (c=0.985 in CHCl₃); ¹H NMR (500 MHz, CDCl₃, the minor conformer is indicated by an asterisk): $\delta =$ 1.13 (d, J=7.2 Hz, 3H), 1.255* (d, J=7.3 Hz, 3H), 1.259 (s, 3H), 1.35 (d, J=7.3 Hz, 3 H), 1.361* (d, J=7.2 Hz, 3 H), 1.51* (s, 3 H), 1.63 (s, 3 H), 1.77* (s, 3H), 1.97 (s, 3H), 2.02* (s, 3H), 2.35 (s, 3H), 2.49* (s, 3H), 3.33 (dd, J=6.3, 12.4 Hz, 1 H), 3.44* (dd, J=6.3, 12.3 Hz, 1 H), 3.68* (s, 3 H), 3.75 (s, 3H), 3.88-3.93 (m, 1H), 3.89-3.94* (m, 1H), 3.94* (s, 3H), 3.97* (dd, J=1.0, 11.0 Hz, 1 H), 4.00 (s, 3 H), 4.02 (dd, J=3.6, 10.1 Hz, 1 H),4.05 (dd, J=4.3, 12.4 Hz, 1 H), 4.14* (dd, J=3.5, 10.1 Hz, 1 H), 4.16* (dd, J=4.3, 12.3 Hz, 1 H), 4.292* (d, J=11.1 Hz, 1 H), 4.293 (d, J=11.1 Hz, 1H), 4.34 (d, J=11.5 Hz, 1H), 4.43 (d, J=12.1 Hz, 1H), 4.44* (d, J= 11.7 Hz, 1H), 4.466 (d, J=12.1 Hz, 1H), 4.468 (dd, J=5.7, 7.5 Hz, 1H), 4.470 (d, J=11.5 Hz, 1 H), 4.52* (d, J=12.1 Hz, 1 H), 4.55 (d, J=5.7 Hz, 1H), 4.58 (qd, J=7.2, 7.9 Hz, 1H), 4.605* (d, J=11.0 Hz, 1H), 4.607 (d, J = 3.0 Hz, 1H), 4.64* (dq, J = 7.2, 7.3 Hz, 1H), 4.67 (d, J = 8.0 Hz, 1H), 4.66-4.76* (m, 3H), 4.68-4.72 (m, 1H), 4.75 (dd, J=7.5, 7.5 Hz, 1H), 4.79* (d, J=11.7 Hz, 1 H), 4.83* (ddd, J=4.3, 7.4, 12.3 Hz, 1 H), 4.90* (dd, J=7.4, 7.4 Hz, 1 H), 5.00 (d, J=3.0 Hz, 1 H), 5.17* (d, J=11.1 Hz, 1H), 5.20 (d, J=11.1 Hz, 1H), 5.26* (d, J=8.0 Hz, 1H), 5.28 (dd, J=8.0, 10.1 Hz, 1 H), 5.46 (d, J=7.9 Hz, 1 H; NH), 5.54 (d, J=3.6 Hz, 1 H), 5.62* $(d, J=3.5 \text{ Hz}, 1 \text{ H}), 5.87^* (dd, J=8.0, 10.1 \text{ Hz}, 1 \text{ H}), 6.27^* (d, J=7.2 \text{ Hz}, 1 \text{ H})$ 1H; NH), 6.69-6.87 (m, 2H), 6.98-7.17 (m, 9H), 7.18-7.43 (m, 1H), 7.43-7.69 (m, 4H), 7.56-8.02 (m, 2H), 8.12* (d, J=1.0 Hz, 1H), 8.25 (d, J=7.2 Hz, 1H), 12.7 (s, 1H; OH), 12.9 ppm* (s, 1H; OH); ¹³C NMR (125 MHz, CDCl₃, without distinction of the two conformers): $\delta = 16.40$, 16.42, 17.5, 18.0, 19.1, 19.5, 19.8, 20.2, 20.30, 20.34, 20.7, 20.8, 47.9, 48.4, 52.2, 52.3, 56.0, 56.1, 61.4, 61.5, 68.7, 68.8, 69.5, 69.7, 70.07, 70.11, 70.2, 70.3, 71.10, 71.11, 71.4, 72.1, 72.8, 73.1, 73.8, 74.4, 74.6, 76.9, 77.2, 77.26, 77.30 (overlapped by a signal of CDCl₃), 77.4, 77.5, 80.2, 98.5, 100.7, 101.1, 101.4, 105.8, 106.0, 107.1, 107.4, 110.4, 110.7, 118.3, 120.5, 121.7, 123.3, 125.2, 126.1, 127.3, 127.4, 127.5, 127.59, 127.61, 127.7, 127.8, 128.0, 128.1, 128.2, 128.31, 128.33, 128.4, 128.5, 128.6, 128.7, 128.9, 129.0, 129.5, 129.7, 129.8, 130.00, 130.1, 132.4, 132.89, 132.94, 133.2, 133.38, 133.43, 133.5, 134.5, 135.3, 136.1, 136.4, 136.5, 136.6, 136.8, 137.2, 137.3, 137.4, 138.0, 138.5, 139.1, 139.9, 140.0, 144.2, 153.0, 154.9, 156.6, 158.6, 163.7, 164.6, 164.89, 164.91, 166.1, 166.2, 166.50, 166.52, 167.0, 168.4, 168.7, 169.6, 169.7, 169.8, 172.86, 172.92, 180.6, 181.1, 185.5, 185.9 ppm; IR (neat): $\tilde{\nu}\!=\!3400,\;2940,\;1730,\;1660,\;1620,\;1600,\;1580,\;1540,\;1500,\;1450,$ 1360, 1310, 1300, 1260, 1220, 1160, 1100, 1060, 1020, 910, 880, 720, 700 cm $^{-1};$ elemental analysis calcd (%) for $C_{81}H_{75}NO_{24}:$ C 67.26, H 5.23, N 0.97; found: C 67.19, H 5.12, N 0.80.

Benanomicin A (2a): 5% Pd/C (39.7 mg) was added to a solution of 79 (30.0 mg, 0.0207 mmol) in MeOH (2.25 mL) and DMF (0.75 mL). After stirring under H₂ (1 atm) at RT for 50 min, the reaction mixture was filtered though a Hyflo Super-Cel pad (washed with CHCl₃/MeOH=9:1) and concentrated in vacuo. The residue was purified by PTLC (CHCl₃/ MeOH=95:5) to afford the corresponding tetrol (20.4 mg) as a red amorphous solid. A 5 M aq. NaOH solution (0.23 mL) was added to the solution of this crude material in H₂O (2.3 mL) at 0 °C. After stirring for 15 h at RT, the reaction was stopped by adding 2 M aq. HCl (pH 3.5) and the mixture concentrated in vacuo. The residue was purified by Diaion HP-20 (H₂O/MeOH=1:0 to 0:1, gradient elution) to give red precipitates which were further purified by Sephadex LH-20 (DMF) (twice) to afford a red amorphous solid. The product was dissolved in DMSO (40 uL) and then CHCl₃ was added. The precipitate was collected by filtration and dried in vacuo at 50°C for 2 days to afford benanomicin A as a red amorphous powder (11.6 mg, 68%). M.p. 242–244 $^{\circ}\mathrm{C}$ (decomp) (lit.: $^{[1b]}$

FULL PAPER

>220 °C); ¹H NMR (500 MHz, [D₆]DMSO, 313 K): $\delta = 1.03$ (d, J =6.3 Hz, 3 H), 1.36 (d, J=7.3 Hz, 3 H), 2.32 (s, 3 H), 2.91-3.38 (m, 6 H), 3.27 (s, 3H), 3.60 (dd, J=5.2, 11.3 Hz, 1H), 3.96 (s, 3H), 4.06 (dd, J=7.1, 9.6 Hz, 1 H), 4.44 (d, J = 7.1 Hz, 1 H), 4.45 (q, J = 6.7 Hz, 1 H), 4.55 (d, J = 10.0 Hz, 1 H), 4.63 (d, J=10.0 Hz, 1 H), 4.72 (d, J=7.7 Hz, 1 H), 4.78 (dd, J=2.9, 7.0 Hz, 1H), 6.91 (d, J=2.5 Hz, 1H), 7.11 (br, 1H), 7.32 (d, J=2.5 Hz, 1H), 7.94 (br, 1H), 8.11 (br, 1H), 8.18 (br, 1H), 12.82 ppm (s, 1 H); ¹³C NMR (500 MHz, $[D_6]$ DMSO, 313 K): $\delta = 16.3$, 16.9, 19.1, 47.7, $56.4,\ 65.6,\ 69.5,\ 70.1,\ 70.4,\ 71.9,\ 70.4,\ 71.9,\ 73.7,\ 76.1,\ 81.6,\ 83.0,\ 104.5,$ 105.3, 106.9, 107.6, 110.1, 113.6, 115.6, 125.6, 127.5, 131.3, 134.3, 137.4, 138.1, 147.9, 151.1, 156.9, 164.7, 166.0, 166.9, 173.0, 174.0, 185.0, 187.5) ppm (signals for four sp² carbon atoms were not detected presumably owing to peak-broadening ascribed to the slow conformational interconversion, diaxial \rightarrow diequatorial; IR (KBr): $\tilde{\nu}$ =3420, 2980, 2940, 1720, 1620, 1520, 1450, 1390, 1340, 1300, 1260, 1210, 1190, 1160, 1130, 1070, 1050, 1000, 980, 900, 860, 800, 750, 660 $\rm cm^{-1};\ LRMS$ (MALDI-TOF, DHBA matrix): m/z: calcd for $C_{39}H_{41}O_{19}NNa$ ([*M*+Na]⁺): 850.2; found: 850.2; HRMS (FAB): m/z: calcd for $C_{39}H_{41}O_{19}NNa$ ([*M*+Na]⁺): 850.2170; found: 850.2162.

Acknowledgements

We are grateful to Professors Toshikazu Oki and Yasuhiro Igarashi for providing us with an authentic sample of pradimicin A. Thanks are also due to Meiji Seika Kaisha, Ltd. for providing us with authentic samples of benanomicins A and B. We are grateful to Professor Dr. Katsuhiko Tomooka and Dr. Kazunobu Igawa for in situ IR analysis. Partial financial support from the 21st Century COE Program (Tokyo Institute of Technology). M.T. is grateful to the JSPS Research Fellowship for Young Scientists. We also thank Professor Dr. Jay S. Siegel and Professor Dr. Dennis P. Curran for helpful discussions and careful proof reading.

- [1] a) T. Oki, M. Konishi, K. Tomatsu, K. Saito, M. Tsunakawa, M. Nishio, T. Miyaki, H. Kawaguchi, J. Antibiot. 1988, 41, 1701-1704; b) T. Takeuchi, T. Hara, H. Naganawa, M. Okada, M. Hamada, H. Umezawa, S. Gomi, M. Sezaki, S. Kondo, J. Antibiot. 1988, 41, 807-811; c) S. Gomi, M. Sezaki, S. Kondo, T. Hara, H. Naganawa, T. Takeuchi, J. Antibiot. 1988, 41, 1019-1028; d) M. Tsunakawa, M. Nishio, H. Ohkuma, T. Tsuno, M. Konishi, T. Naito, T. Oki, H. Kawaguchi, J. Org. Chem. 1989, 54, 2532-2536; e) Y. Sawada, M. Nishio, H. Yamamoto, M. Hatori, T. Miyaki, M. Konishi, T. Oki, J. Antibiot. 1990, 43, 771-777; f) Y. Sawada, M. Hatori, H. Yamamoto, M. Nishio, T. Miyaki, T. Oki, J. Antibiot. 1990, 43, 1223-1229; g) Y. Sawada, T. Tsuno, H. Yamamoto, M. Nishio, M. Konishi, T. Oki, J. Antibiot. 1990, 43, 1367-1374; h) K. Saitoh, Y. Sawada, K. Tomita, T. Tsuno, M. Hatori, T. Oki, J. Antibiot. 1993, 46, 387-397; i) K. Saitoh, K. Suzuki, M. Hirano, T. Furumai, T. Oki, J. Antibiot. 1993, 46, 398-405; j) K. Saitoh, T. Tsuno, M. Kakushima, M. Hatori, T. Furumai, T. Oki, J. Antibiot. 1993, 46, 406-411; k) Y. Sawada, T. Tsuno, T. Ueki, H. Yamamoto, Y. Fukagawa, T. Oki, J. Antibiot. 1993, 46, 507-510; I) T. Furumai, T. Hasegawa, M. Kakushima, K. Suzuki, H. Yamamoto, S. Yamamoto, M. Hirano, T. Oki, J. Antibiot. 1993, 46, 589-597; m) T. Furumai, H. Yamamoto, Y. Narita, T. Hasegawa, S. Aburaki, M. Kakushima, T. Oki, J. Antibiot. 1993, 46, 1589 - 1597.
- [2] a) T. Oki, O. Tenmyo, M. Hirano, K. Tomatsu, H. Kamei, J. Antibiot. 1990, 43, 763–770; b) M. Kakushima, M. Nishio, K. Numata, M. Konishi, T. Oki, J. Antibiot. 1990, 43, 1028–1030; c) T. Oki, M. Kakushima, M. Nishio, H. Kamei, M. Hirano, Y. Sawada, M. Konishi, J. Antibiot. 1990, 43, 1230–1235; d) A. Tanabe-Tochikura, S. Tanabe-Tochikura, O. Yoshida, T. Oki, N. Yamamoto, Virology 1990, 176, 467–473; e) T. Oki, M. Kakushima, M. Hirano, M. Takahashi, A. Ohta, S. Masuyoshi, M. Hatori, H. Kamei, J. Antibiot. 1992, 45, 1512–1517; f) H. Kamachi, S. Iimura, S. Okuyama, H. Hoshi, S. Tamura, M. Shinoda, K. Saitoh, M. Konishi, T. Oki, J. Antibiot. 1992, 45, 1518–1525; g) S. Aburaki, H. Yamashita, T. Ohnuma, H. Kamachi,

T. Moriyama, S. Masuyoshi, H. Kamei, M. Konishi, T. Oki, J. Antibiot. 1993, 46, 631–640; h) T. Ueki, K. Numata, Y. Sawada, T. Nakajima, Y. Fukagawa, T. Oki, J. Antibiot. 1993, 46, 149–161; i) M. Watanabe, S. Gomi, H. Tohyama, K. Ohtsuka, S. Shibahara, S. Inouye, H. Kobayashi, S. Suzuki, S. Kondo, T. Takeuchi, H. Yamaguchi, J. Antibiot. 1996, 49, 366–373; j) T. Mizuochi, M. Nakata, Jpn. J. Clin. Med. 1995, 53, 2340–2349; k) T. Ueki, K. Numata, Y. Sawada, M. Nishio, H. Ohkuma, S. Toda, H. Kamachi, Y. Fukagawa, T. Oki, J. Antibiot. 1993, 46, 455–464; l) K. Fujikawa, Y. Tsukamoto, T. Oki, Y. C. Lee, Glycobiology 1998, 8, 407–414.

- [3] a) T. Nishizuka, S. Hirosawa, S. Kondo, D. Ikeda, T. Takeuchi, J. Antibiot. 1997, 50, 755-764; b) K. Krohn, S. Bernhard, U. Flörke, N. Hayat, J. Org. Chem. 2000, 65, 3218-3222; c) F. M. Hauser, H. Liao, Y. Sun, Org. Lett. 2002, 4, 2241-2443; d) T. R. Kelly, W. Xu, Z. Ma, Q. Li, V. Bhushan, J. Am. Chem. Soc. 1993, 115, 5843-5844; e) T. R. Kelly, Q. Li, V. Bhushan, Tetrahedron Lett. 1990, 31, 161-164; for a comprehensive review of bioactive antibiotics bearing biaryl structural motifs, see: f) G. Bringmann, C. Günther, M. Ochse, O. Schupp, S. Tasler, Progress in the Chemistry of Organic Natural Products, Vol.82 (Eds.: W.Herz, H. Falk, G. W. Kirby, R. E. Moore), Springer, Wien, 2001.
- [4] a) M. Kitamura, K. Ohmori, T. Kawase, K. Suzuki, Angew. Chem.
 1999, 111, 1308–1311; Angew. Chem. Int. Ed. 1999, 38, 1229–1232;
 b) K. Ohmori, M. Tamiya, M. Kitamura, H. Kato, M. Oorui, K. Suzuki, Angew. Chem. 2005, 117, 3939–3942; Angew. Chem. Int. Ed. 2005, 44, 3871–3874.
- [5] K. Ohmori, M. Kitamura, K. Suzuki, Angew. Chem. 1999, 111, 1304–1307; Angew. Chem. Int. Ed. 1999, 38, 1226–1229.
- [6] K. Ohmori, M. Kitamura, Y. Ishikawa, H. Kato, M. Oorui, K. Suzuki, *Tetrahedron Lett.* 2002, 43, 7023–7026.
- [7] H. Kato, K. Ohmori, K. Suzuki, Synlett 2001, 1003–1005.
- [8] P. S. Baily, R. E. Erickson, Org. Synth. 1973, Coll. Vol. V, 489-493.
- [9] For the preparation of SmI₂ in THF, see: P. Girard, J. L. Namy, H. B. Kagan, J. Am. Chem. Soc. 1980, 102, 2693–2698.
- [10] a) R. N. Armstrong, D. A. Lewis, H. L. Ammon, S. M. Prasad, J. Am. Chem. Soc. 1985, 107, 1057–1058; b) R. N. Armstrong, D. A. Lewis, J. Org. Chem. 1985, 50, 907–908; c) D. I. Cobb, D. A. Lewis, R. N. Armstrong, J. Org. Chem. 1983, 48, 4139–4141; d) D. M. Jerina, H. Selander, H. Yagi, M. C. Wells, J. F. Davey, V. Mahadevan, D. T. Gibson, J. Am. Chem. Soc. 1976, 98, 5988–5996.
- [11] The enantiomeric purity of **6** was assessed by ¹H NMR spectroscopy (400 MHz) after conversion into the (+)- and (-)-MTPA derivatives: J. A. Dale, D. L. Dull, H. S. Mosher, *J. Org. Chem.* **1969**, *34*, 2543–2549. In addition, the acetonide was used to fix the conformation for the measurement of the CD spectrum (CD (MeOH): λ_{ext} = 248.9 nm ($\Delta \varepsilon$ = +1.59×10² M⁻¹ cm⁻¹) because conformational change leads to the opposite helicity, which shows the reverse sign in the CD curve (see ref. [10]).
- [12] a) K. Mislow, B. H. Harvey, J. Am. Chem. Soc. 1962, 84, 3018–3020;
 b) R. N. Armstrong, B. Kedzierski, W. Levin, D. M. Jerina, J. Biol. Chem. 1981, 256, 4726–4733; c) D. A. Lewis, R. N. Armstrong, Biochemistry 1983, 22, 6297–6303; d) E. L. Eliel, S. H. Wilen, Stereochemistry of Organic Compounds, Wiley, New York 1994, chapter 14-5; e) H. Kato, K. Ohmori, K. Suzuki, Chirality 2000, 12, 548–550.
- [13] a) G. Bringmann, J. R. Jansen, H.-P. Rink, Angew. Chem. 1986, 98, 917–919; Angew. Chem. Int. Ed. Engl. 1986, 25, 913–915; b) P. P. Deshande, O. R. Martin, Tetrahedron Lett. 1990, 31, 6313–6318; c) T. Matsumoto, T. Hosoya, K. Suzuki, J. Am. Chem. Soc. 1992, 114, 3568–3570; d) T. Hosoya, E. Takashiro, T. Matsumoto. K. Suzuki, J. Am. Chem. Soc. 1994, 116, 1004–1015.
- [14] For enantioselective reactions, see: a) G. Bringmann, T. Hartung, Angew. Chem. 1992, 104, 782–783; Angew. Chem. Int. Ed. Engl. 1992, 31, 761–762; b) G. Bringmann, T. Hartung, Tetrahedron 1993, 49, 7891–7902; c) G. Bringmann, M. Breuning, Synlett 1998, 634– 636; for diastereoselective reactions, see: d) G. Bringmann, H. Reuscher, Angew. Chem. 1989, 101, 1725–1726; Angew. Chem. Int. Ed. Engl. 1989, 28, 1672–1673; e) G. Bringmann, M. Breuning, R. Walter, A. Wuzik, K. Peters, E.-M. Peters, J. Org. Chem. 1999, 64,

3047–3055; f) G. Bringmann, M. Breuning, S. Tasler, H. Endress, C. L. J. Ewers, L. Göbel, K. Peters, E.-M. Peters, *Chem. Eur. J.* **1999**, 5, 3029–3038; for a review, see: g) G. Bringmann, M. Breuning, S. Tasler, *Synthesis* **1999**, 525–558.

- [15] a) J. Savard, P. Brassard, *Tetrahedron* **1984**, 40, 3455–3464; b) S. Danishefsky, Acc. Chem. Res. **1981**, 14, 400–406.
- [16] a) J. Chiarello, M. M. Joullié, *Tetrahedron* **1988**, 44, 41–48; b) K. van Laak, H.-D. Scharf, *Tetrahedron* **1989**, 45, 5511–5516.
- [17] S. Cacchi, P. G. Ciattini, E. Morera, G. Ortar, *Tetrahedron Lett.* 1986, 27, 3931–3934.
- [18] S. Kajigaeshi, T. Kakinami, H. Yamasaki, S. Fujisaki, M. Kondo, T. Okamoto, *Chem. Lett.* **1987**, 2109–2112.
- [19] W. M. Owton, P. T. Gallagher, A. Juan-Montesinos, Synth. Commun. 1993, 23, 2119–2125.
- [20] J. L. Loomer, K. W. Stagliano, J. A. Gazzillo, J. Org. Chem. 1993, 58, 7906–7912.
- [21] M. F. Comber, M. V. Sargent, J. Chem. Soc., Perkin Trans. 1 1991, 2783–2787.
- [22] The regiochemistry of 22 (R=H) was assigned by HMBC analysis.
- [23] Condensation with a carbodiimide (e.g., EDCI, DCC) gave a low yield of the ester (\approx 50%) with a sizable amount of the acyl urea derived from carboxylic acid **26** and the carbodiimide (\approx 30%).
- [24] J. Inanaga, K. Hirata, H. Saeki, T. Katsuki, M. Yamaguchi, Bull. Chem. Soc. Jpn. 1979, 52, 1989–1993.
- [25] J. P. Brown, E. B. McCall, J. Chem. Soc. 1955, 3681-3687.
- [26] G. Casiraghi, F. Bigi, G. Casnati, G. Sartori, P. Soncini, G. G. Fava, M. F. Belicchi, J. Org. Chem. 1988, 53, 1779–1785.
- [27] The requisite *M* chirality of diastereomer **37a** was determined from the CD spectrum of the two enantiomers of **38** derived from **37a** and **37b**, respectively.
- [28] The enantiomeric purity of the biaryl dialdehyde **39** was assessed by HPLC analysis (DAICEL CHIRALCEL OD-H (0.46 cm $\varphi \times 25$ cm), hexane/*i*PrOH=9:1, flow rate=0.7 mLmin⁻¹, $t_{\rm R}$ =18.2 min for the *M* isomer, 26.4 min for the *P* isomer).
- [29] The enantiomeric purity of diol **40** was assessed by HPLC analysis (DAICEL CHIRALCEL OD-H (0.46 cm $\varphi \times 25$ cm), hexane/ *i*PrOH=9:1, flow rate=1.0 mLmin⁻¹, $t_{\rm R}$ =10.9 min for the *R*,*R* isomer, 15.0 min for *S*,*S* isomer).
- [30] The same conversion was applied to the racemate of 38. Sample 46, thus obtained, showed additional ¹H NMR peaks that could be assigned to the 5,6-bis-epimer (relative to the D-alanine moiety).
- [31] M. Kitamura, unpublished results: The glycosidation of 51 with quinone containing alcohol resulted in the recovery of the starting material.
- [32] a) K. Suzuki, H. Maeta, T. Matsumoto, G. Tsuchihashi, *Tetrahedron Lett.* 1988, 29, 3571–3574; b) K. Suzuki, H. Maeta, T. Matsumoto, *Tetrahedron Lett.* 1989, 30, 4853–4856.
- [33] For the preparation of glycosyl donor 51 and the glycosylation study, see: H. Kato, K. Ohmori, K. Suzuki, *Tetrahedron Lett.* 2000, 41, 6827–6832. See, also the Supporting Information.
- [34] See the Supporting Information.
- [35] All the physical data (¹H and ¹³C NMR, IR, $[\alpha]_D$, and m.p.) of the synthetic material were identical to those of benanomicin B-HCl.
- [36] K. Soai, T. Yamanoi, H. Oyamada, Chem. Lett. 1984, 251-254.
- [37] M. Yatagai, T. Ohnuki, J. Chem. Soc., Perkin Trans. 1 1990, 1826– 1828.
- [38] S. Yamada, Y. Mori, K. Morimatsu, Y. Ishizu, Y. Ozaki, R. Yoshioka, T. Nakatani, H. Seko, J. Org. Chem. 1996, 61, 8586–8590.
- [39] Acyl rearrangement was very slow in the absence of valinol.
- [40] The relative axial stereochemistry was determined by X-ray crystallographic analysis. CCDC-262629 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.
- [41] For the SmI₂/BF₃·OEt₂ reduction system, see: A. Studer, D.P. Curran, Synlett 1996, 255–257.
- [42] D. H. Eargle, Jr., R. Emrich, J. Org. Chem. 1970, 35, 3744-3747.

Chem. Eur. J. 2007, 13, 9791-9823

- [43] a) A. I. Meyers, M. Shipman, J. Org. Chem. 1991, 56, 7098-7102;
 b) S. Boisnard, L. Neuville, M. Bois-Choussy, J. Zhu, Org. Lett. 2000, 2, 2459-2462.
- [44] T. Tsunoda, M. Suzuki, R. Noyori, *Tetrahedron Lett.* 1980, 21, 1357– 1358.
- [45] The enantiomeric purity of **59** was assessed by HPLC analysis [DAICEL CHIRALPAK AD-H (0.46 cm $\varphi \times 25$ cm), hexane/ *i*PrOH=9:1, flow rate = 1.0 mLmin⁻¹, $t_{\rm R}$ =5.4 min for the *M* isomer, 8.0 min for the *P* isomer].
- [46] The enantiomeric purity of **60** was analyzed by HPLC analysis [DAICEL CHIRALPAK AD-H (0.46 cm $\varphi \times 25$ cm), hexane/ EtOH=8:2, flow rate = 1.0 mL min⁻¹, $t_{\rm R}$ =6.6 min for the *S*,*S* isomer, 9.3 min for the *R*,*R* isomer].
- [47] The relative stereochemistry of the two stereogenic centers at the B ring was determined by X-ray crystallographic analysis. CCDC-622146 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_ request/cif.
- [48] The lowest temperature for activation of the glycosyl fluoride was -36°C, and 68 underwent decomposition with an excess of Cp₂HfCl₂ (e.g., 1.2 equiv) and AgClO₄ (e.g., 2.4 equiv).
- [49] DMF was necessary for the completion of this step as the intermediates were insoluble in MeOH. When only DMF was used as the solvent, the reaction did not go to completion.
- [50] The reaction did not go to completion without the addition of 1 M aq. HCl.

- [51] For selected examples of the N-monoalkylation of primary amines, see: a) R. K. Olsen, J. Org. Chem. 1970, 35, 1912–1915; b) E. M. Briggs, G. W. Brown, J. Jiricny, M. F. Meidine, Synthesis 1980, 295–296; c) S. Krishnamurthy, Tetrahedron Lett. 1982, 23, 3315–3318; d) P. A. Grieco, A. Bahsas, J. Org. Chem. 1987, 52, 5746–5749, and references cited therein.
- [52] H. Staudinger, J. Meyer, *Helv. Chim. Acta* **1919**, *2*, 635–646; for a review of the Staudinger reaction, see: Y. G. Gololobov, L. F. Kasukhin, *Tetrahedron* **1992**, *48*, 1353–1406.
- [53] We also attempted the conventional method for the *N*-monoalkylation of primary amines, which resulted in failure: After treatment of azide **69** with Me₃P and H₂O, the resulting primary amine was protected by a trifluoroacetyl group and its *N*-methylation was attempted (NaH, MeI). The reaction only gave a complex mixture of unidentified products.
- [54] C. D. Meo, M. N. Kamat, A. V. Demchenko, Eur. J. Org. Chem. 2005, 706–711.
- [55] Removal of the trifluoroacetyl group was sluggish in the presence of MeOH.
- [56] Although the ¹H NMR spectrum of **1a** proved highly dependent on the pH, the measurement at pH 3.5 allowed us to compare each sample of synthetic/authentic **1a**.
- [57] For the preparation of glycosyl donor **80**, see the Supporting Information.

Received: June 6, 2007 Published online: September 28, 2007